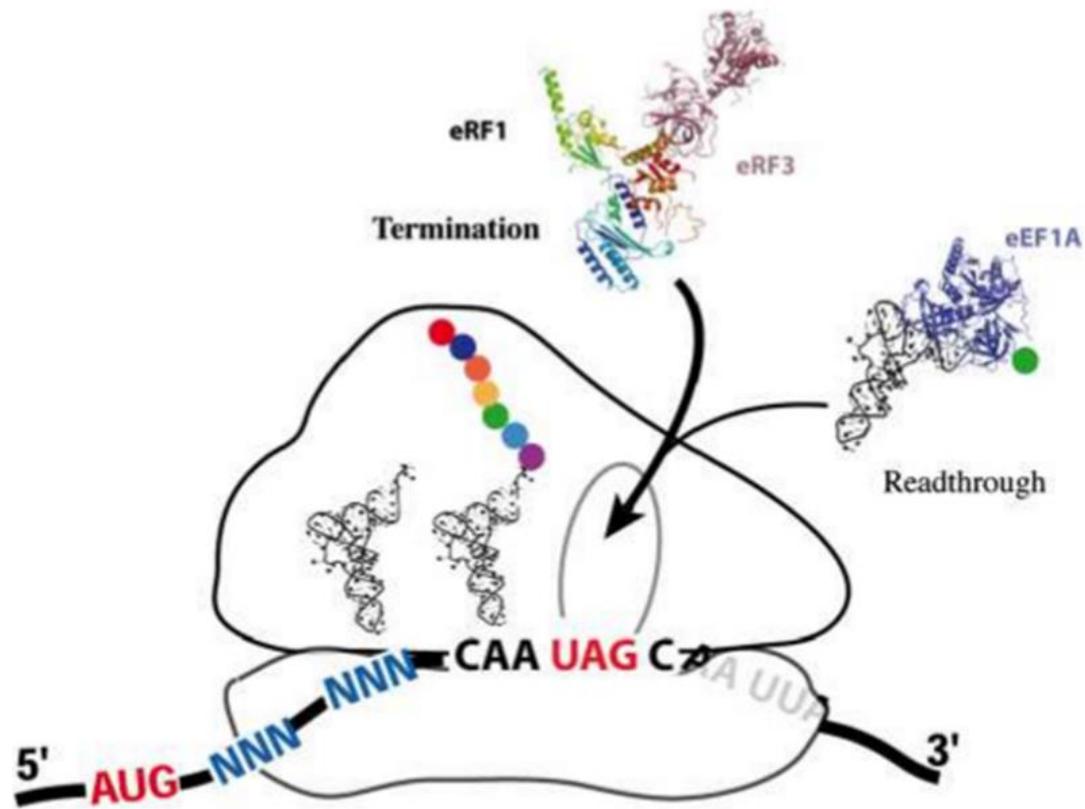


Ribosome readthrough



Starting from the base...PROTEIN SYNTHESIS

Eukaryotic translation can be divided into four stages: Initiation, Elongation, Termination and Recycling

During translation, the ribosome catalyzes the sequential addition of amino acids to a growing polypeptide chain, using an mRNA as template and aminoacyl-tRNAs as substrates

Correct base pairing between the three bases of the codon on mRNA and those of the anticodon of the aa-tRNA dictates the sequence of the polypeptide chain

Translation termination

When the ribosome arrives at a **stop codon**, there is **no corresponding tRNA**. Instead, a **release factor** enters the assembly site and **synthesis is terminated**, releasing the completed polypeptide from the ribosome.

Translation termination

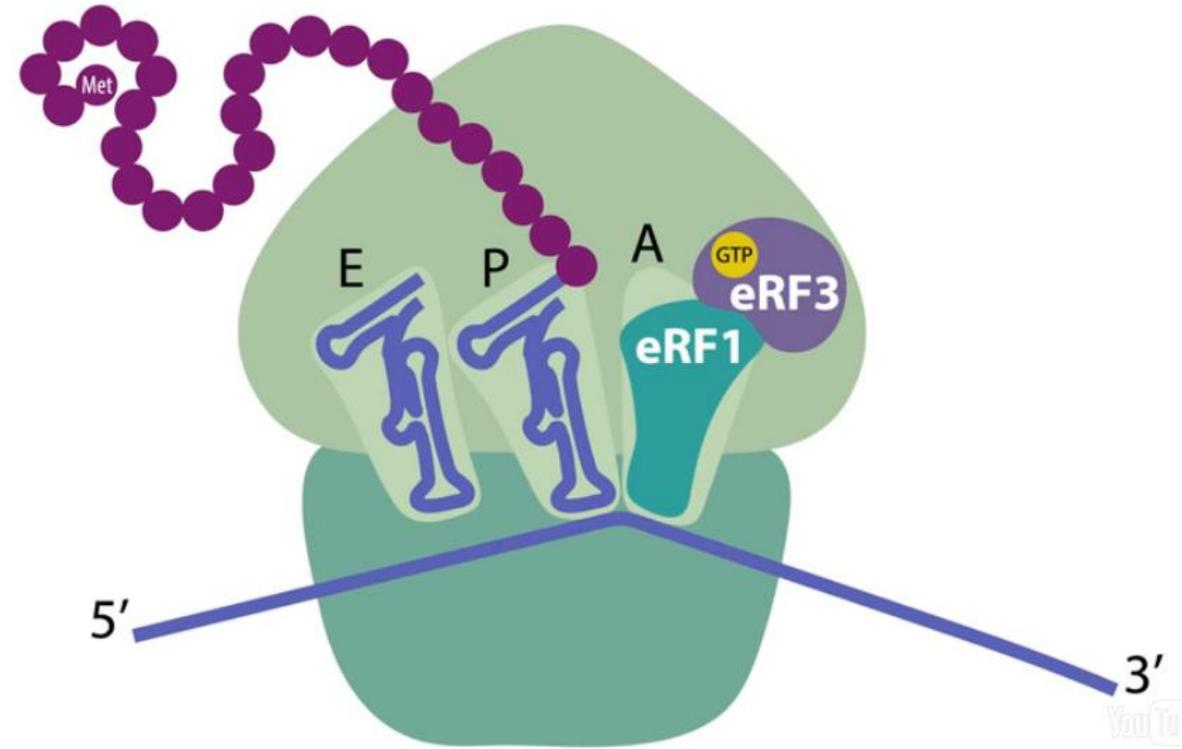
When the ribosome arrives at a **stop codon**, there is **no corresponding tRNA**. Instead, a **release factor** enters the assembly site and **synthesis is terminated**, releasing the completed polypeptide from the ribosome.

Translation termination in eukaryotes is mediated by two release factors:

- **eRF1** recognizes each of the three stop codons (UAG, UAA, and UGA) and facilitates release of the nascent polypeptide chain
- **eRF3** is a GTP binding protein that facilitate the termination process

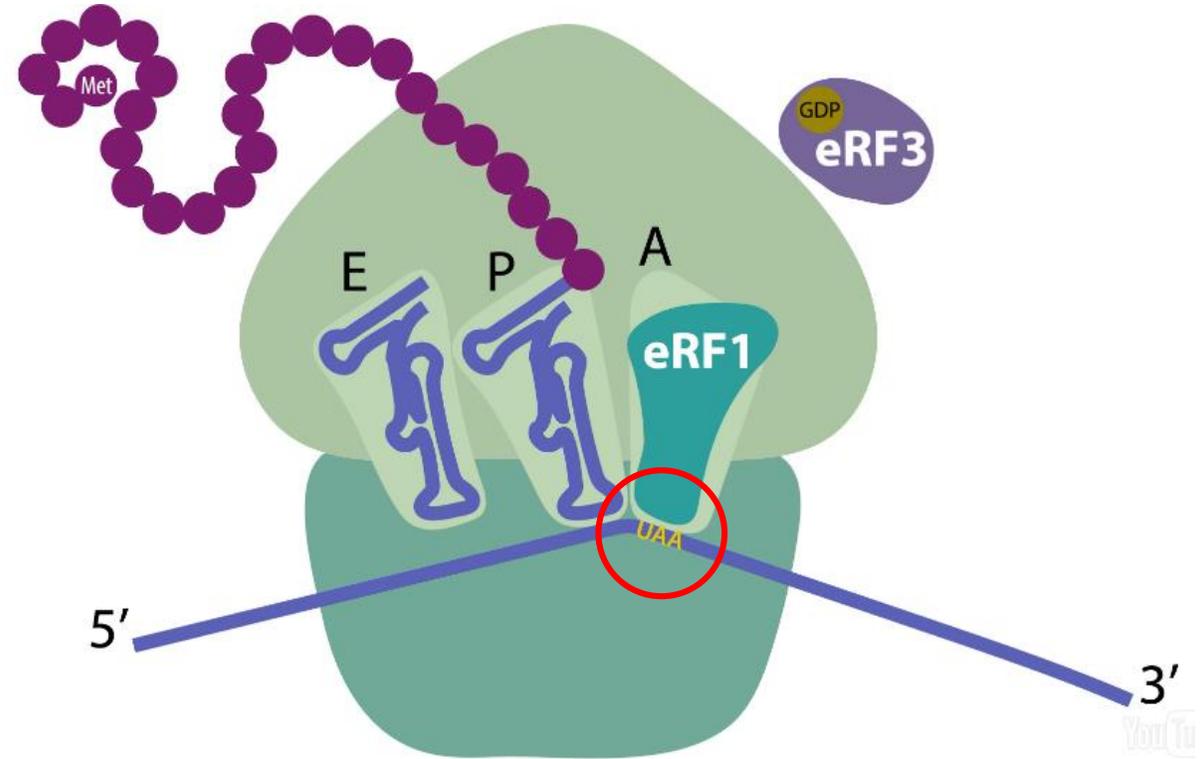
Translation termination

1. A complex formed by **eRF1** and **eRF3-GTP** enters the ribosome



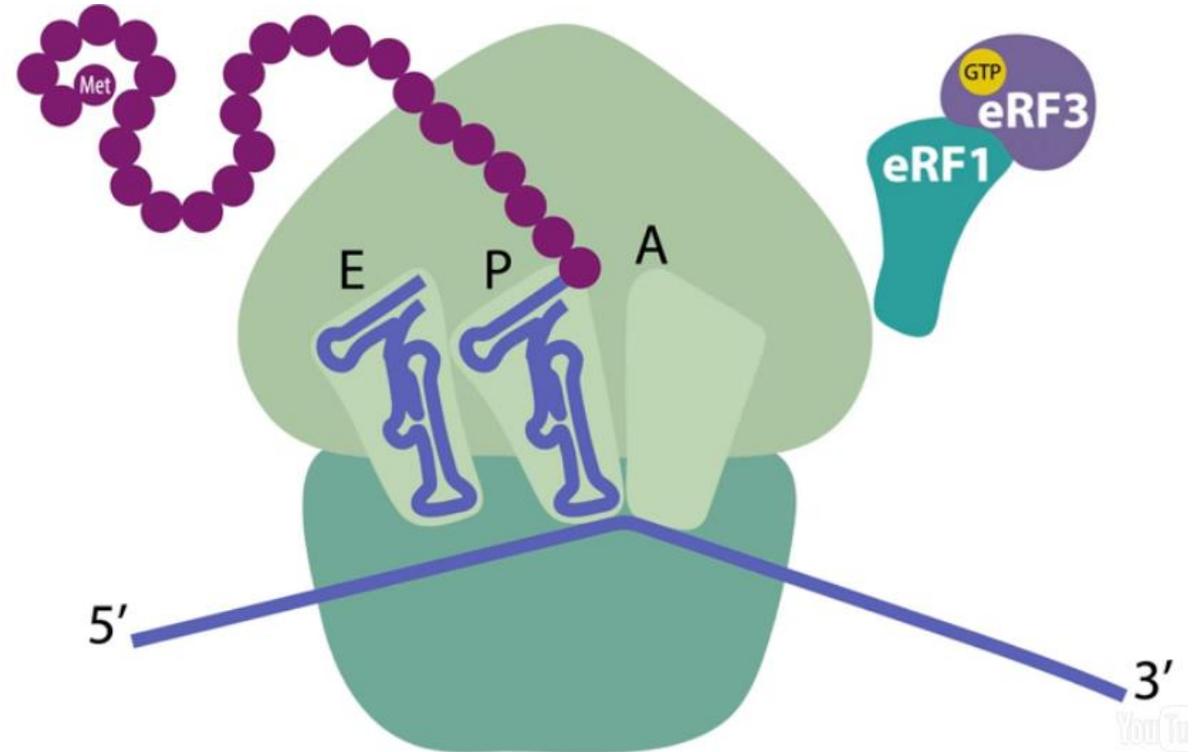
Translation termination

1. A complex of eRF1 and eRF3-GTP enters the ribosome
2. The **stable interaction** between eRF1 and a stop codon in the ribosomal A site stimulates **GTP hydrolysis** by eRF3



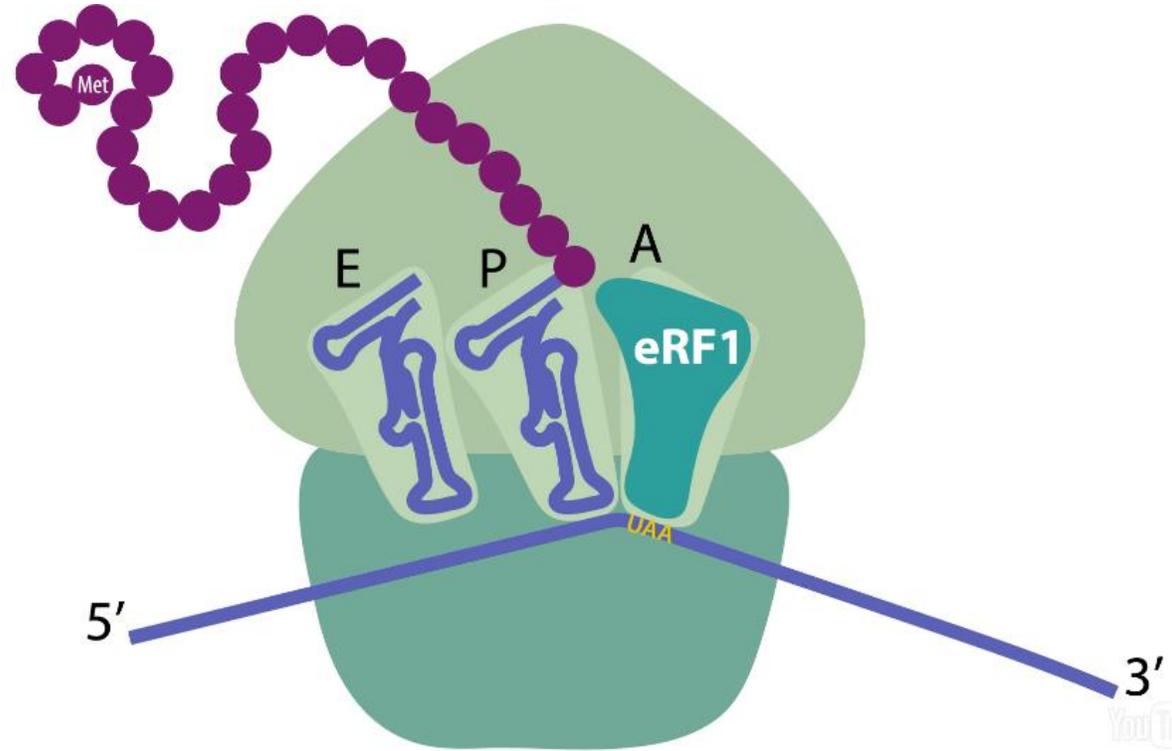
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Translation termination

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3. GTP hydrolysis **activates eRF1** so that it can efficiently stimulate nascent chain release

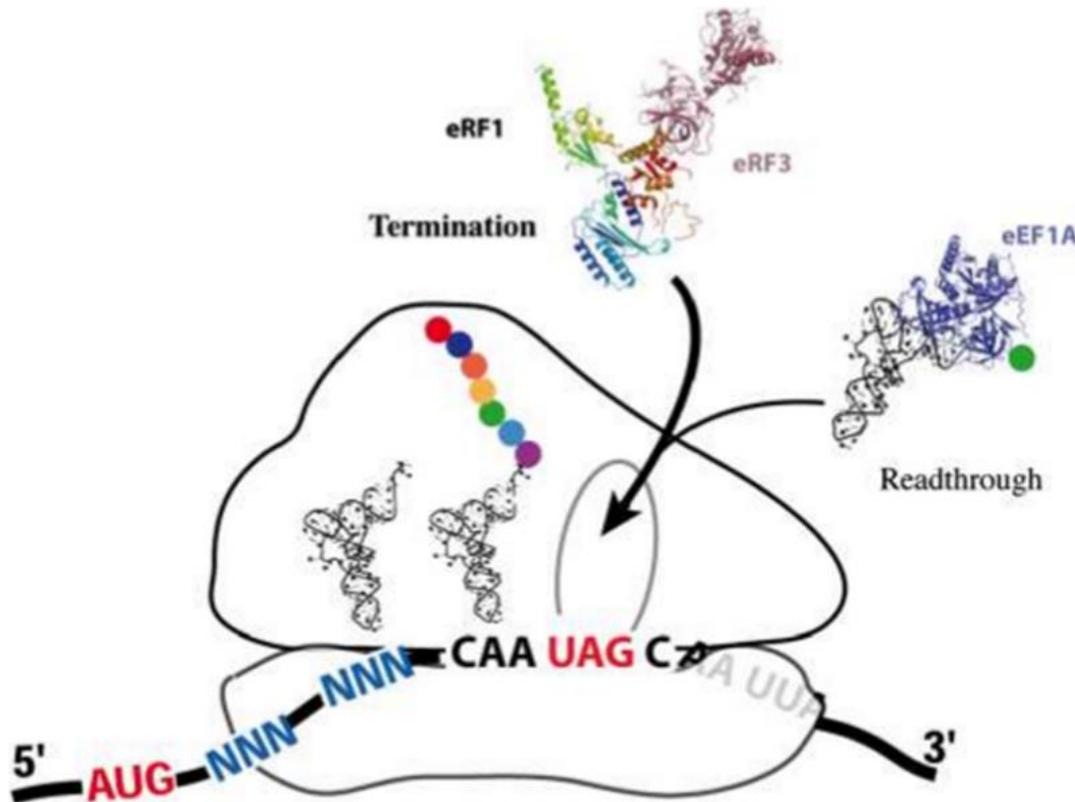


Translation termination is normally a highly efficient process

BUT occasionally stop codon recognition by eRF1 can be superseded by selected aminoacyl-tRNAs, resulting in **stop codon suppression**

