Stop codons & Readthrough

U. III

A

Fidelity in protein synthesis

DNA replication and transcription are based on complementarity and correctly matched base-pairing.

During translation, each tRNA is covalently bound to an amino acid in order to be accomodated in the ribosomal A-site due to correspondence of codon (on mRNA), anticodon (on tRNA), and amino acid.

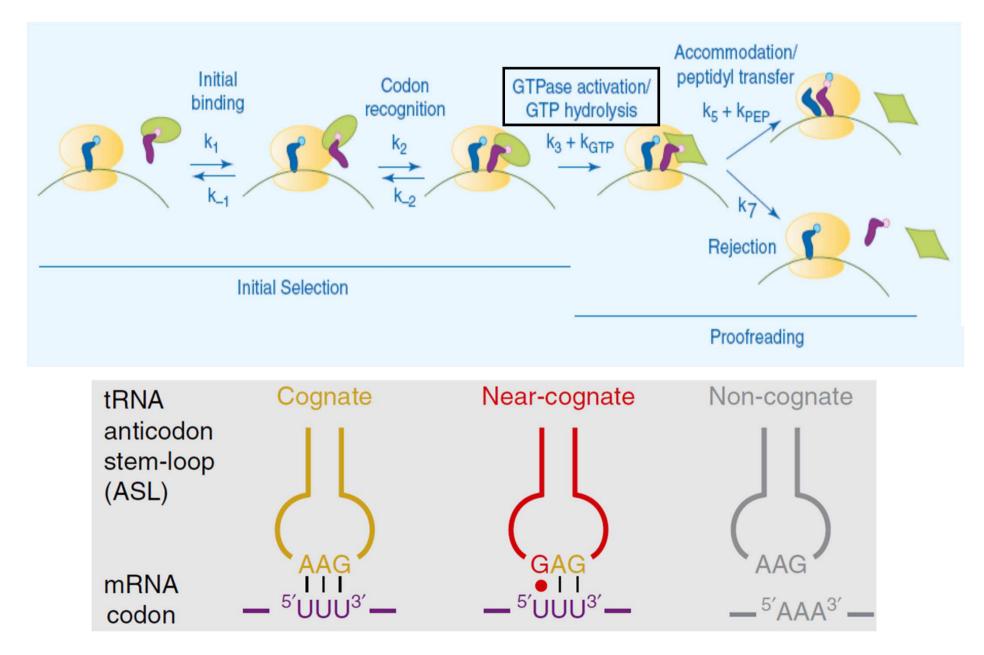
Three strategies ensure a balance between <u>velocity</u> (3-5 aa/sec) and <u>accuracy</u> (**error rate** $\sim 10^{-4}$):

1. Editing (tRNA/amino acid)------ Aa-tRNA synthetase

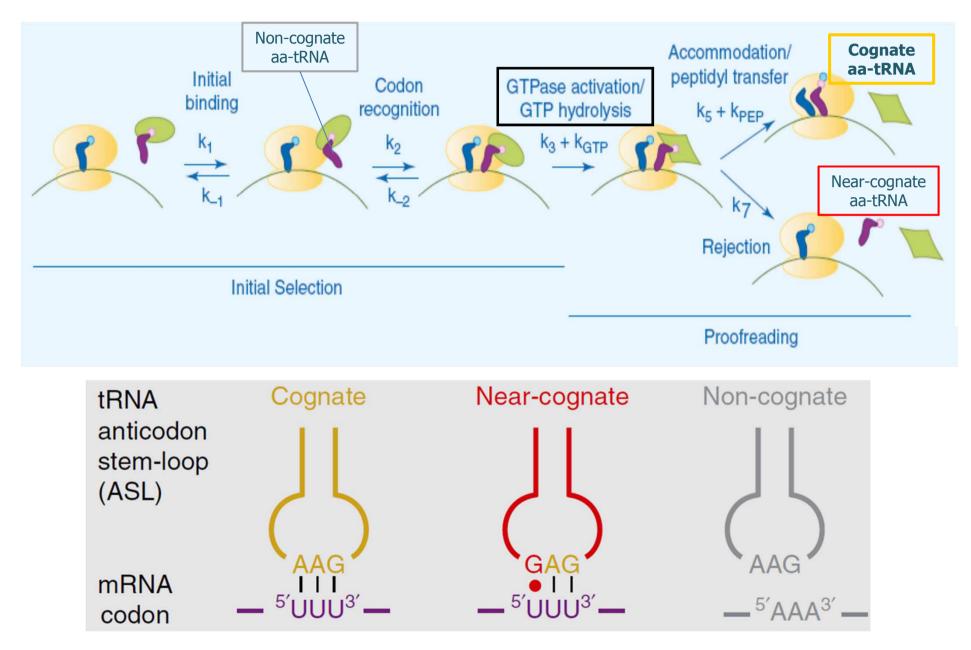
Ribosome

- 2. Kinetic proofreading (codon/anticodon)-----
- 3. Induced fit (codon/anticodon)------ⁱ

Kinetic proofreading



Kinetic proofreading



Translational reprogrammed genetic decoding (RECODING) during protein synthesis

Recoding: regulatory mechanisms of protein expression that include several non-canonical events, opposite to the DNA \rightarrow RNA \rightarrow Protein central dogma of biology

Recoding was found to be associated to elongation and termination phases:

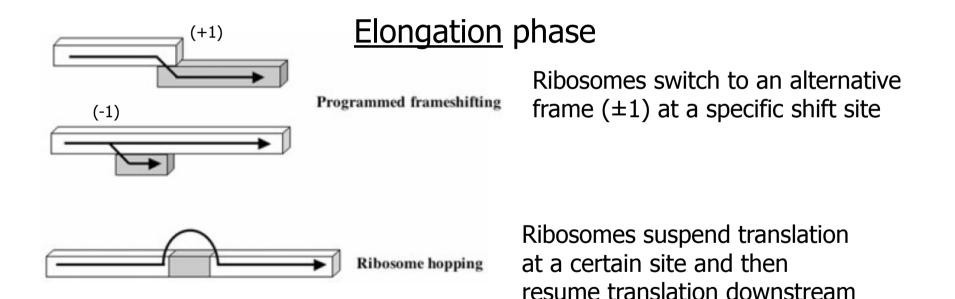
Elongation phase

- +1 Frameshifting
- -1 Frameshifting
- Ribosome hopping

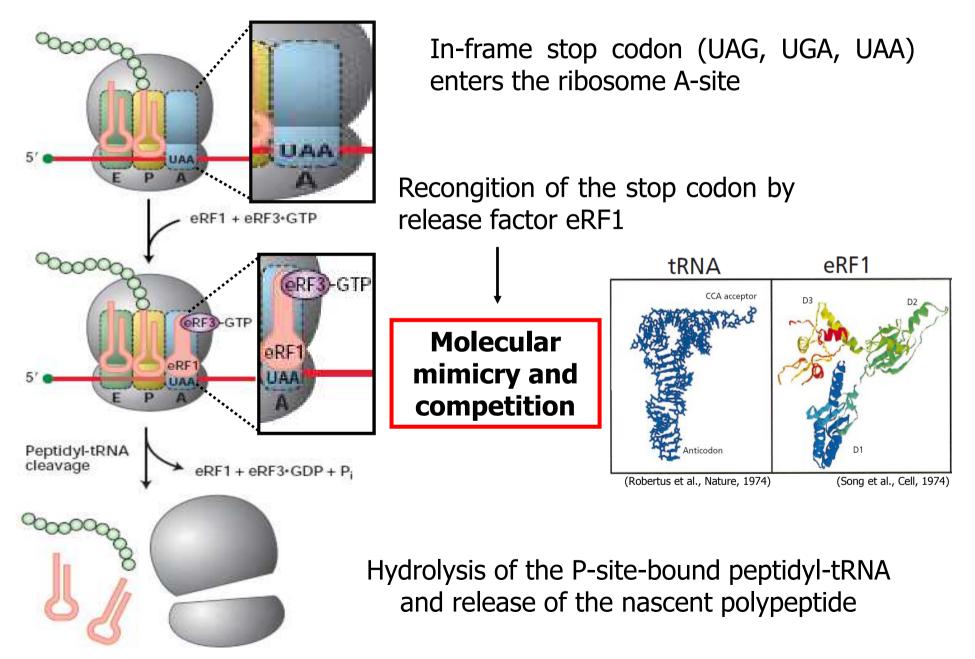
Termination phase• Stop codon Readthrough(frequency: 10-4)

Translational reprogrammed genetic decoding (RECODING) during protein synthesis

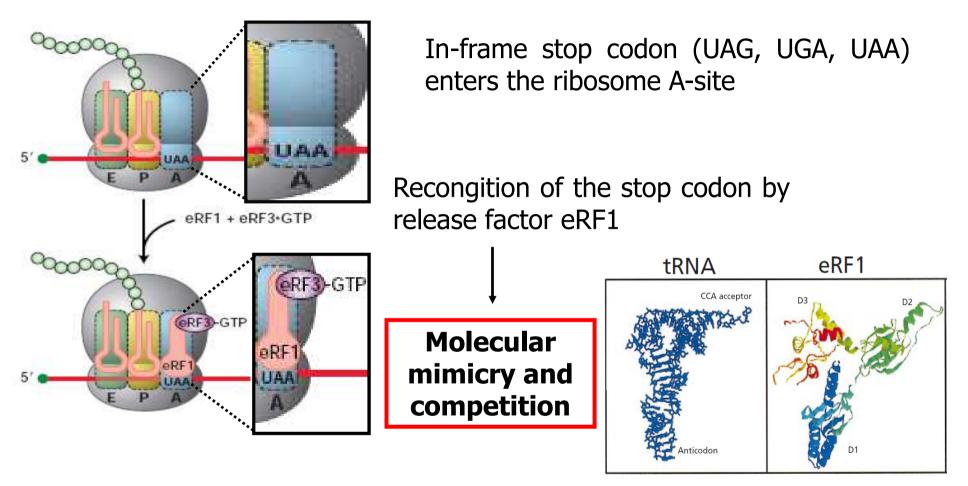
Recoding: regulatory mechanisms of protein expression that include several non-canonical events, opposite to the DNA \rightarrow RNA \rightarrow Protein central dogma of biology



Translation termination



Translation termination



Termination phase



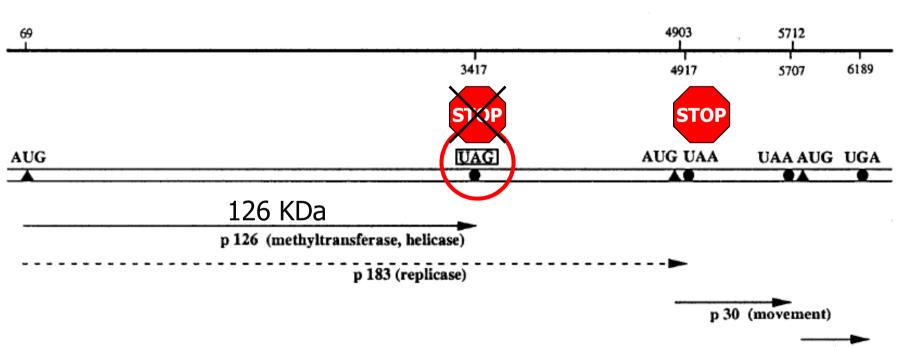
Stop codon readthrough

Translation is continued <u>beyond</u> the stop codon

Translational READTHROUGH

A regulatory mechanism of gene expression, extensively used by ssRNA viruses, which provides the differential production of more than one polypeptide from a single mRNA

Virus (small genome) \rightarrow **Readthrough** \rightarrow Expansion of genetic information



TMV RNA (6395 nt)

р 17.5 (ср)

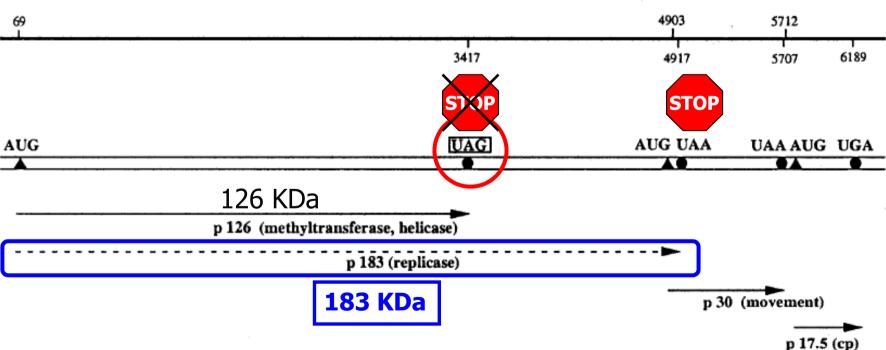
Beier and Grimm, Nucleic Acids Res (2001)

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Beier and Grimm, Nucleic Acids Res (2001)