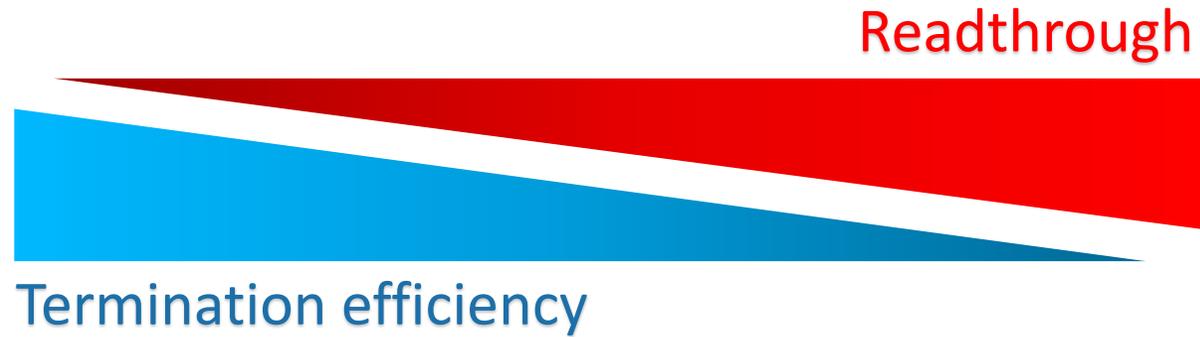
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This event is called **READTHROUGH** and corresponds to the **incorporation of a tRNA**, or natural suppressor, **at the stop codon**, allowing translation to continue in the same frame until the ribosome reaches the next stop

The more efficient is translation termination, the less frequent is readthrough (and viceversa)



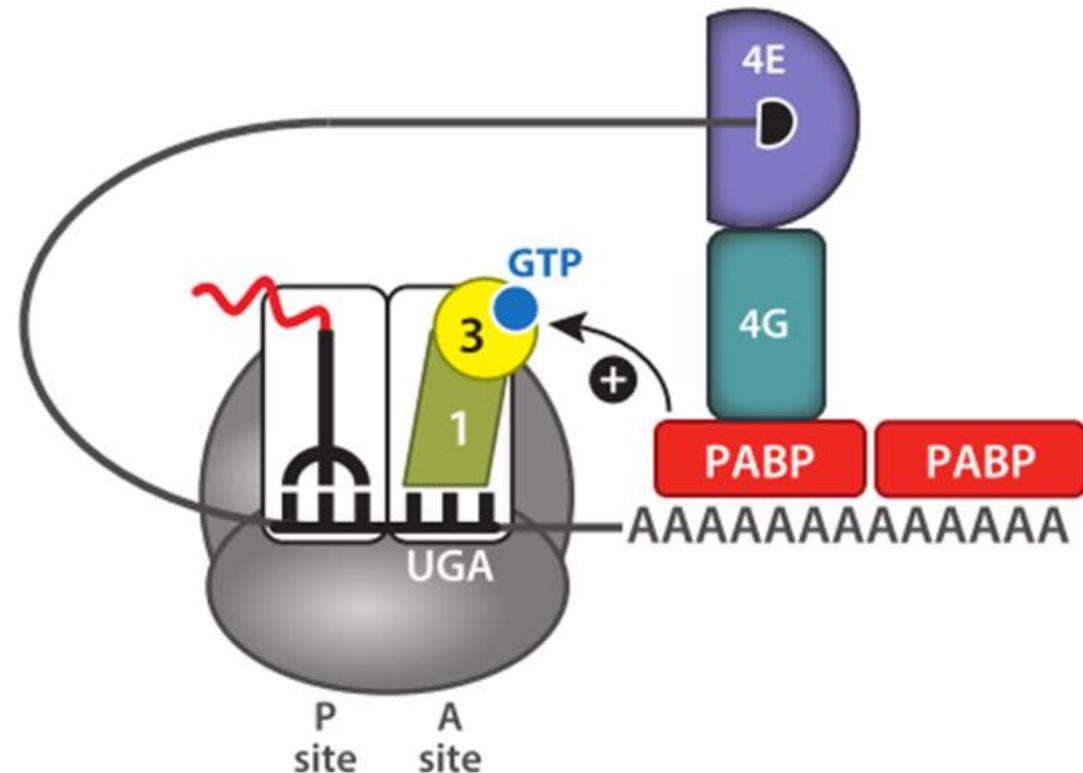
The efficiency of translation termination (and the occurrence of readthrough) can vary depending on many factors

The efficiency of translation termination (and the occurrence of readthrough) can vary depending on:

1) The efficiency of termination differs between normal stop codons and premature termination codons (PTC)

Normal STOP codon

PABP interacts with eRF3, promoting translation termination

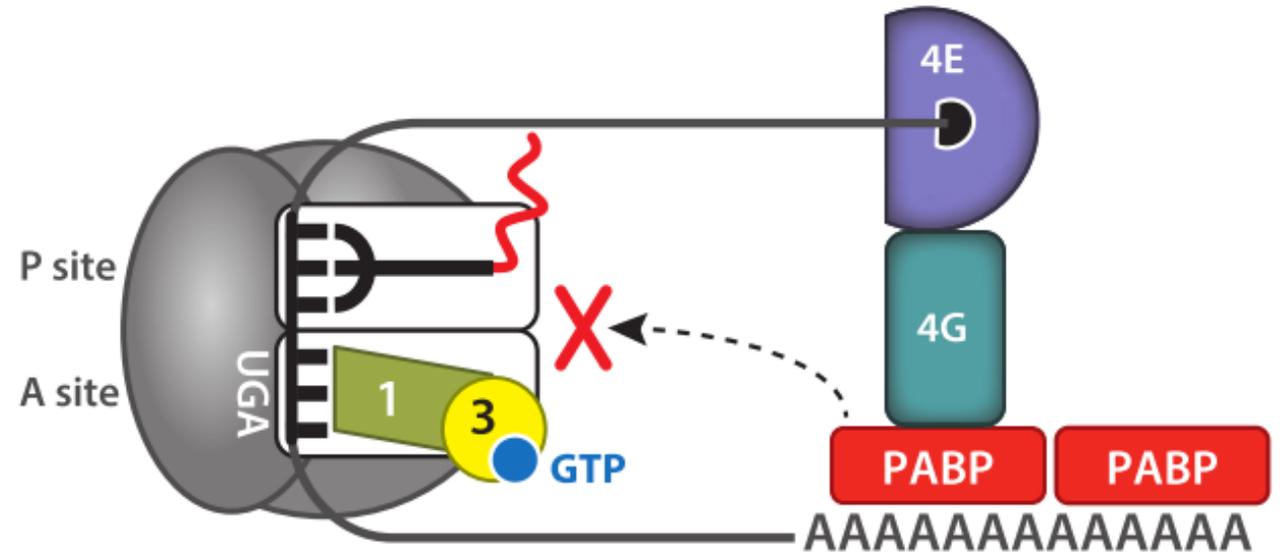


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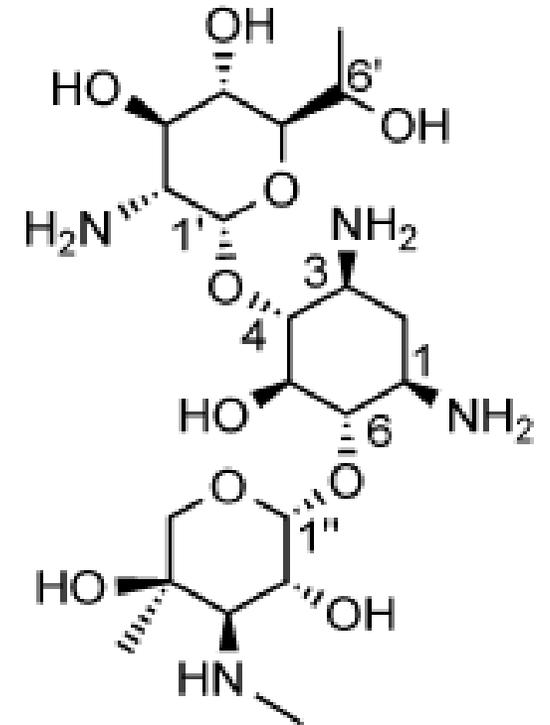
Premature STOP codon

PABP can not interact with eRF3, leading to prolonged ribosomal pausing at PTC and increasing aa-tRNA sampling



2) Aminoglycosides can decrease the fidelity of translation, causing higher frequencies of readthrough

Aminoglycosides are a class of antibiotics that interfere with bacterial-protein synthesis. They all have a common 2-deoxystreptamine ring structure, which binds to the ribosome decoding center.



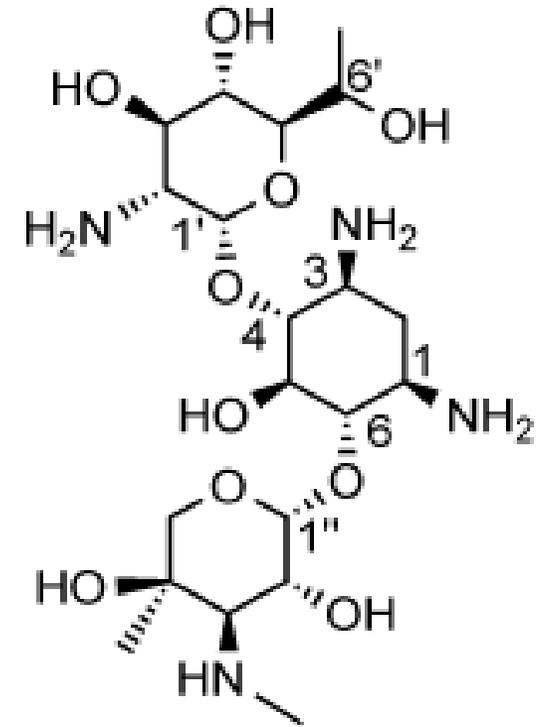
Geneticin (G418)

2) Aminoglycosides can decrease the fidelity of translation, causing higher frequencies of readthrough

Aminoglycosides are a class of antibiotics that interfere with bacterial-protein synthesis. They all have a common 2-deoxystreptamine ring structure, which binds to the ribosome decoding center.

Ribosome is composed by **two subunits**:

- The large subunit contains the peptidyl transferase center, in which peptide bonds are formed
- The **small subunit** contains the **decoding center**, a region in which the correct codon-anticodon pairing between mRNA and tRNAs is monitored

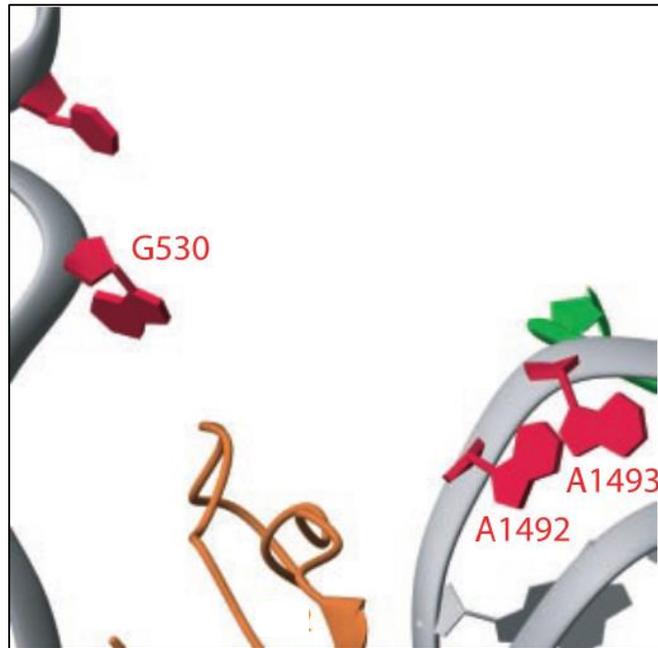


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Mechanism of action: aminoglycosides binding to ribosome decoding site induces a conformational change similar to the transition caused by a tRNA binding

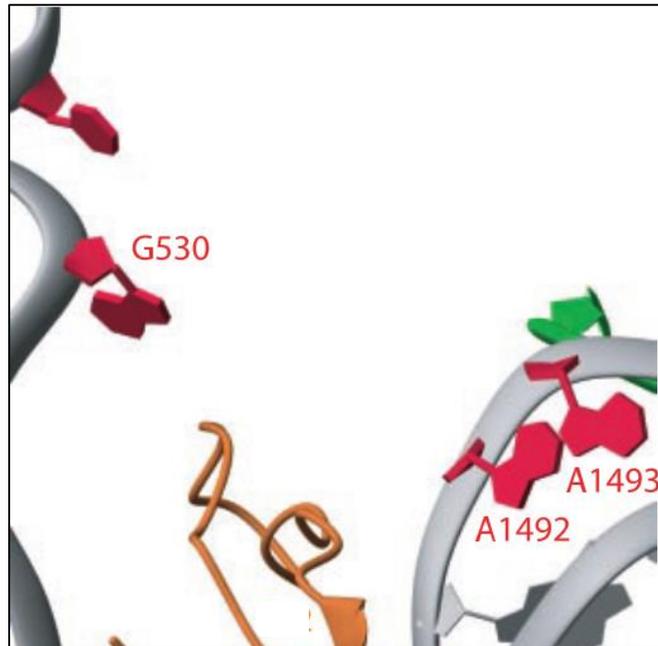
NATIVE STRUCTURE



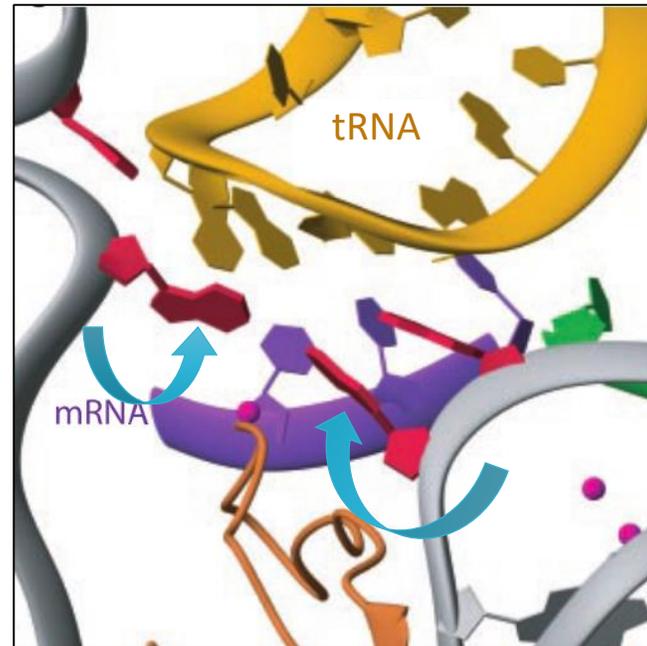
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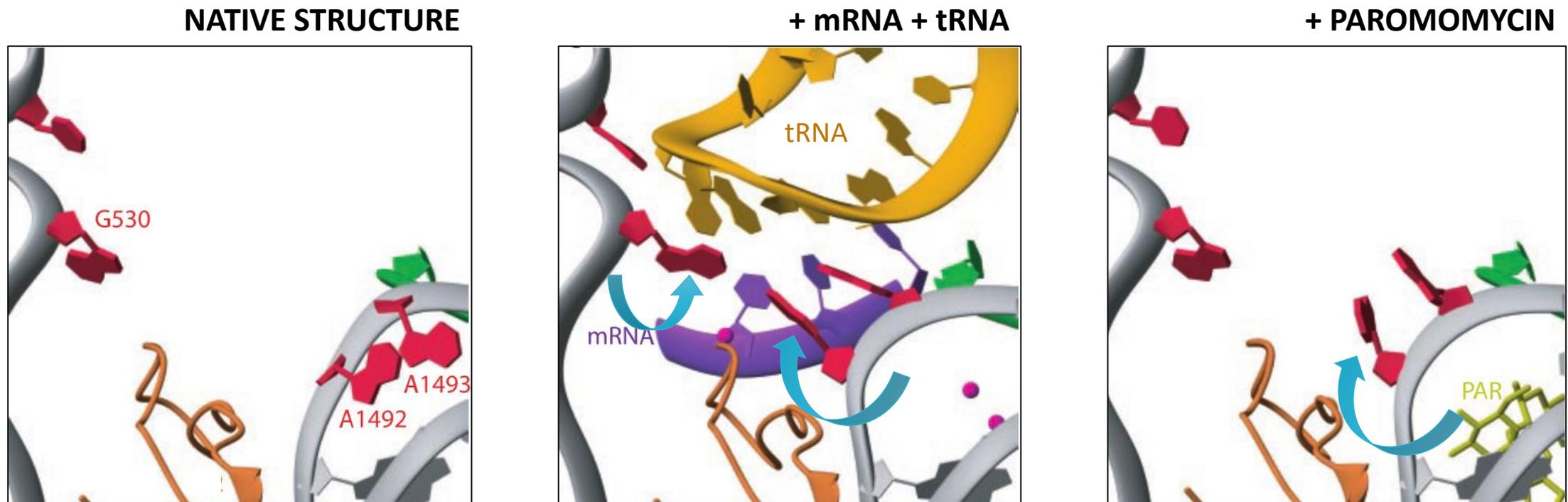


+ mRNA + tRNA



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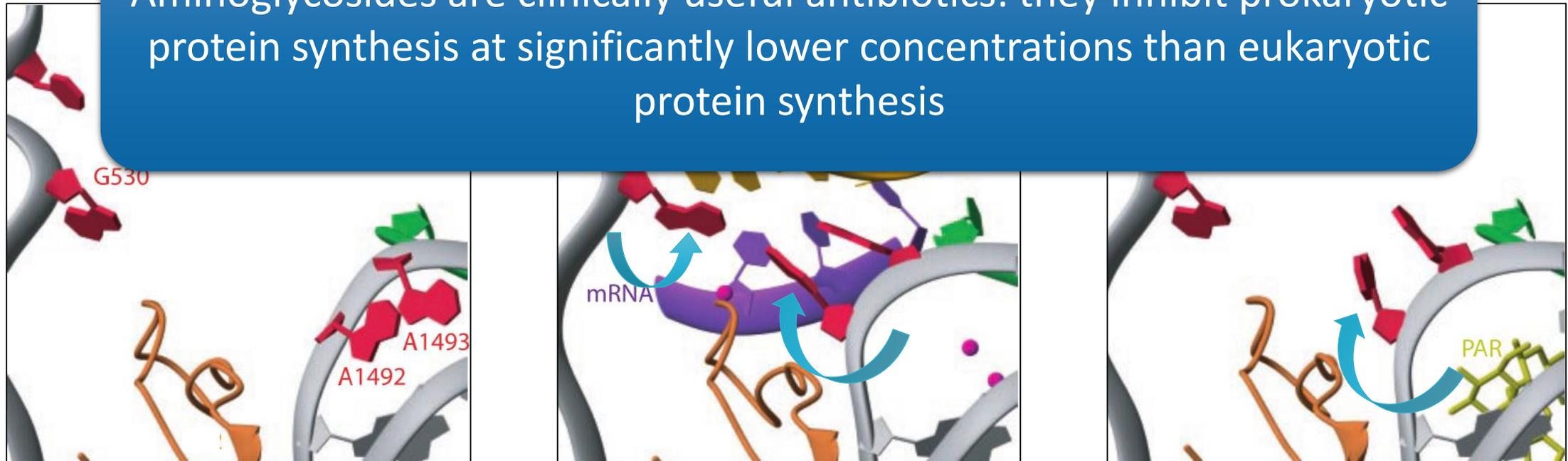
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Aminoglycosides are clinically useful antibiotics: they inhibit prokaryotic protein synthesis at significantly lower concentrations than eukaryotic protein synthesis



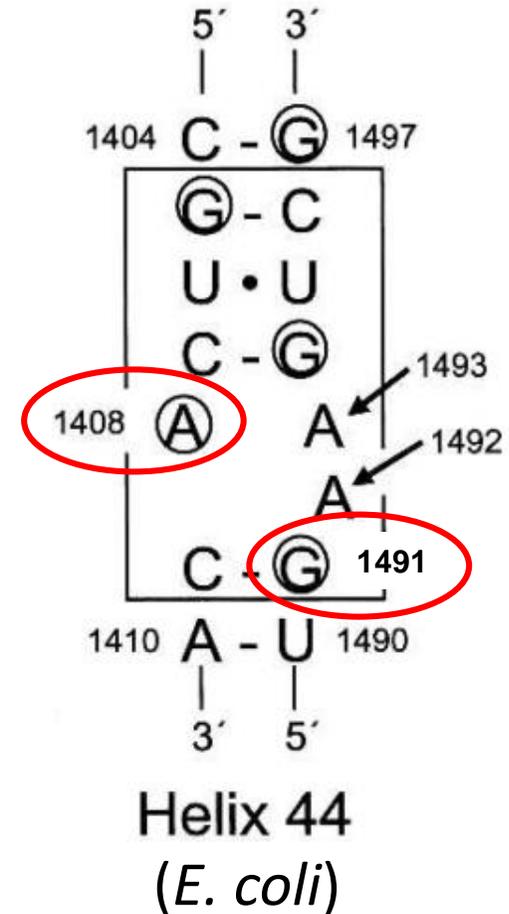
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The major determinants of the **differential** aminoglycoside **sensitivity** between prokaryotes and eukaryotes are **two non conserved residues** of the **decoding center**

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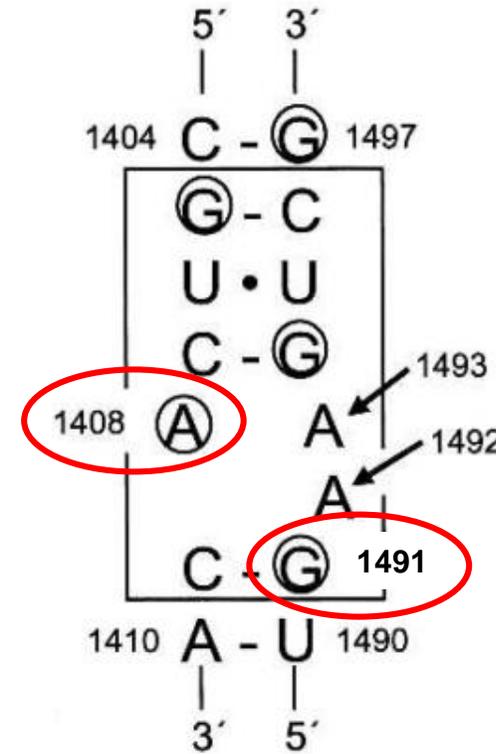
A1408 and **G1491**



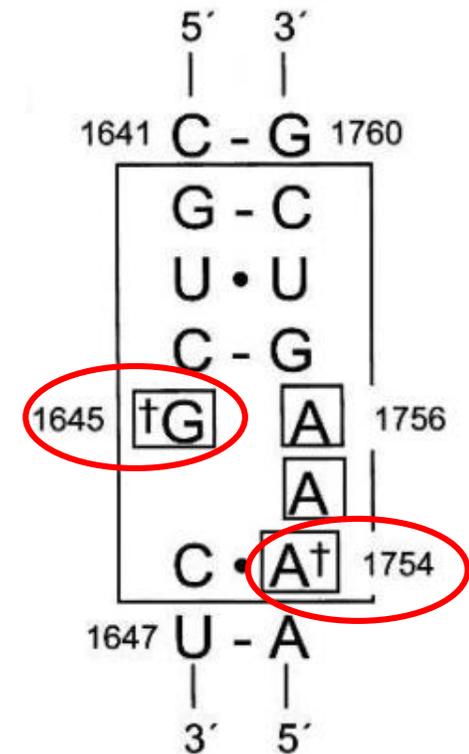
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A1408 and **G1491** VS **G1645** and **A1754**



Helix 44
(*E. coli*)



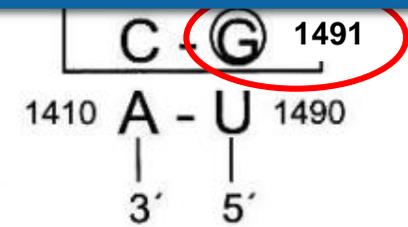
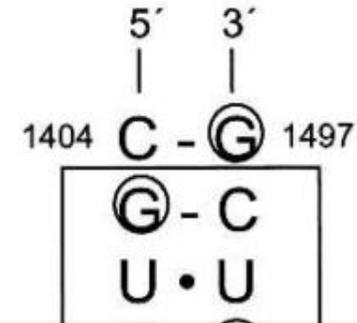
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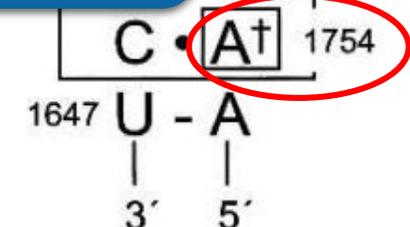
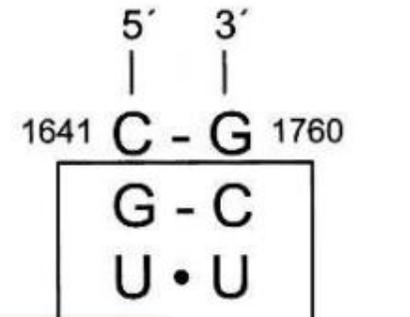
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In eukaryotes aminoglycosides decrease ribosome fidelity and they can be used to induce readthrough

A1408 and G1491 VS G1645 and A1754



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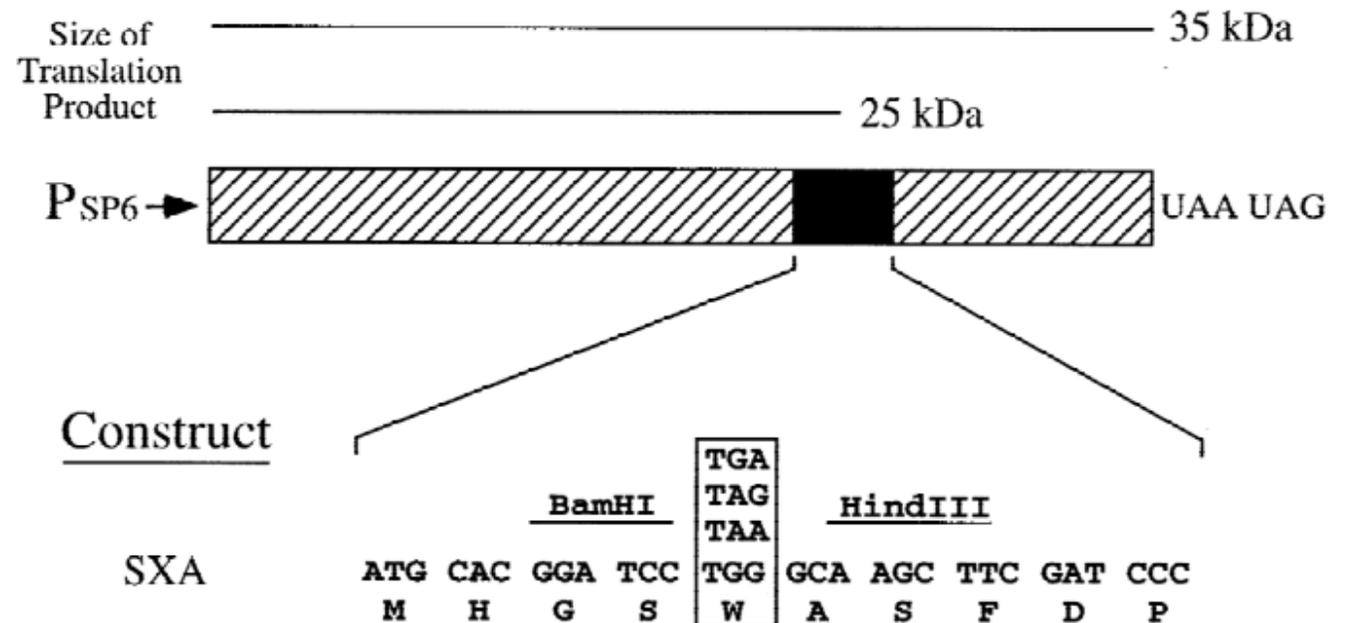
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3) The **nucleotide context** strongly influences efficiency of translation termination and, as a consequence, occurrence of readthrough

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Different stop codons promote translation termination with different efficiencies

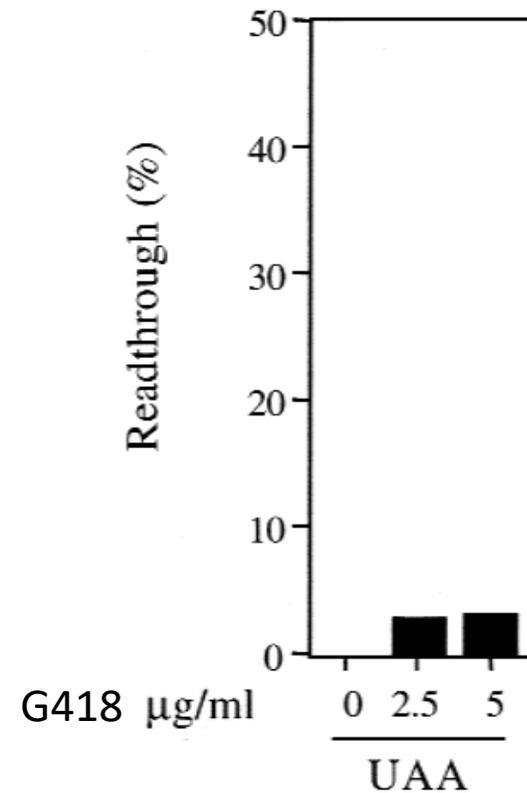
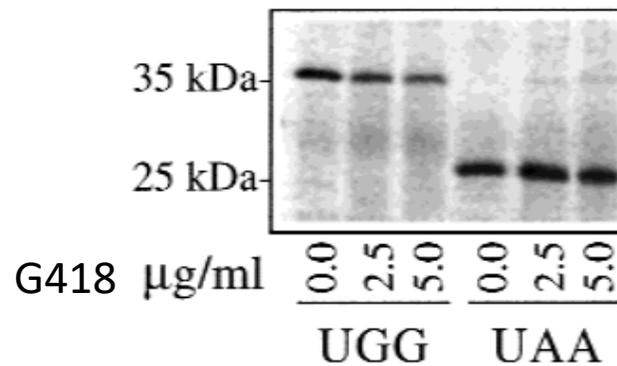
Readthrough reporter system:



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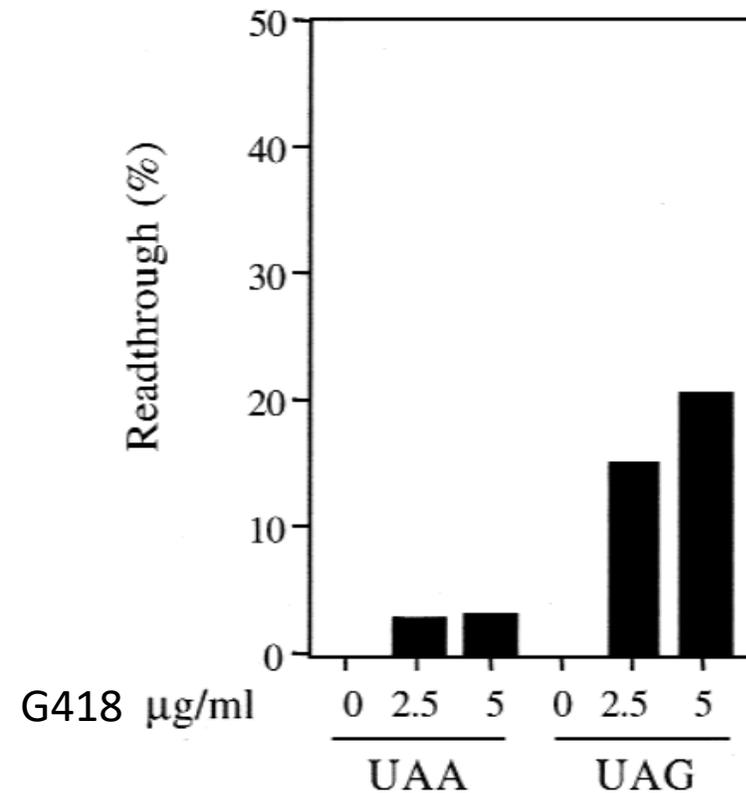
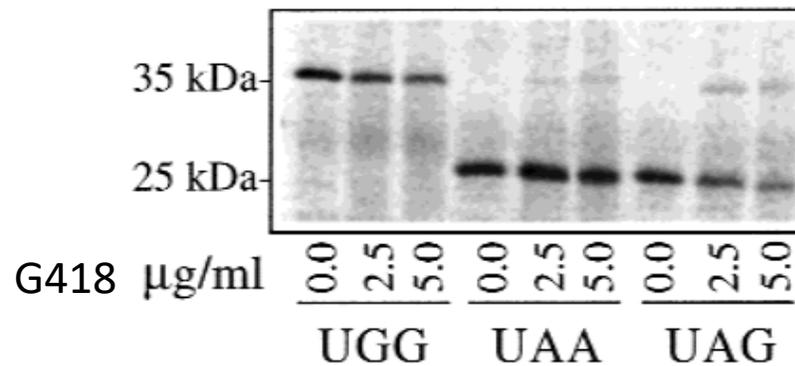
UAA