Termination codons can be "Leaky" stop signals

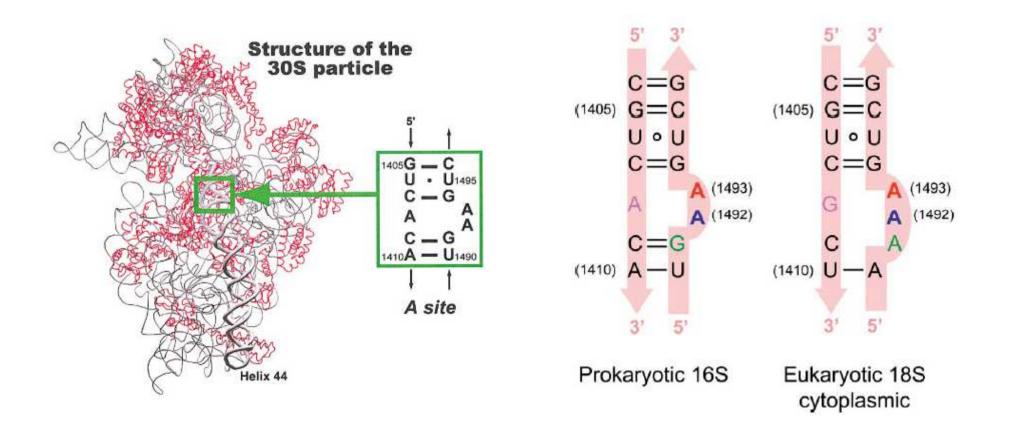
Virus	Genus	"Leaky" termination codon	Readthrough product function
Enterobacteria phage Qβ	Allolevivirus	UGA	Coat protein extension; assembly
Murine leukemia virus (MuLV)	Gammaretrovirus	UAG	Reverse Transcriptase
Sindbis virus (SIN)	Alphavirus	UGA	Replicase
Tomato bushy stunt virus (TBSV)	Tombusvirus	UAG	Replicase
Carnation mottle virus (CarMV)	Carmovirus	UAG	Replicase
Tobacco necrosis virus (TNV)	Necrovirus	UAG	Replicase
Maize chlorotic mottle virus (MCMV)	Machlomovirus	UAG	Replicase
Barley yellow dwarf virus (BYDV)	Luteovirus	UAG	Coat protein extension: aphid transmission
Potato leafroll virus (PLRV)	Polerovirus	UAG	Coat protein extension; aphid transmission
Pea enation mosaic virus (PEMV)	Enamovirus		
RNA-1		UGA	Cost protein extension; aphid transmission
Tobacco mosaic virus (TMV)	Tobamovirus	UAG	Replicase
Tobacco rattle virus (TRV)	Tobravirus		
RNA-1	202025104893034950	UGA	Replicase
Peanut clump virus (PCV)	Pecluvirus		The second s
RNA-1		UGA	Replicase
Soil-borne wheat mosaic virus (SBWMV)	Furovirus		
RNA-1		UGA	Replicase
RNA-2	1	UGA	Coat protein extension;
			fungus transmission
Potato mop-top virus (PMTV)	Pomovirus	100 million (1975)	
RNA-1		UGA	Replicase
RNA-3	Level and	UAG	Coat protein extension
Beet soil-borne virus (BSBV)	Pomovirus	and the second sec	
RNA-1		UAA	Replicase
RNA-2	1	UAG	Coat protein extension
Broad bean necrosis virus (BBNV)	Pomovirus		2
RNA-2	1	UAA	Coat protein extension
Beet necrotic yellow vein virus (BNYVV)	Benyvirus		
RNA-2	÷	UAG	Coat protein extension; fungus transmission
Turnip yellow mosaic virus (TYMV)	Tymovirus	UAG	Replicase extension ?

Beier and Grimm, Nucleic Acids Res (2001)

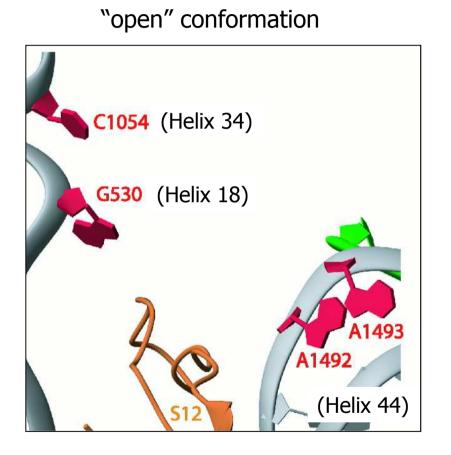
Ribosome Decoding Site

Region located within the **A-site** in the ribosomal small subunit

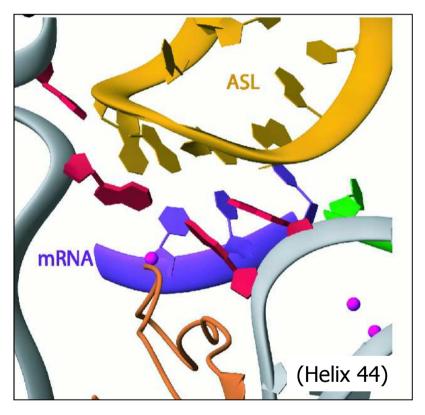
The Decoding Site contains two **adenine nucleotides (A1492 and A1493)** that **monitor codon/anticodon base pairing**



The decoding site switches to a "closed" conformation when the correct (cognate) aa-tRNA enters the A-site



"closed" conformation



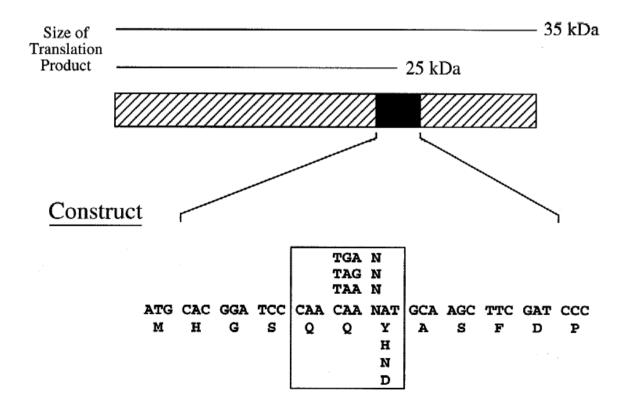
When the codon and a tRNA-ASL bind in the A-site, A1492 and A1493 flip out to monitor the codon-anticodon interaction

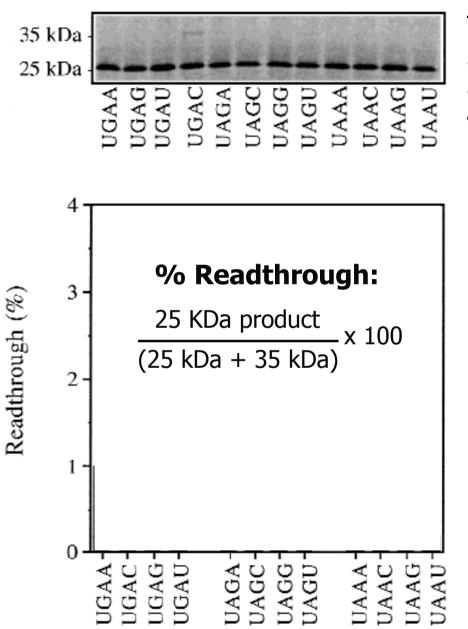
RNA (2000), 6:1044–1055. Cambridge University Press. Printed in the USA. Copyright © 2000 RNA Society.

Aminoglycoside antibiotics mediate <u>context-dependent suppression</u> of termination codons in a mammalian translation system

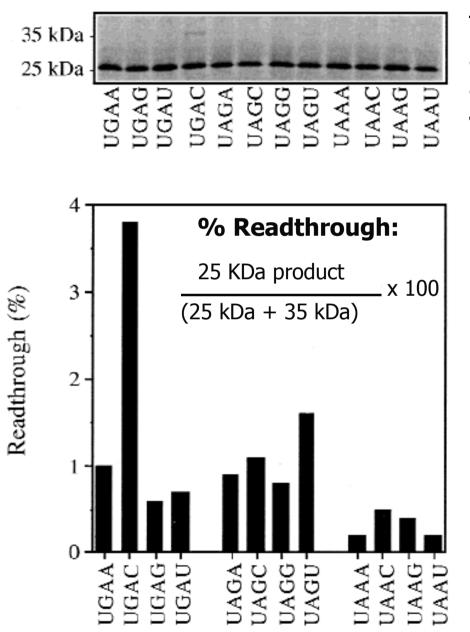
MARINA MANUVAKHOVA,^{1,3} KIM KEELING,² and DAVID M. BEDWELL^{1,2}

Experimental model: synthetic constructs bearing different termination signals are translated *in-vitro* in the presence of [³⁵S]-Met/Cys and the resulting [³⁵S]-labeled polypeptides are analyzed by SDS-PAGE