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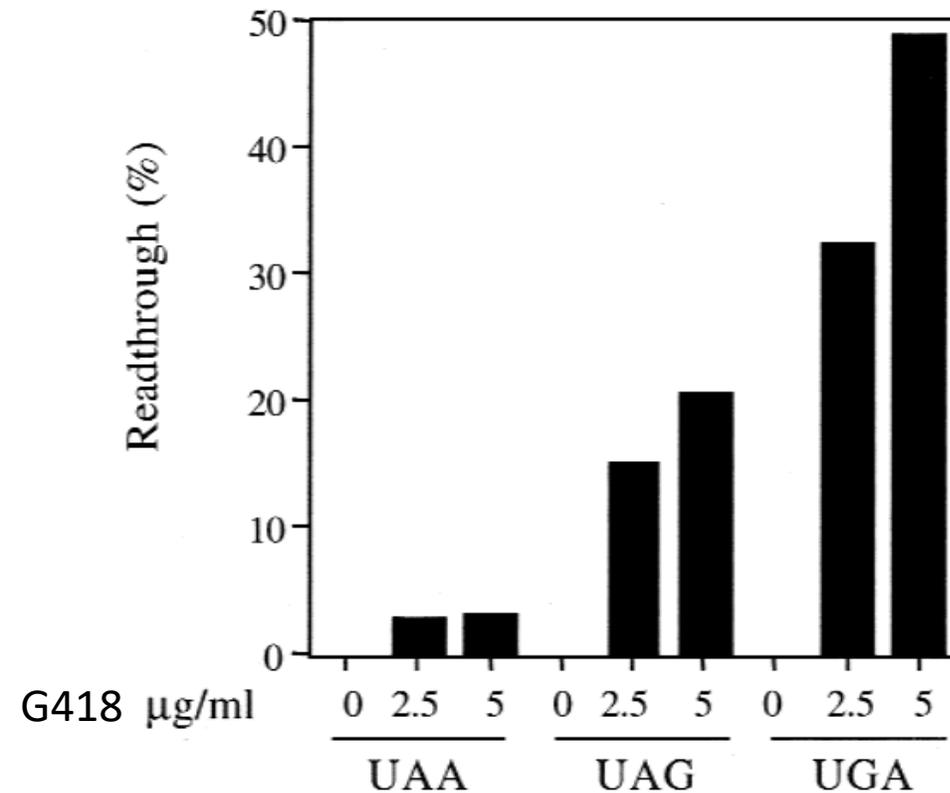
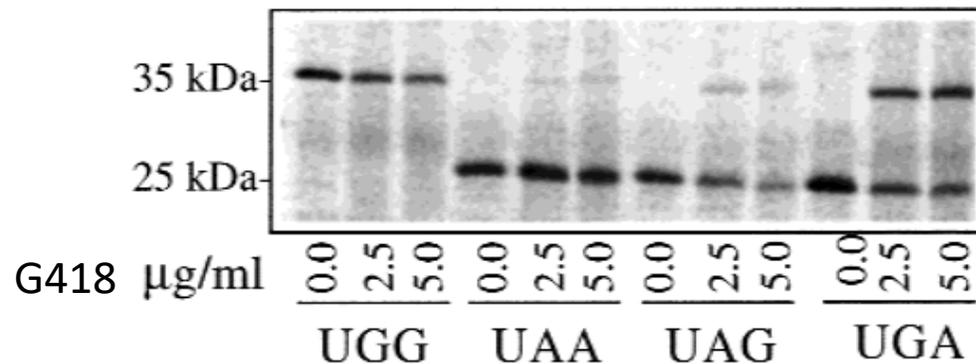
UAA > UAG



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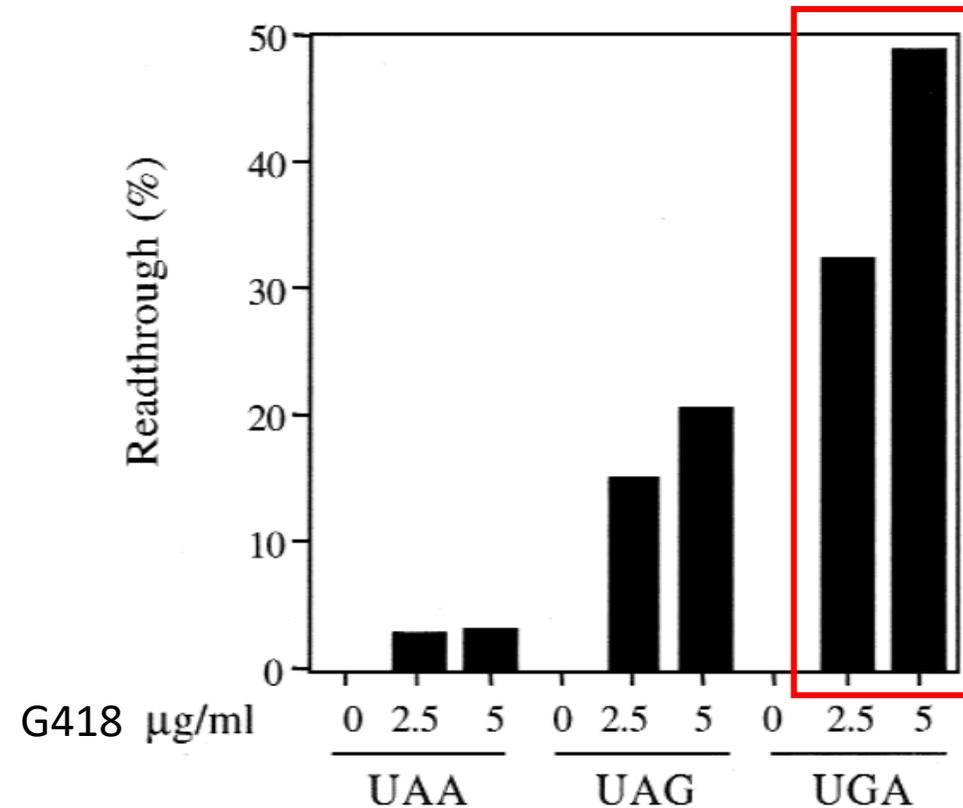
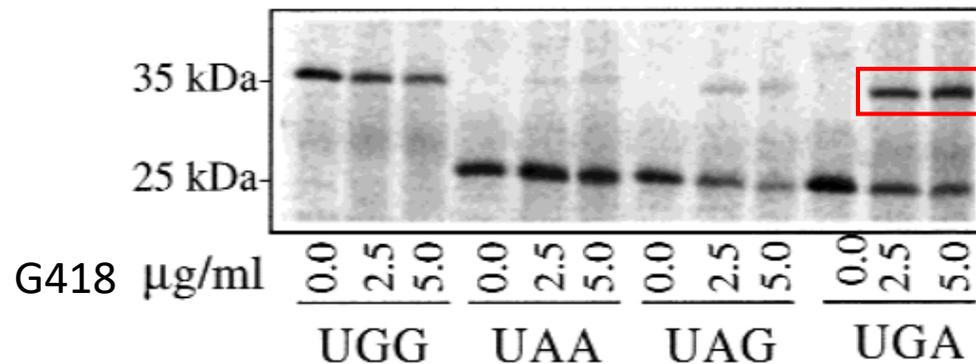
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$UAA > UAG \geq UGA$

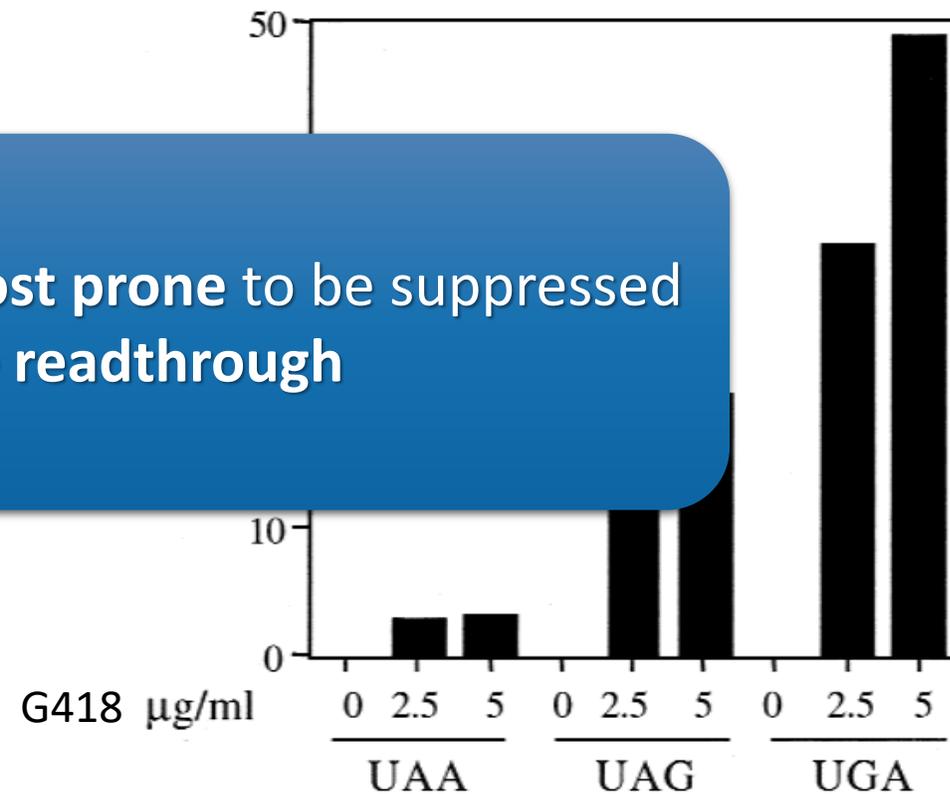
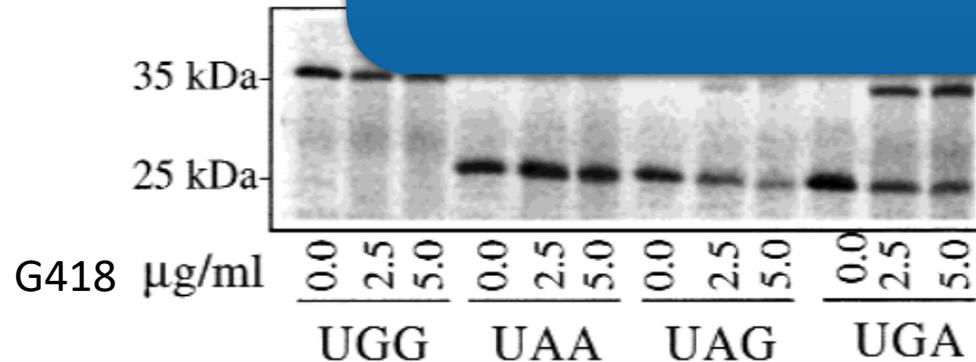


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UAA

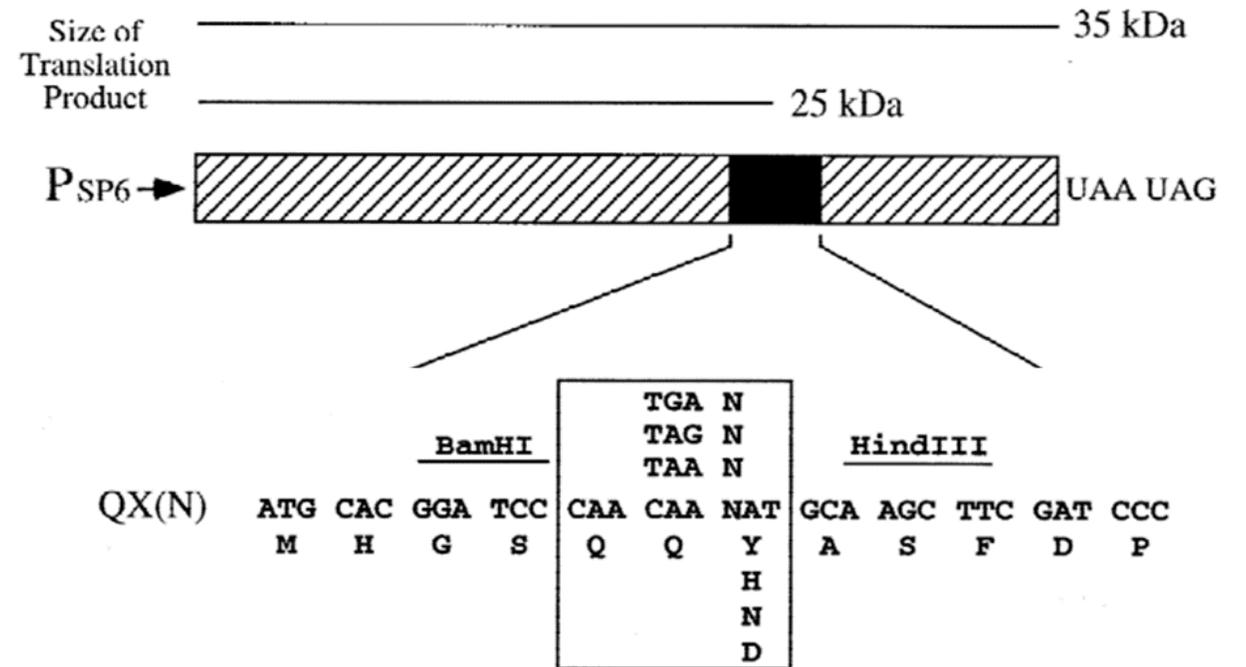
UGA stop codon is the most prone to be suppressed by ribosome readthrough



Also the 4th nucleotide strongly influences the occurrence of readthrough

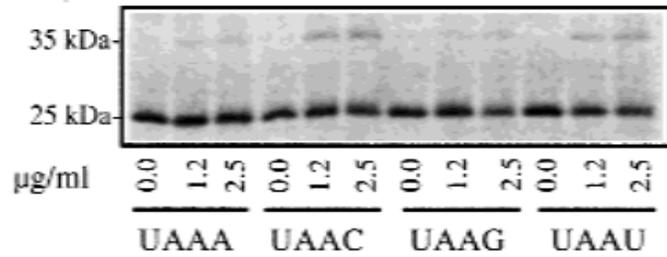
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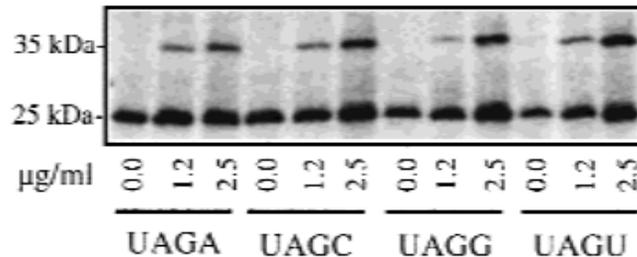


All possible combinations of stop codon and 4th nucleotide

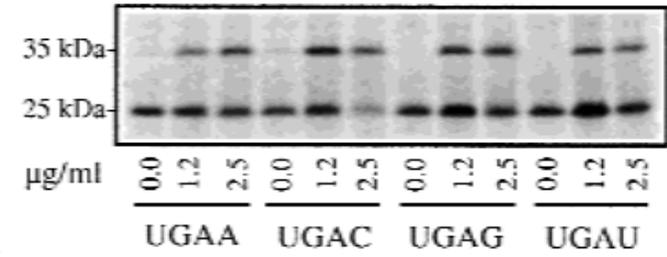
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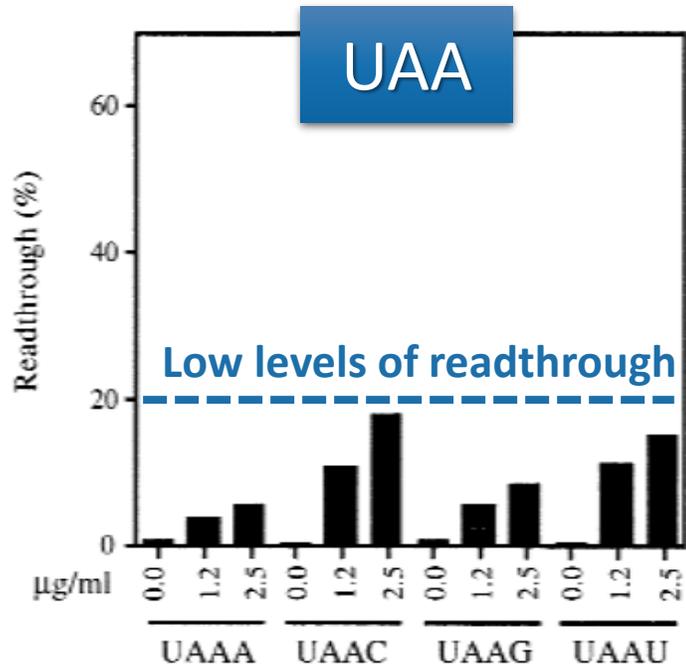
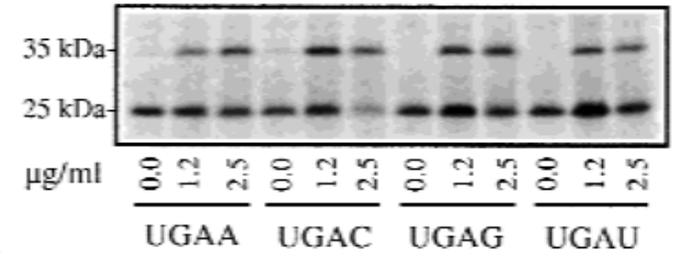
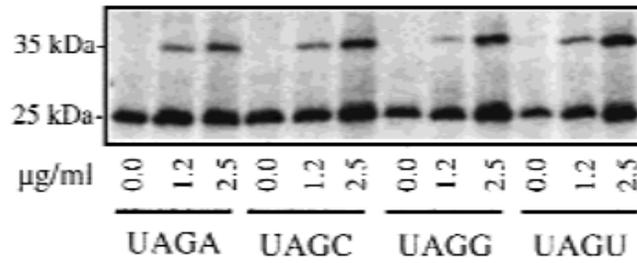
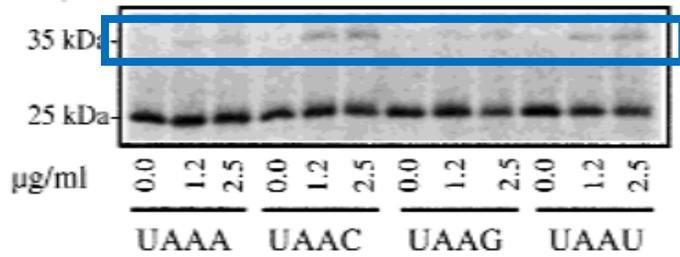
UAG



UGA

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Efficient translation termination

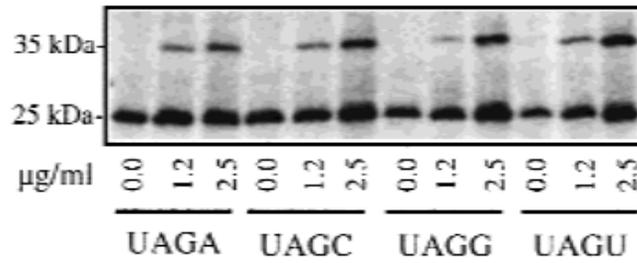
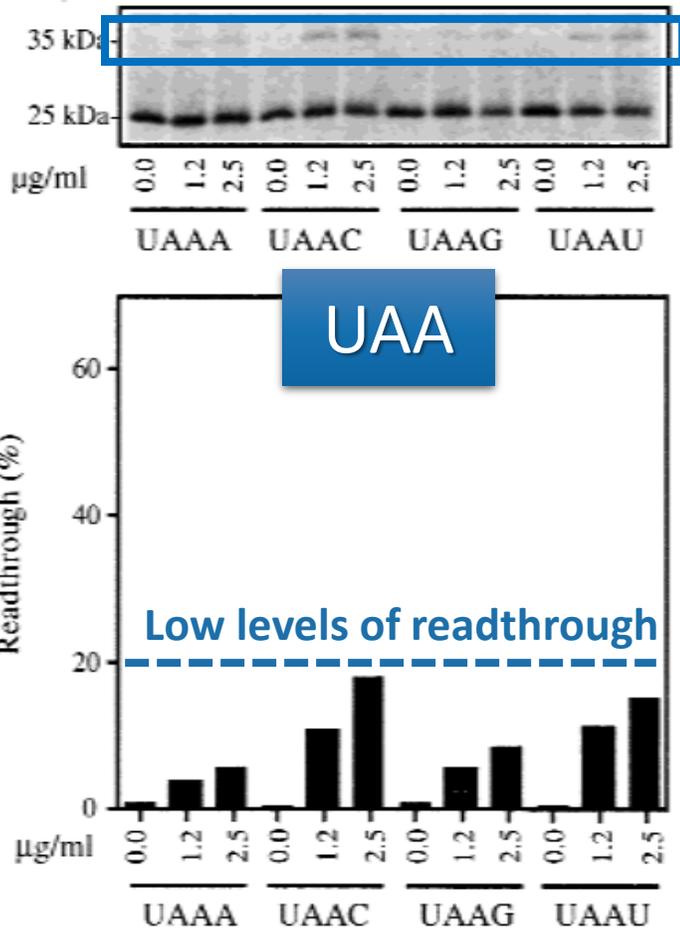


UAG

UGA

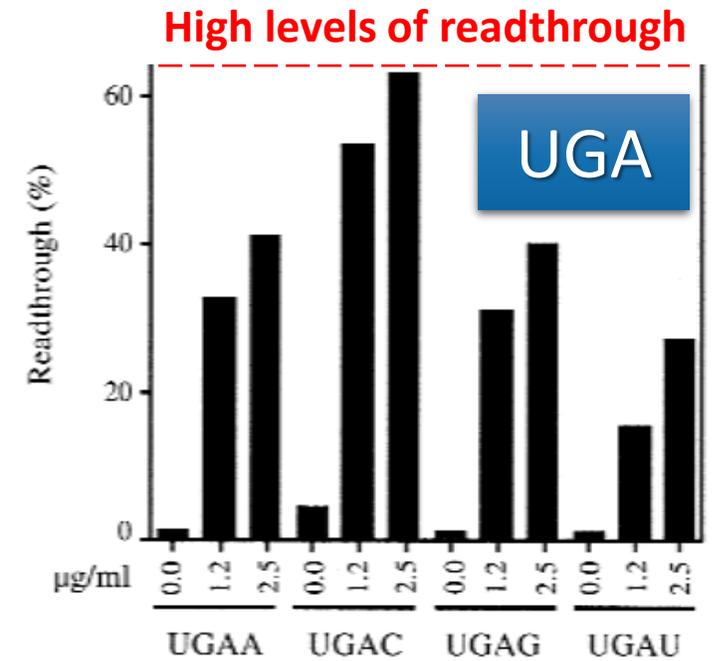
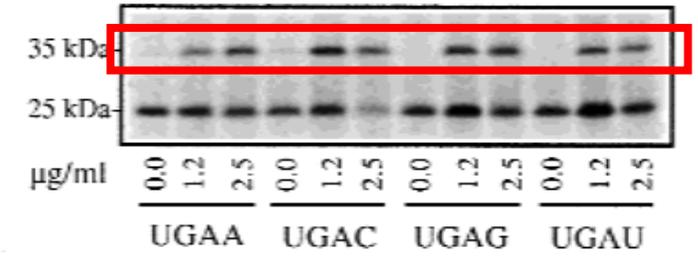
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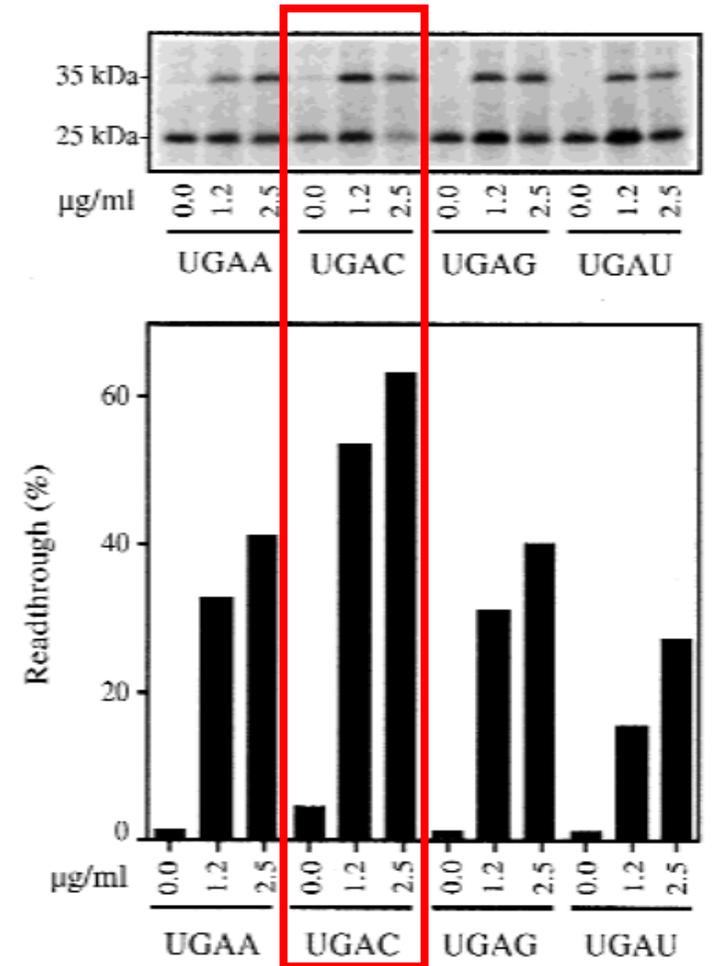
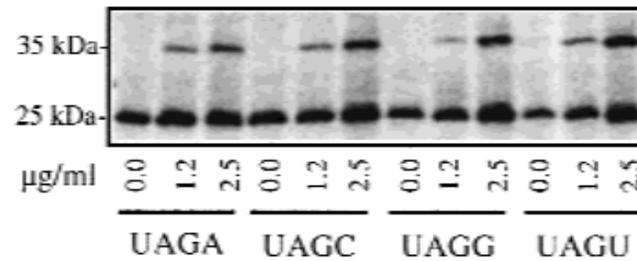
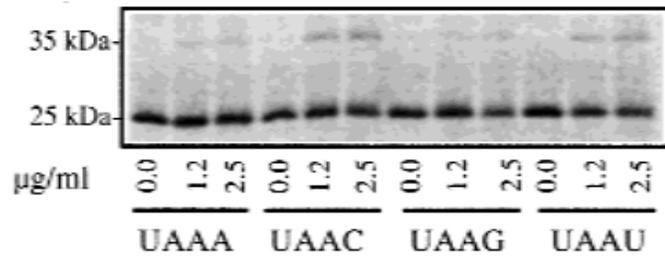


UAG

Less efficient translation termination



Also the 4th nucleotide strongly influences the occurrence of readthrough



The tetranucleotide **UGAC** shows the most frequent stop codon suppression, with readthrough occurring at a frequency of 3–4% (spontaneous) and 63% (G418-induced)

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- 1) The efficiency of termination differs between normal stop codons and premature termination codons
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The restoration of protein activity depends strongly on the identity of the amino acid inserted during PTC readthrough

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doi: 10.1093/nar/gku663

New insights into the incorporation of natural suppressor tRNAs at stop codons in *Saccharomyces cerevisiae*

Sandra Blanchet¹, David Cornu², Manuela Argentini² and Olivier Namy^{1,3,*}

Development of an *in vivo* reporter system to study amino acid insertion at all stop codons

The restoration of protein activity depends strongly on the identity of the amino acid inserted during PTC readthrough

Results

- The main determinant of amino acid incorporation is the sequence of the stop codon

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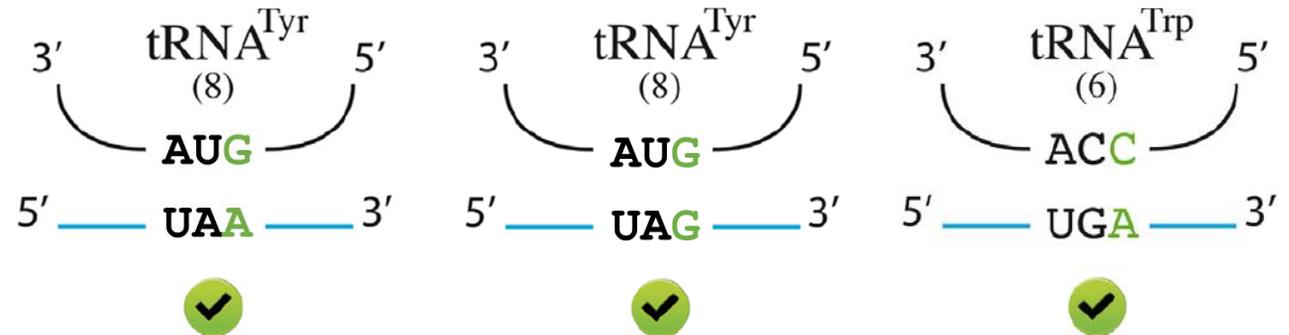
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Tyrosine, glutamine and **lysine** can be inserted at **UAA** and **UAG** codons

Tryptophan, cysteine and **arginine** can be inserted at **UGA** codons



Near-cognate tRNAs: different in only one position of the codon-anticodon pairing