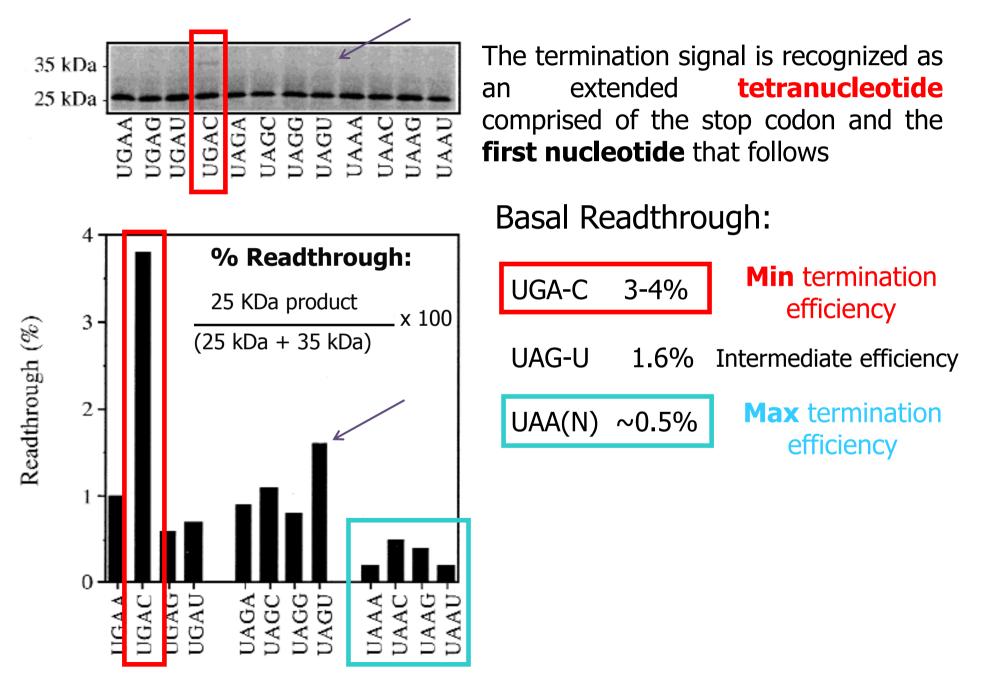


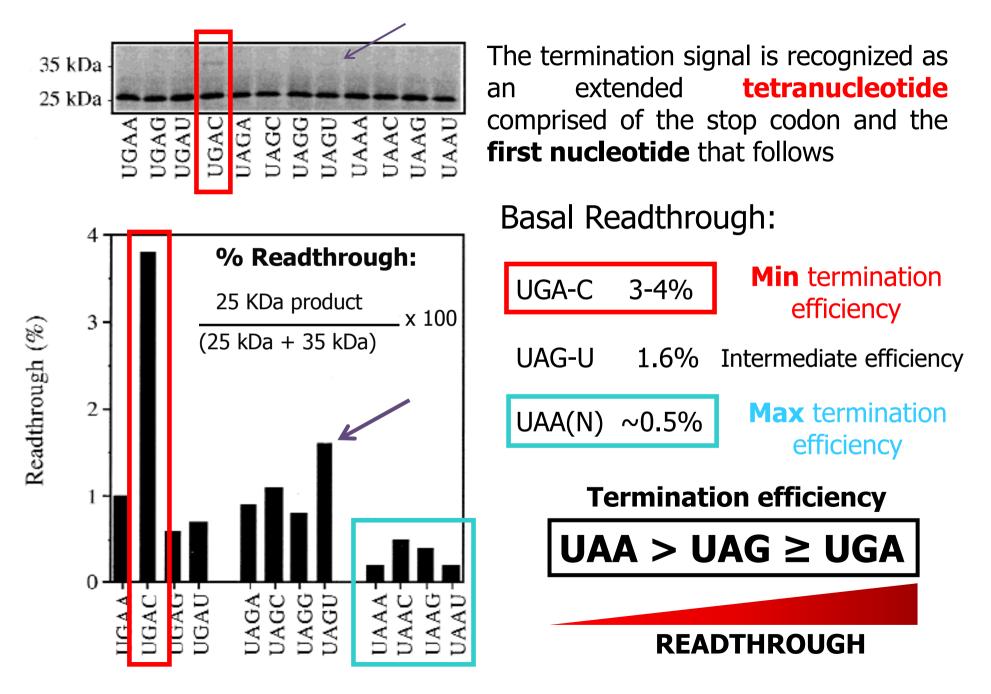


The termination signal is recognized as an extended **tetranucleotide** comprised of the stop codon and the **first nucleotide** that follows

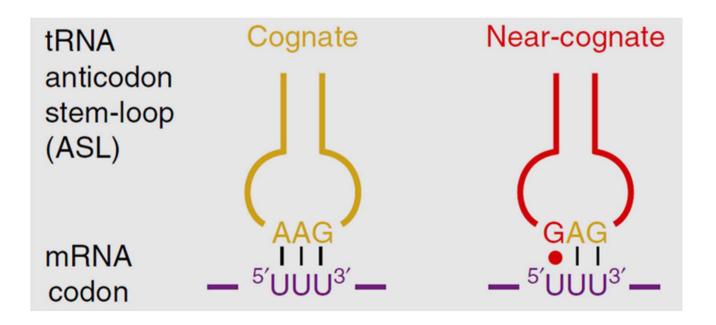


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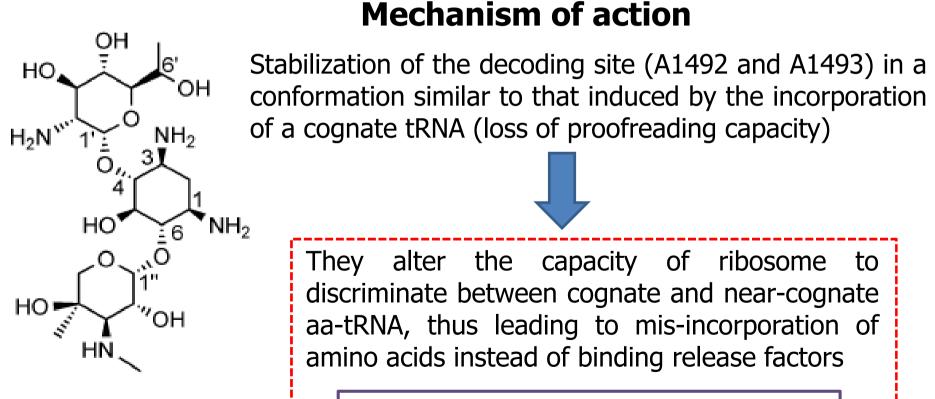
The occurrence of a basal readthrough prepares the ground for the use of molecules that are able to decrease the efficiency of translation termination, thus increasing the efficiency of readthrough itself



Aminoglycosides

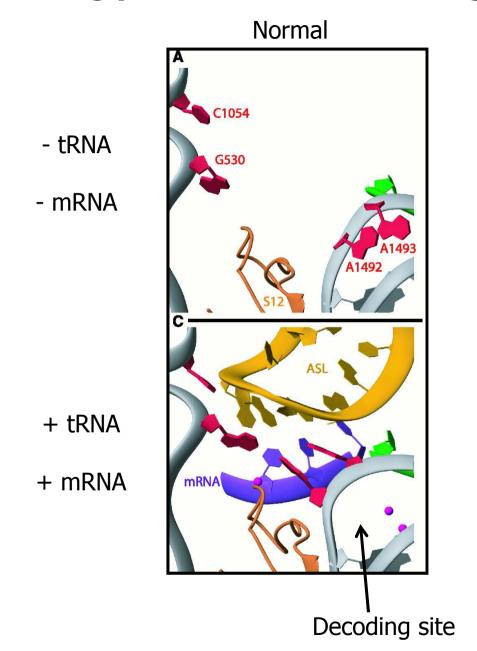
A group of molecules belonging to the class of antibiotics