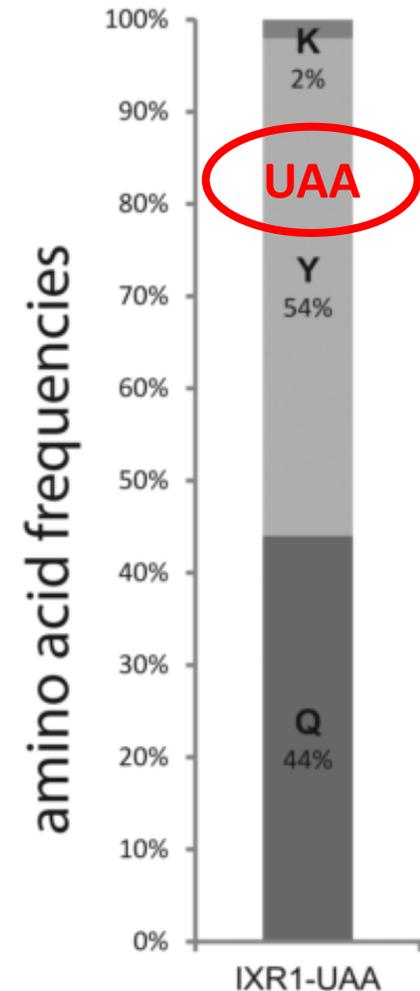
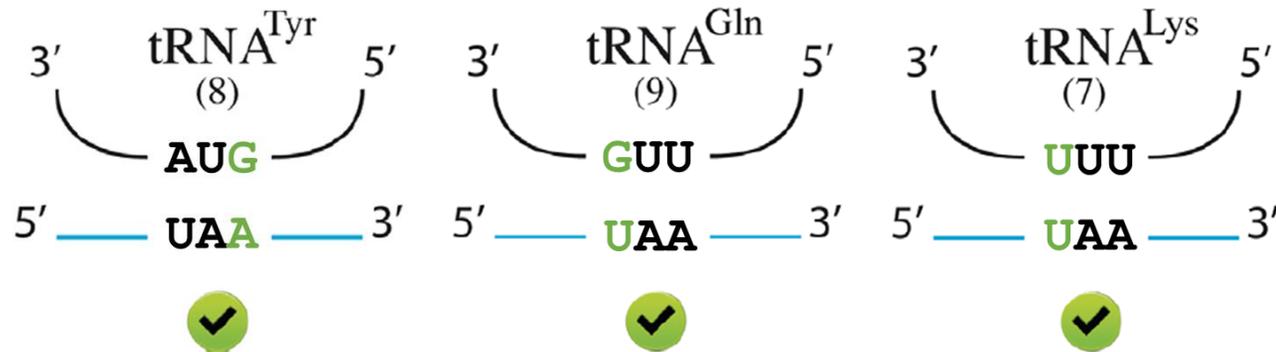
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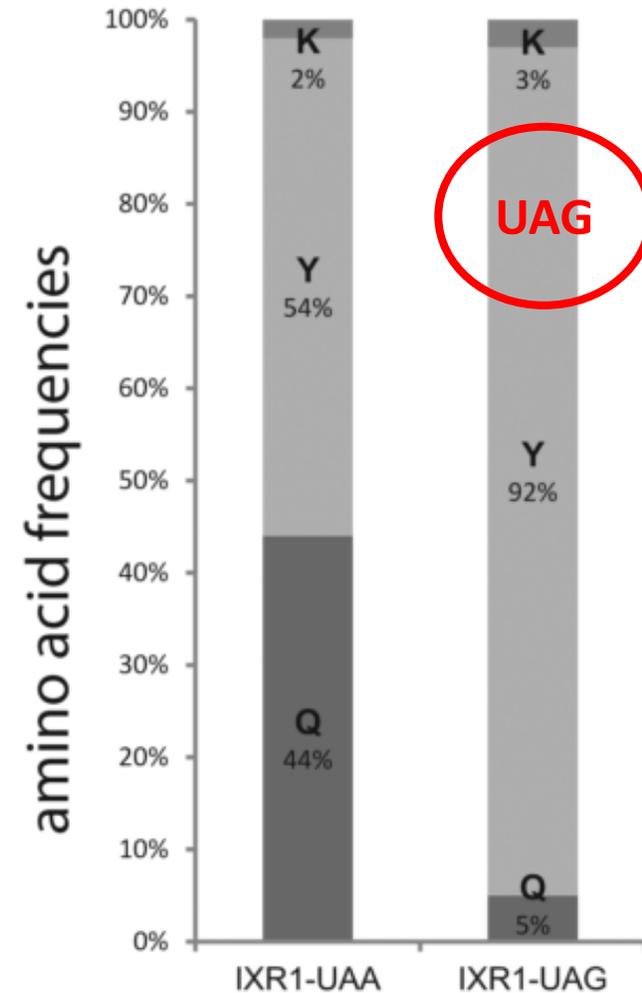
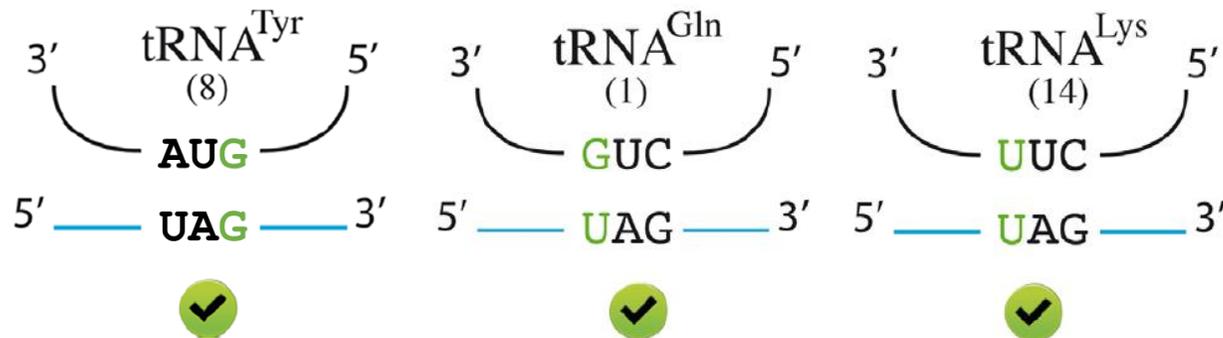
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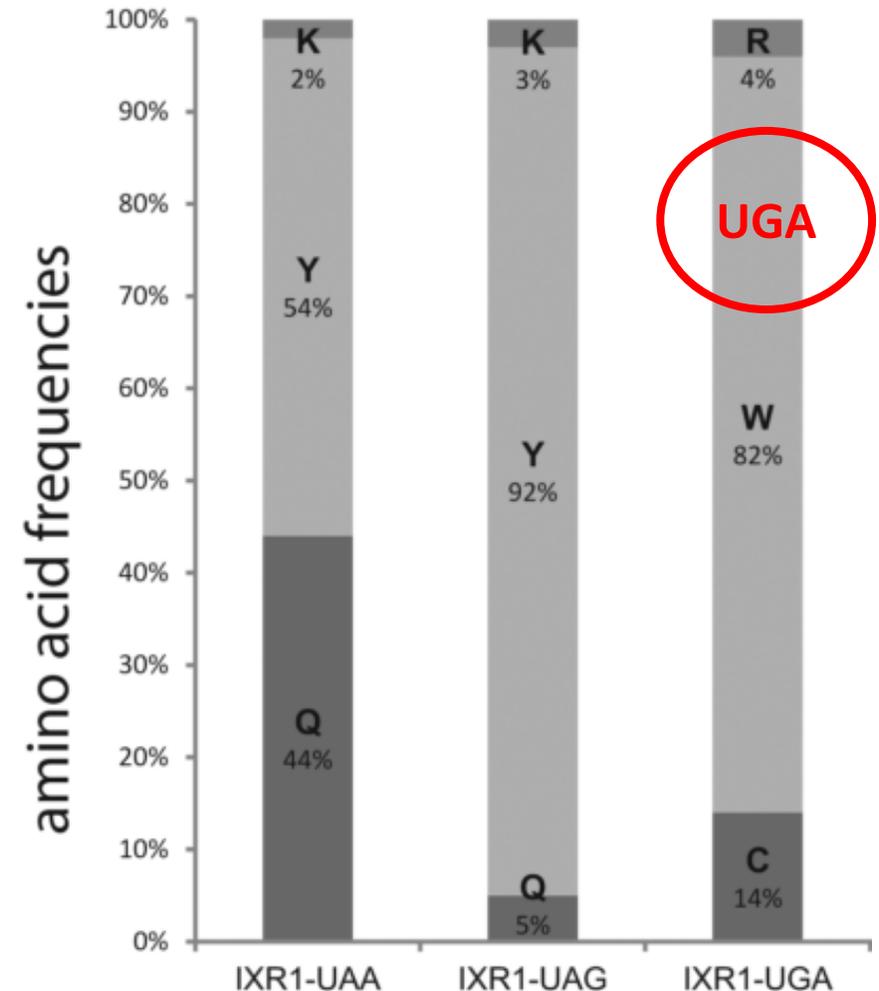
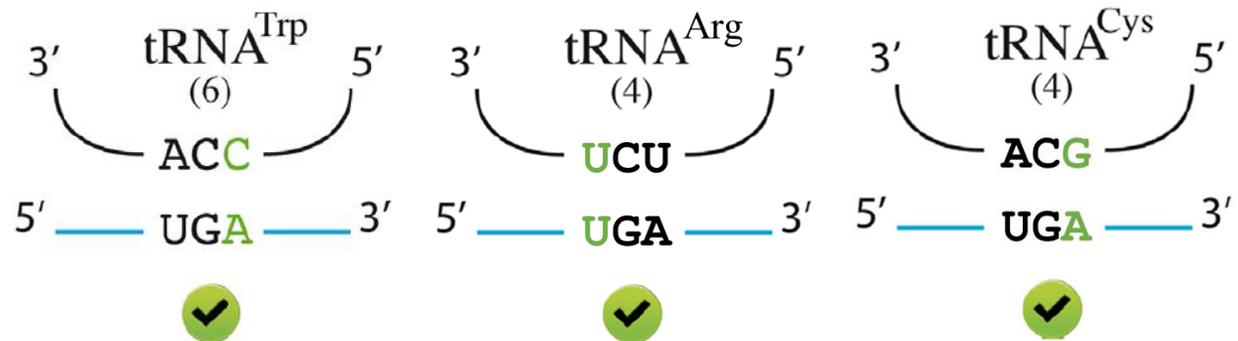
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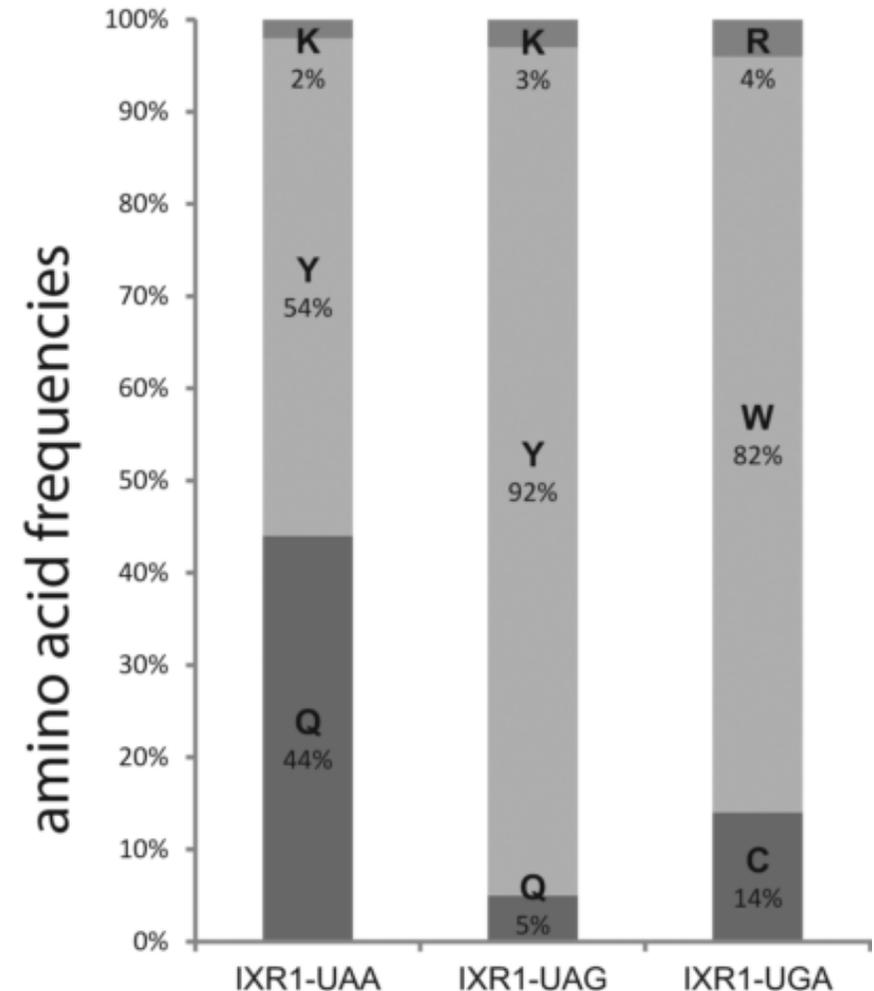
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The restoration of protein activity depends strongly on the identity of the amino acid inserted during PTC readthrough

- Suppressor tRNAs are not incorporated at the same frequency at each stop codon

It is possible to predict the probable protein sequences arising from a readthrough event on the basis of the stop codon present



Brief overview...

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BRIEF REPORT

Differential functional readthrough over homozygous nonsense mutations contributes to the bleeding phenotype in coagulation factor VII deficiency

A. BRANCHINI,^{*†} M. FERRARESE,^{*} S. LOMBARDI,^{*} R. MARI,[‡] F. BERNARDI^{*†} and M. PINOTTI^{*†}

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Objective

Evaluate the spontaneous and drug-induced readthrough levels of two nonsense mutations in coagulation factor VII (FVII) : **p.Ser112X** and **p.Cys132X**

Expected to be lethal

BRIEF REPORT

Differential functional readthrough over homozygous nonsense mutations contributes to the bleeding phenotype in coagulation factor VII deficiency

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Life-threatening
bleeding
symptoms

Moderate
bleeding
symptoms

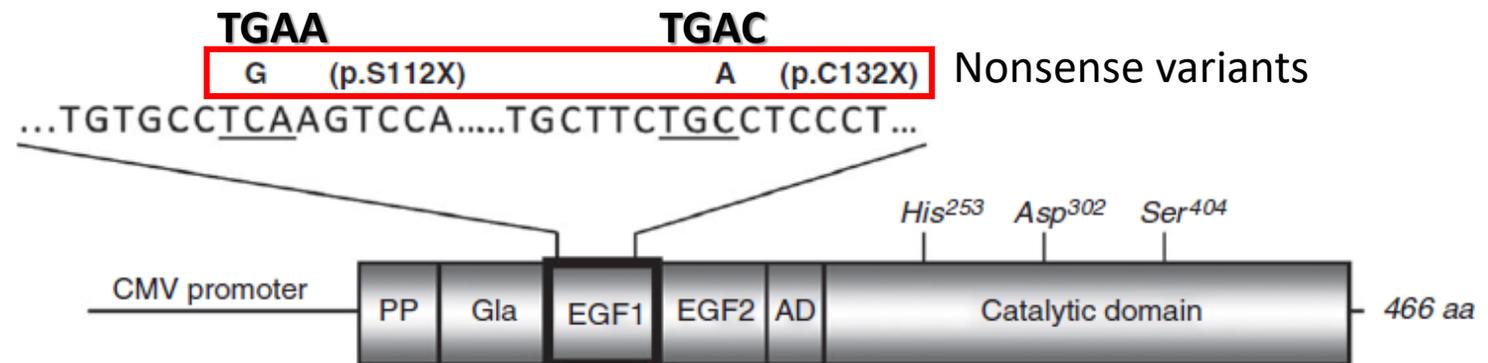
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Methods

1. Creation of a cellular model through the transient expression of recombinant FVII **nonsense** variants
2. Evaluation of secreted levels of rFVII by ELISA



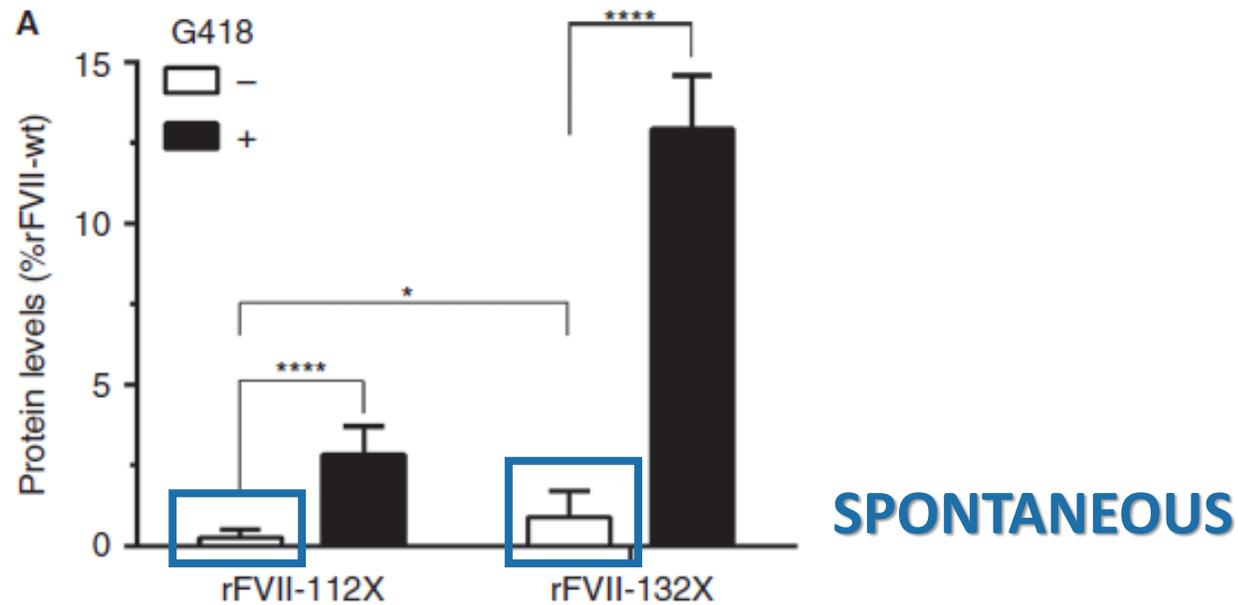
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Results

Secretion levels of rFVII nonsense variants



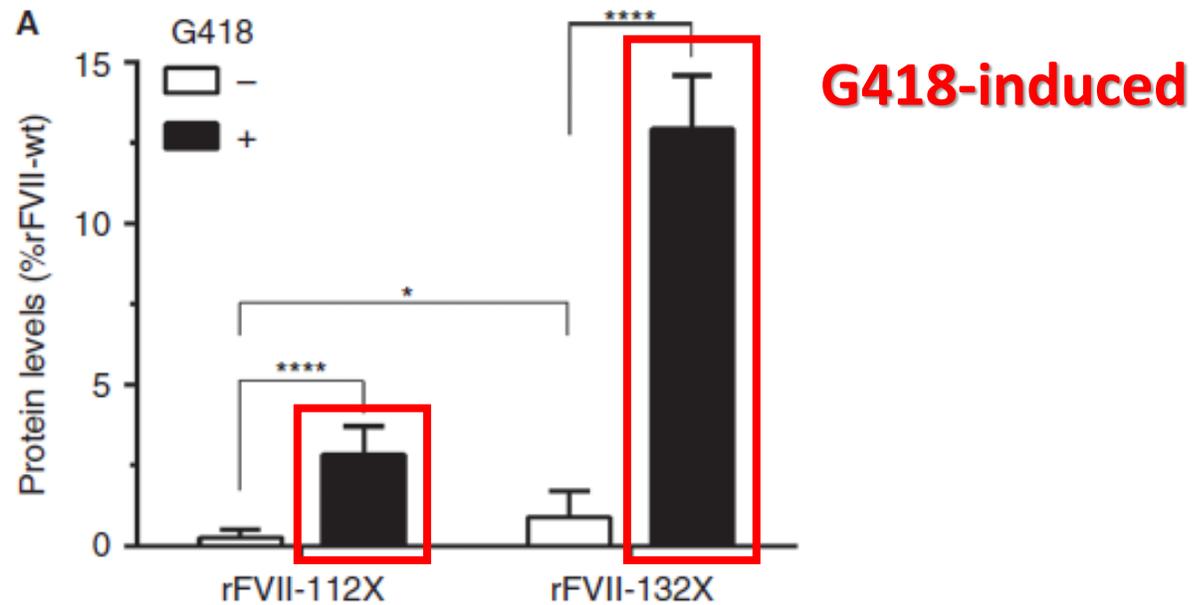
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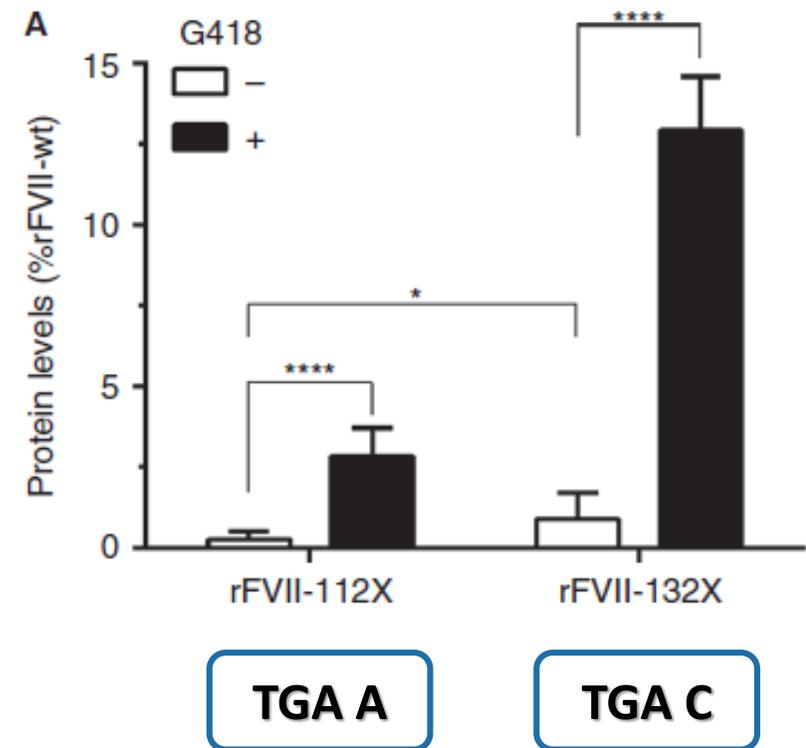
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Secretion and activity levels of rFVII nonsense variants

rFVII C132X shows the higher degree of suppression, probably due to the more readthrough-favourable sequence context



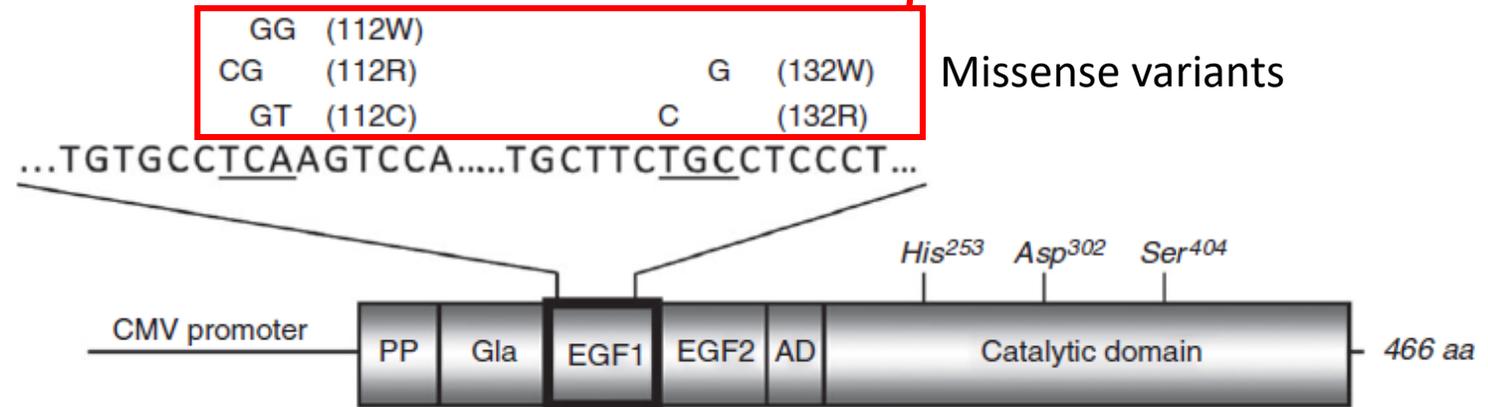
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Methods

1. Creation of a cellular model through the transient expression of recombinant FVII **missense** variants
2. Evaluation of rFVII protein and **functional** levels by ELISA and activity assays



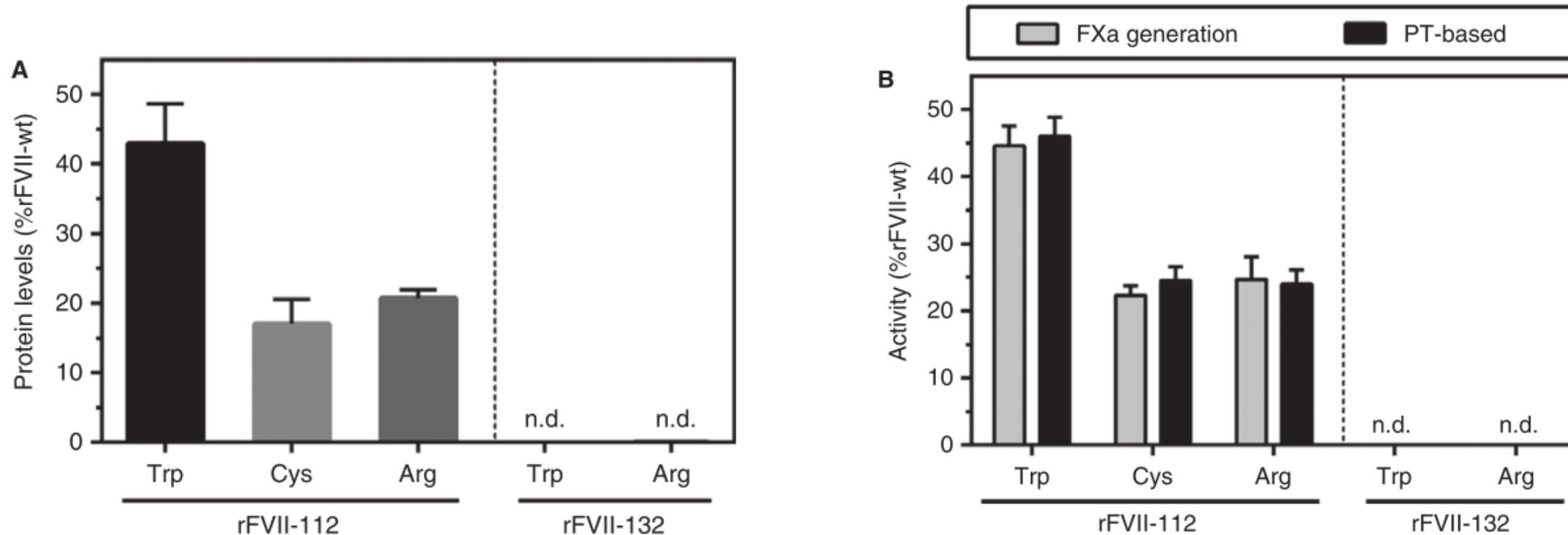
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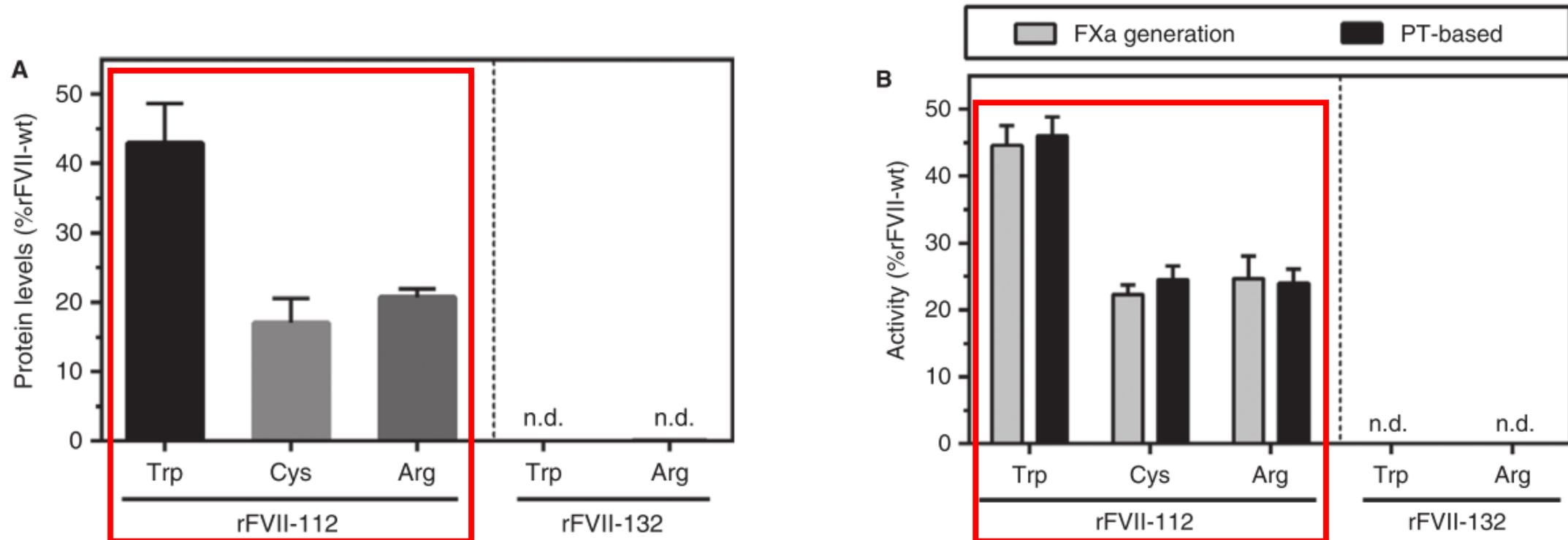
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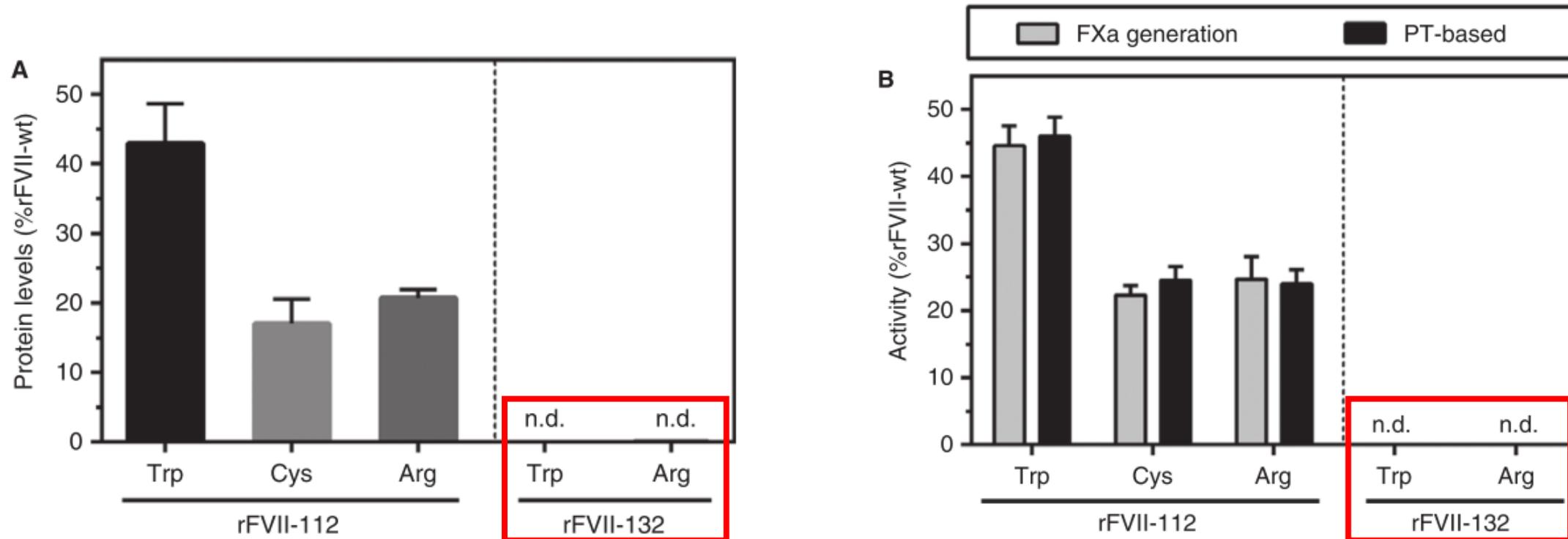
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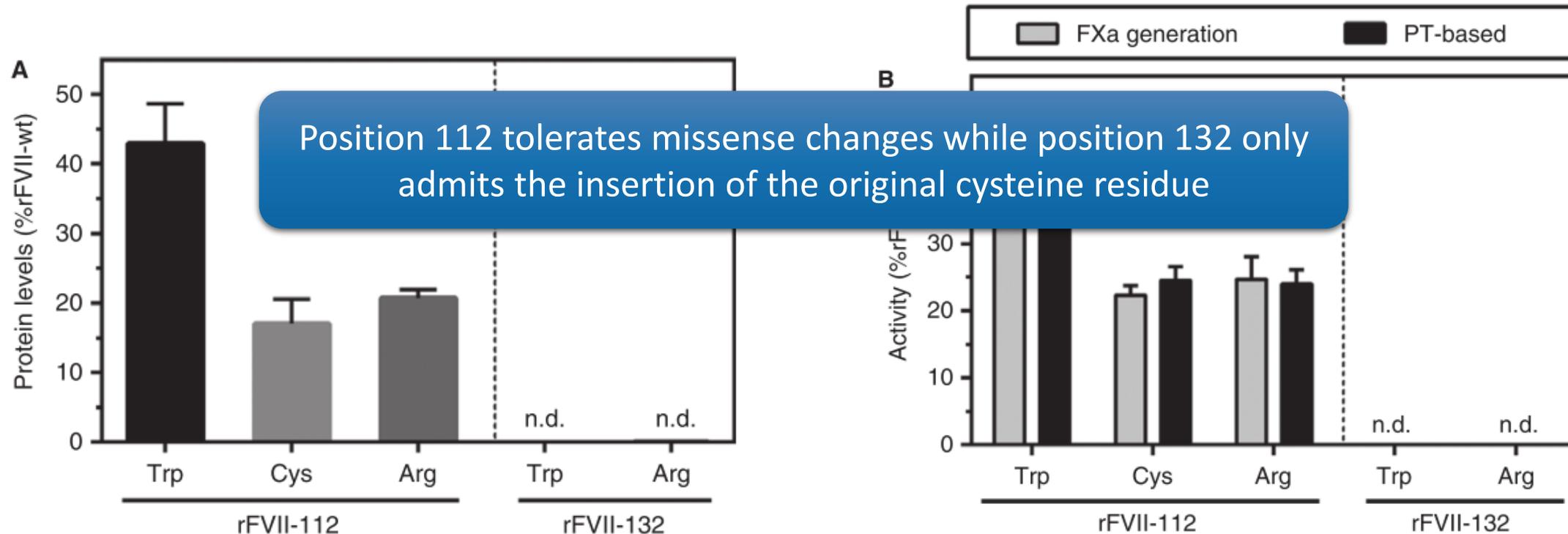
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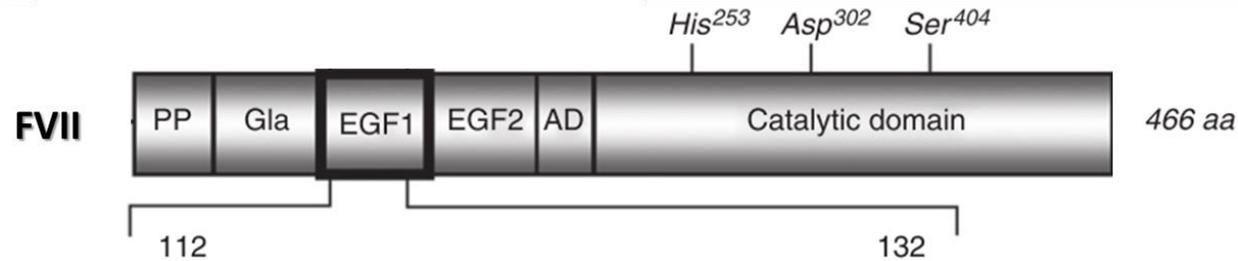
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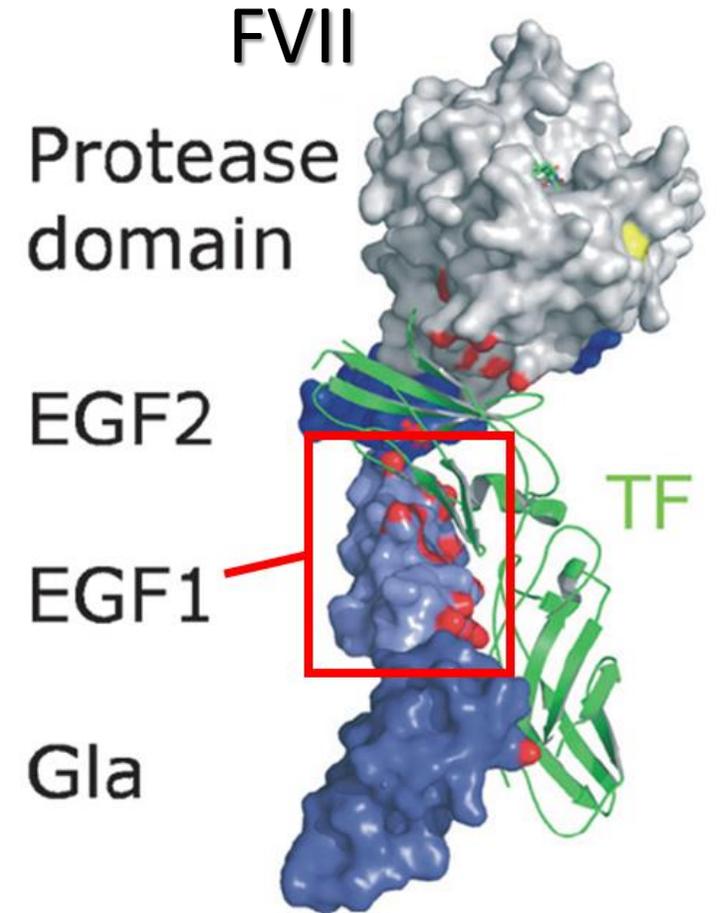
Secretion and activity levels of rFVII missense variants