Aminoglycosides bind the decoding site within the A-site in the ribosomal small subunit

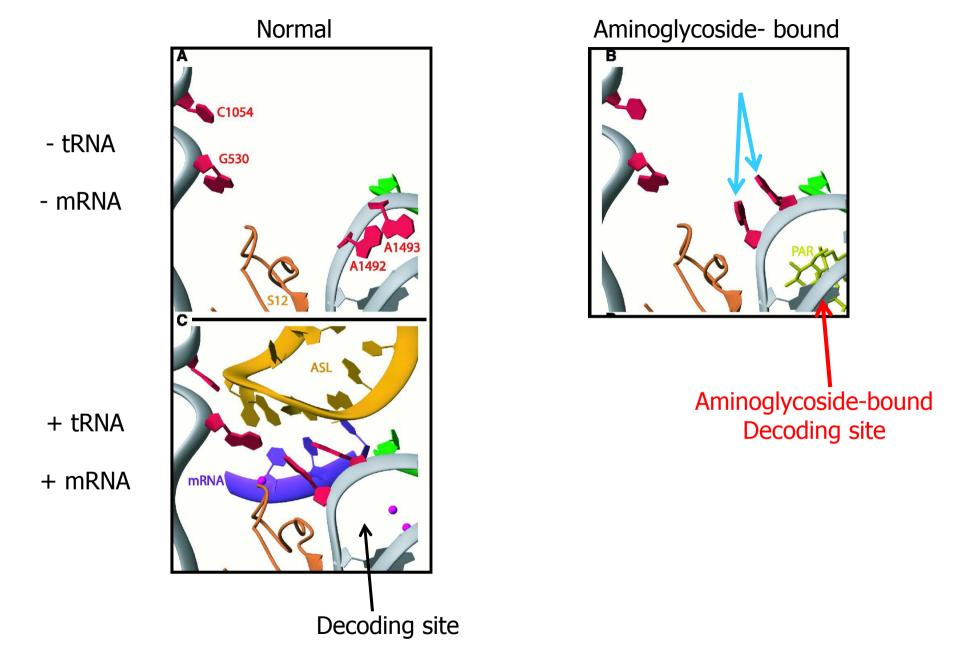


Geneticin (G418)

near-cognate tRNA \approx cognate tRNA

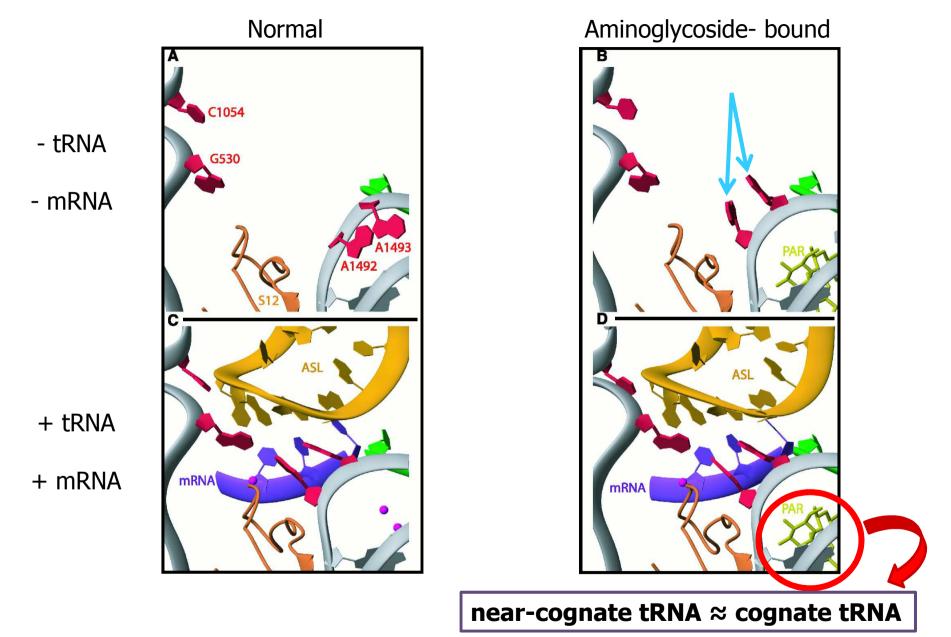


Aminoglycosides bind the decoding site and reduce ribosome fidelity



Aminoglycosides bind the decoding site and reduce ribosome fidelity

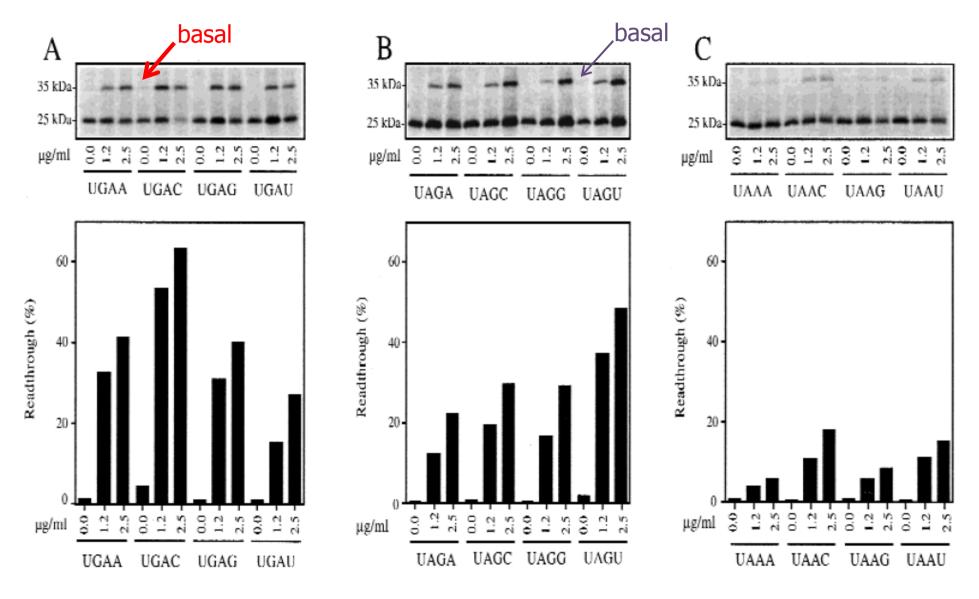
Ogle et al., Science (2001)



Aminoglycosides bind the decoding site and reduce ribosome fidelity

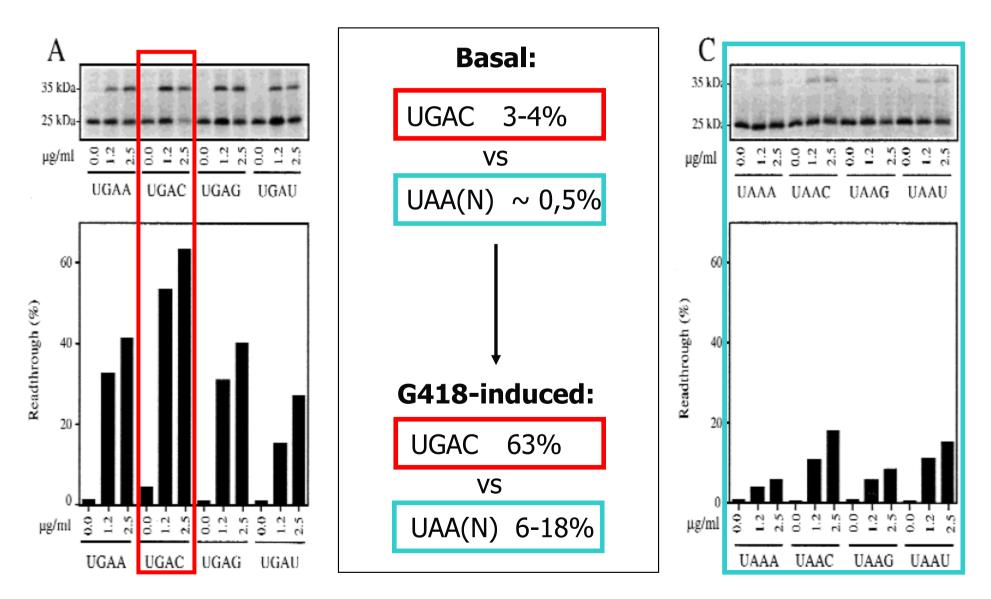
Treatment with aminoglycosides (G418) enhances readthrough

The context and/or stop codon with the highest basal readthrough (**UGAC** or UGA in general) display the most efficient G418-induced readthrough.



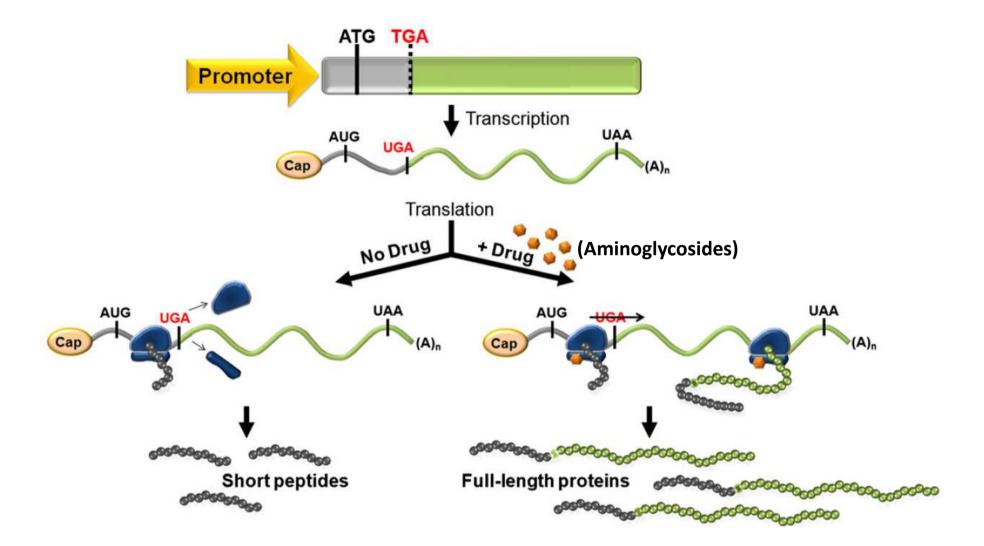
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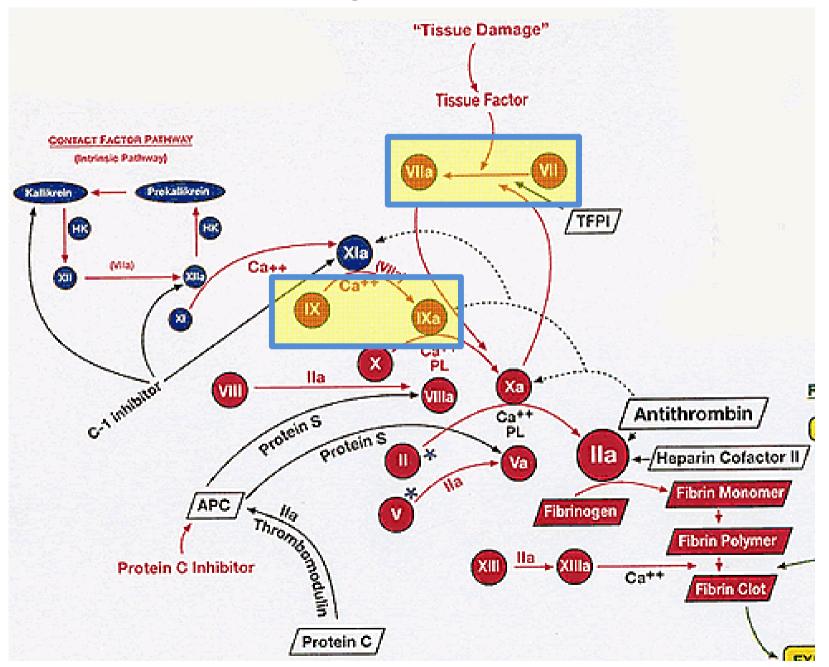


Parte II Metodologie Sperimentali

The readthrough process



The coagulation cascade



Workflow

1. Creation of expression vector(s)

- 2. Expression studies in eukaryotic cells- treatment of cells with readthrough-inducing drug
- 3. Analysis of protein isoforms
- 4. Evaluation of (pro-coagulant) activity

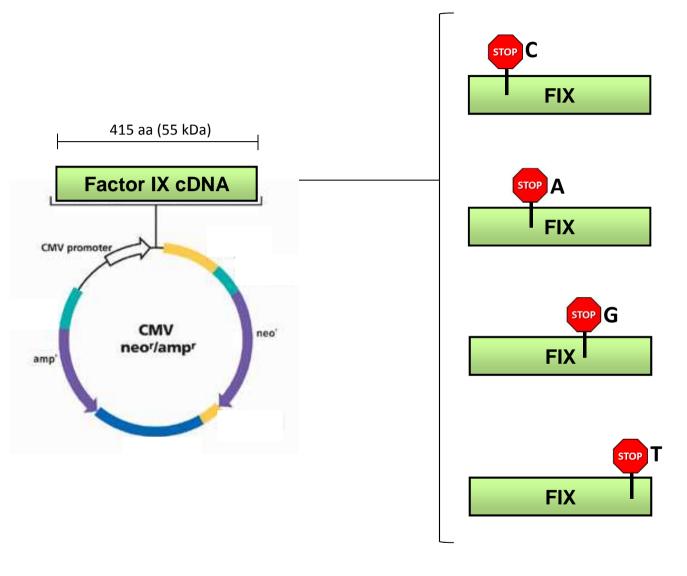
Workflow

1. Creation of expression vector(s)

2. Expression studies in eukaryotic cells- treatment of cells with readthrough-inducing drug

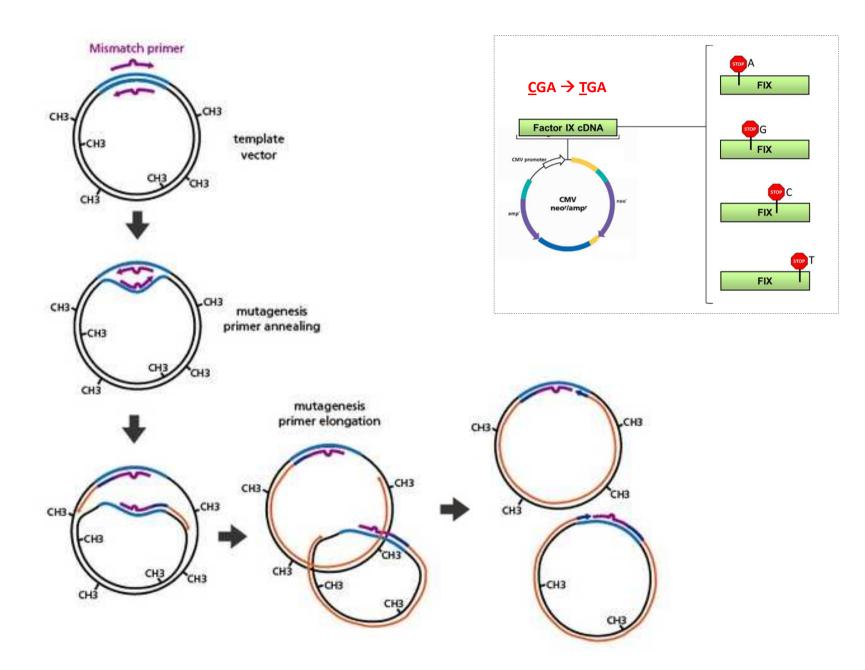
3. Analysis of protein isoforms

1. Creation of expression vectors - Site-directed mutagenesis



Nonsense mutation inserted: <u>CGA → TGA</u> + C-A-G-T (4th nucleotide)

1. Creation of expression vectors - Site-directed mutagenesis



Workflow

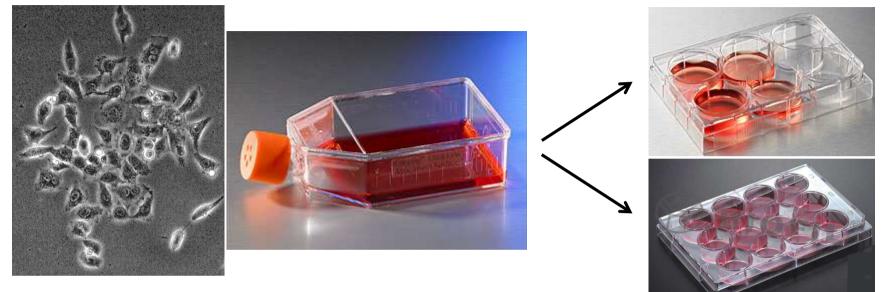
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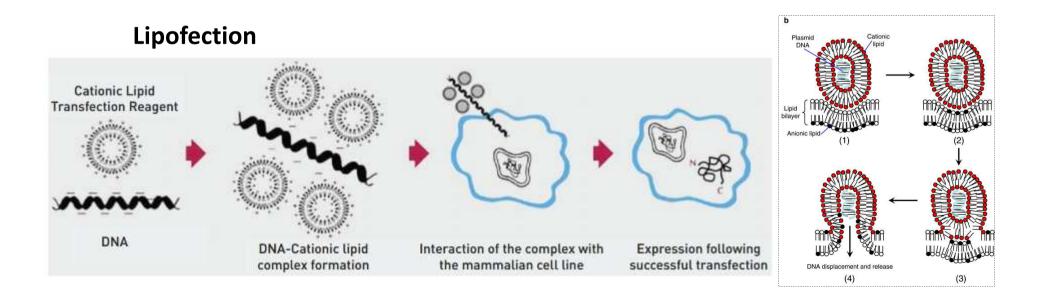
2. Expression studies in eukaryotic cells- treatment of cells with readthrough-inducing drug

3. Analysis of protein isoforms

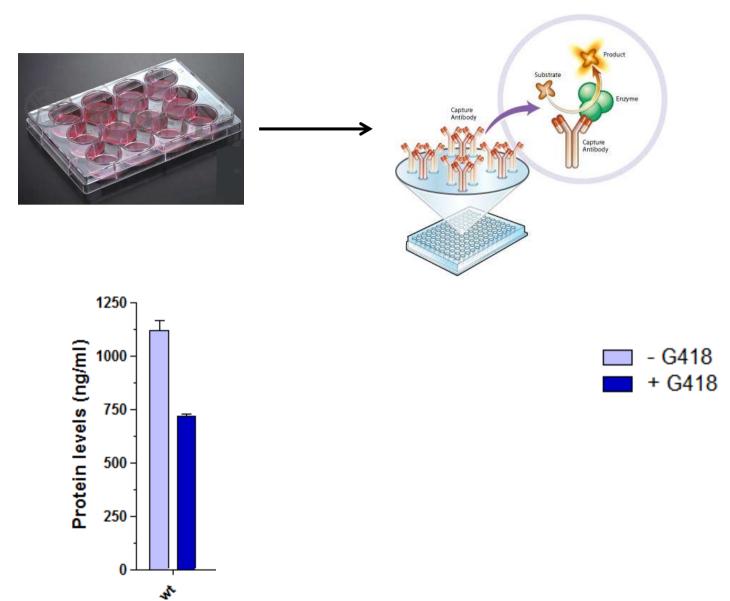
2. Expression studies in eukaryotic cells - Transfection

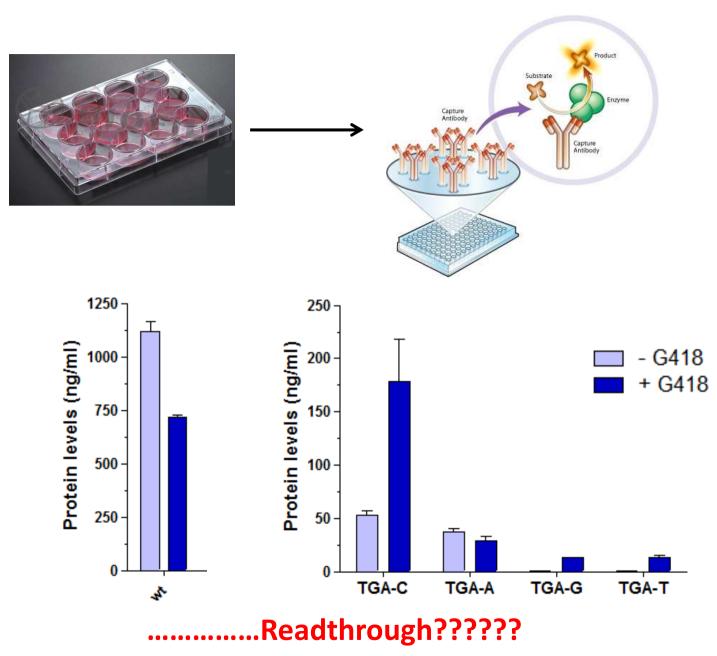
<u>Human Embryonic Kidney (HEK) cells</u>





2. Expression studies in eukaryotic cells - ELISA





2. Expression studies in eukaryotic cells - ELISA

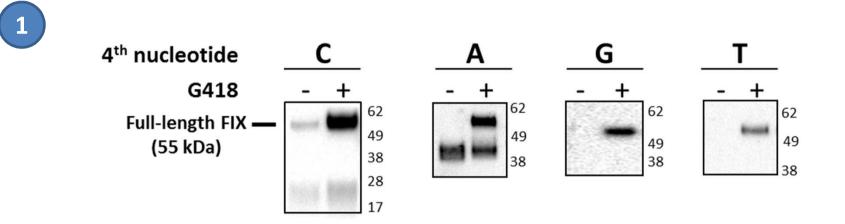
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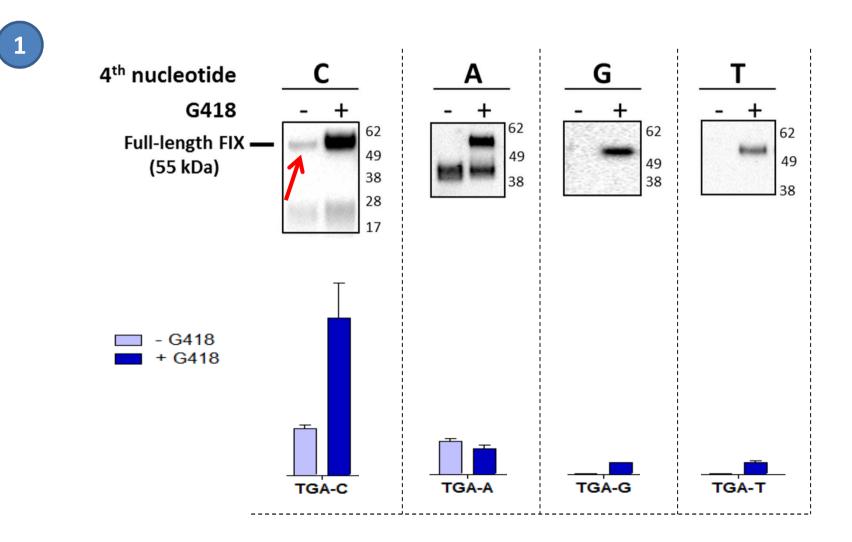
2. Expression studies in eukaryotic cells- treatment of cells with readthrough-inducing drug

3. Analysis of protein isoforms

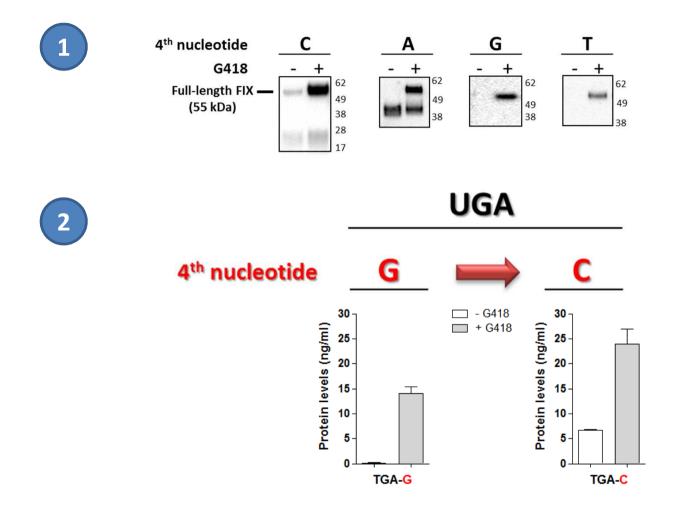
3. Analysis of protein isoforms – Western blotting



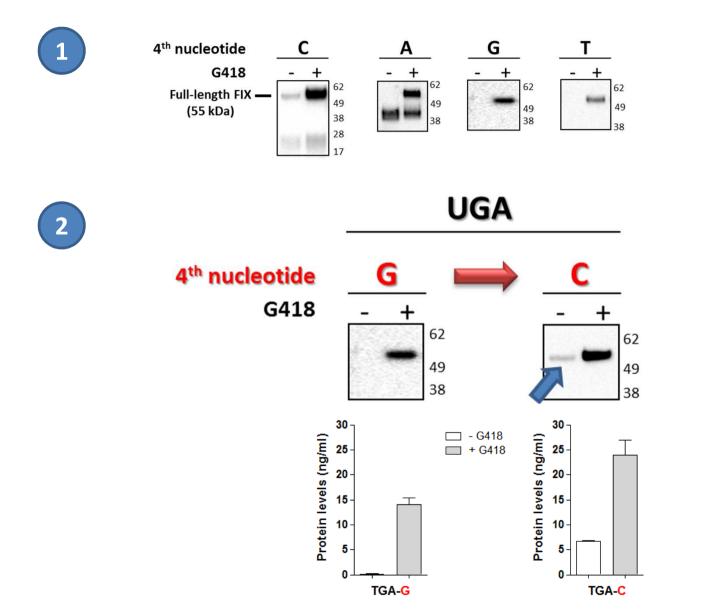
3. Analysis of protein isoforms – Western blotting



3. Analysis of protein isoforms – Sequence context effect



3. Analysis of protein isoforms – Sequence context effect

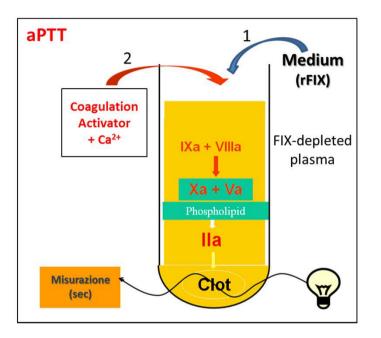


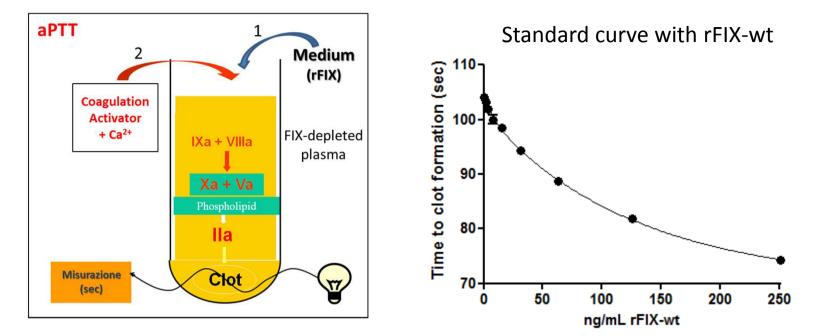
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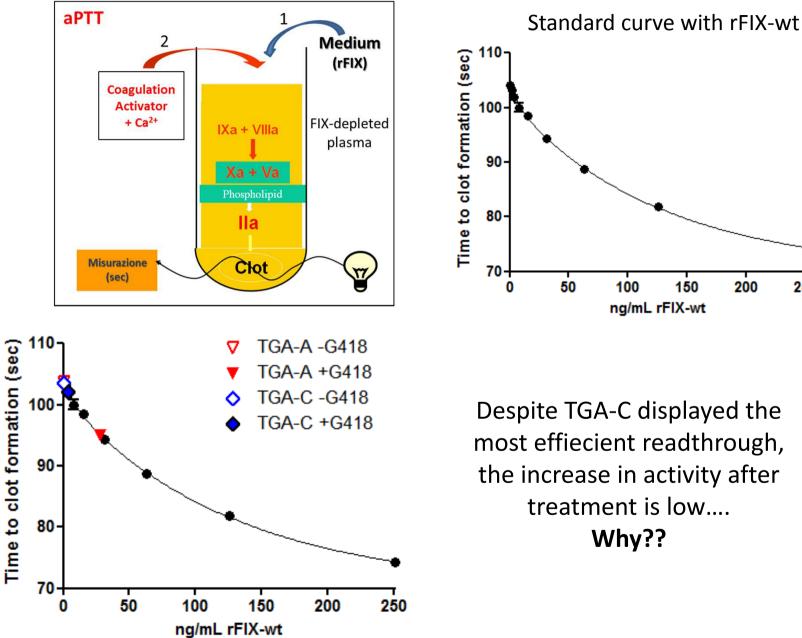
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4. Evaluation of (pro-coagulant) activity



250

Summary