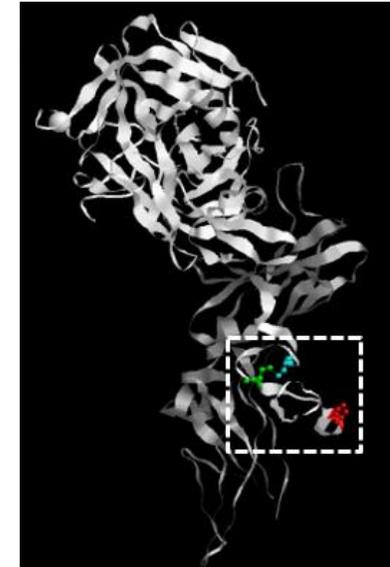
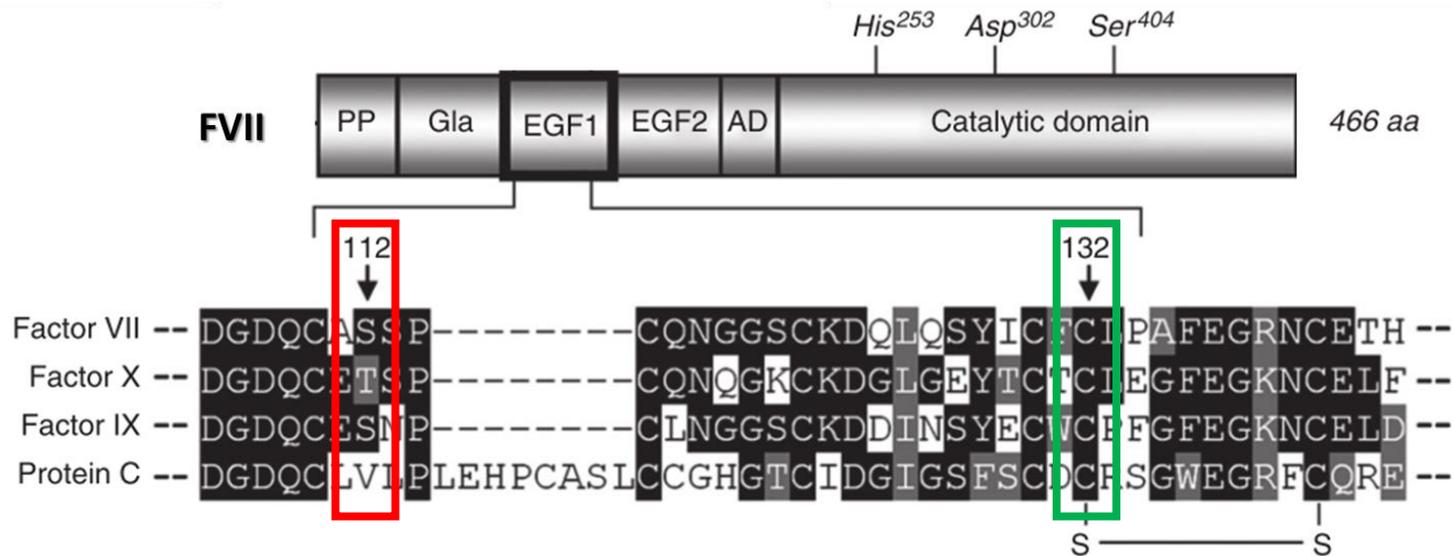
Position 112 tolerates missense changes while position 132 only admits the insertion of the original cysteine residue



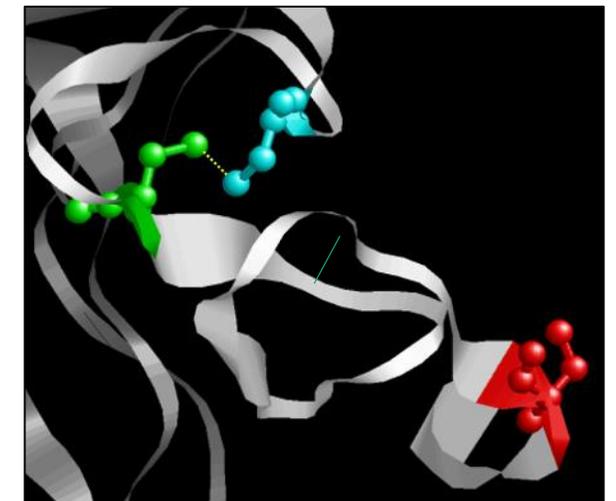
Both residues are part of the first EGF domain of FVII, which is involved in the interaction with tissue factor (TF, the FVII cofactor)



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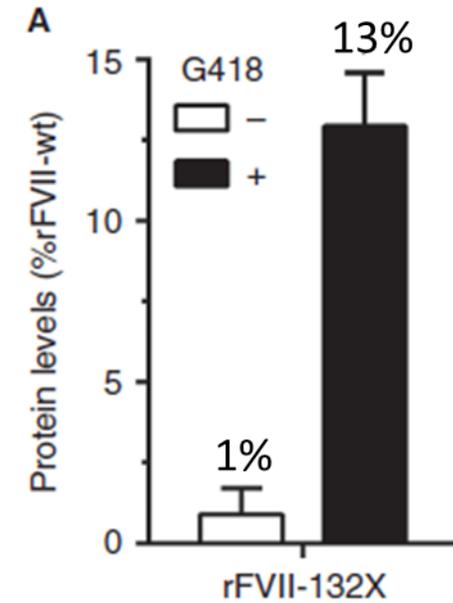
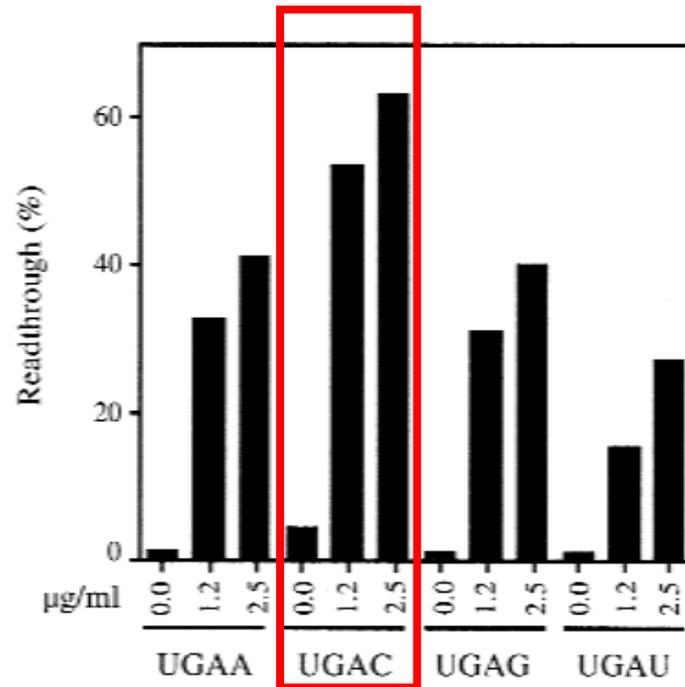


But whereas **Ser112** is surface-exposed and only partially conserved among others coagulation factors, **Cys132** forms a disulfide bridge with **Cys141** within EGF1 and is fully conserved.



In conclusion:

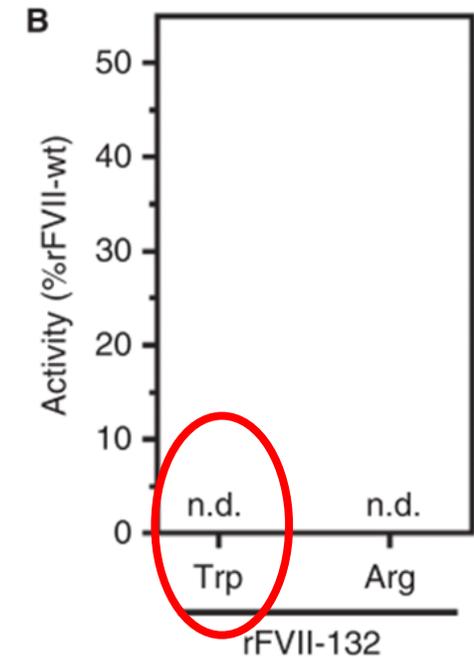
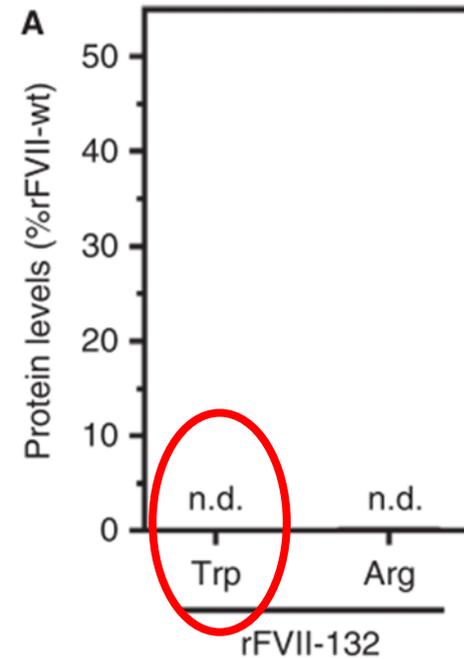
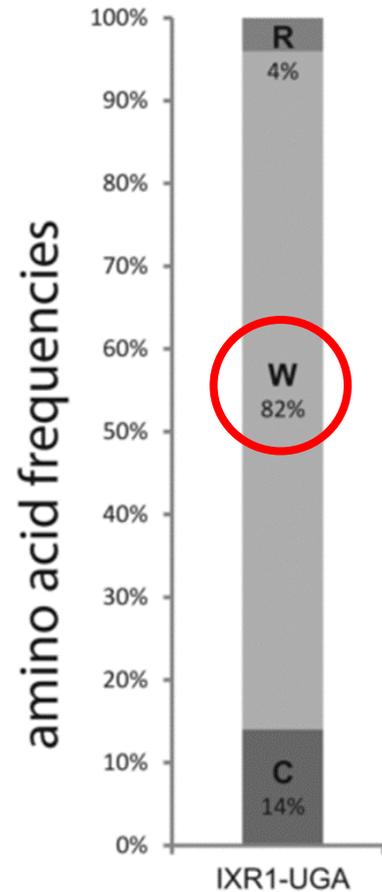
- **p.C132X** variant shows the **most readthrough favourable sequence context**



Moderate bleeding phenotype

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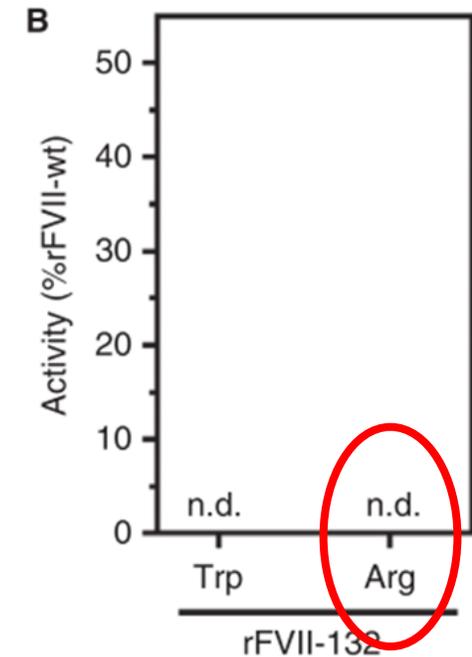
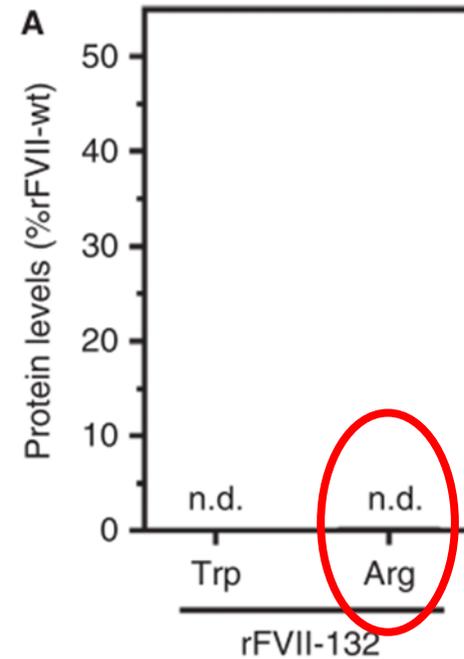
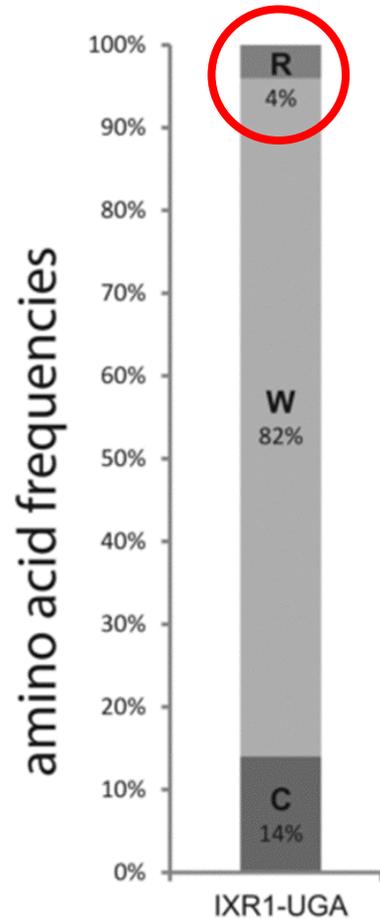
- **p.C132X** variant shows the most readthrough favourable sequence context
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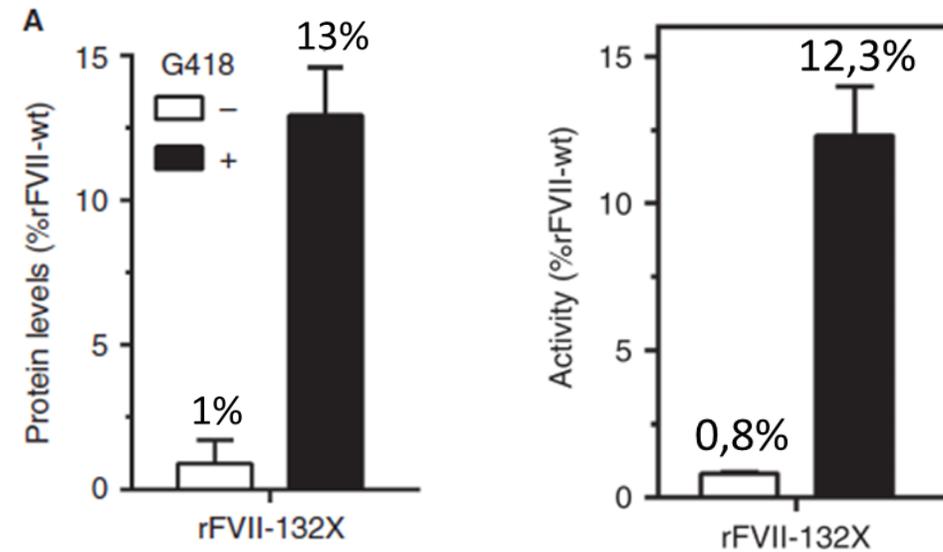
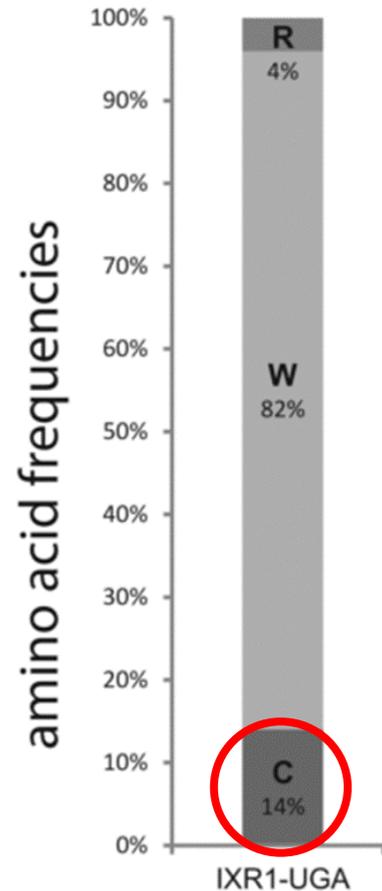
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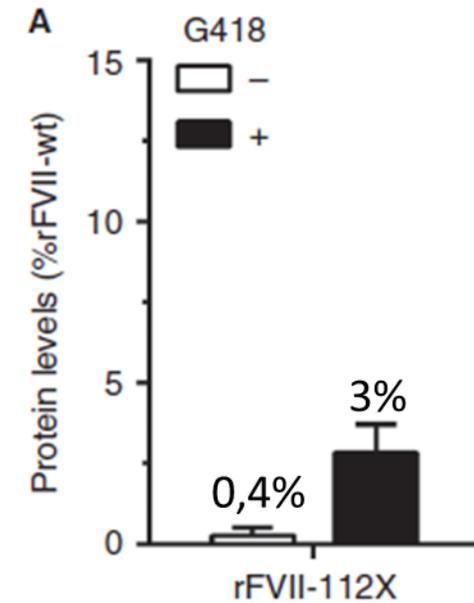
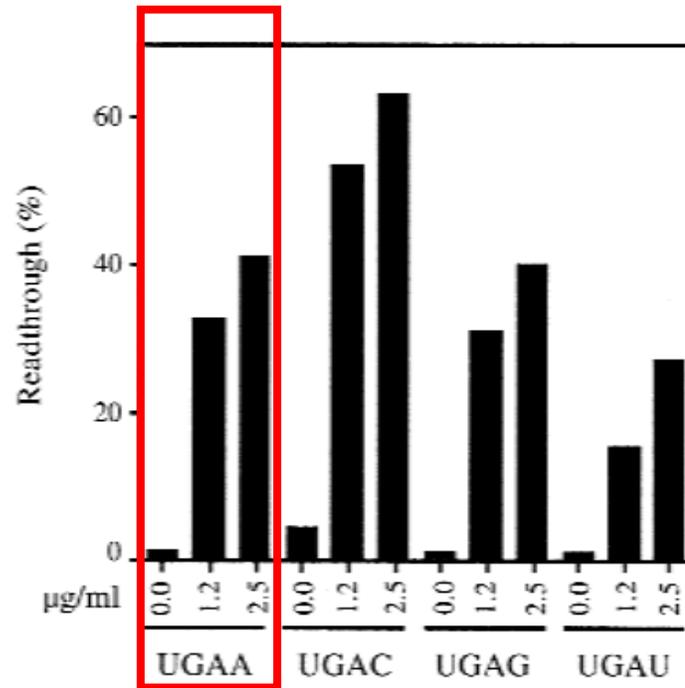


Moderate
bleeding
phenotype

Insertion of original amino acid and
production of full-length wild-type protein

In conclusion:

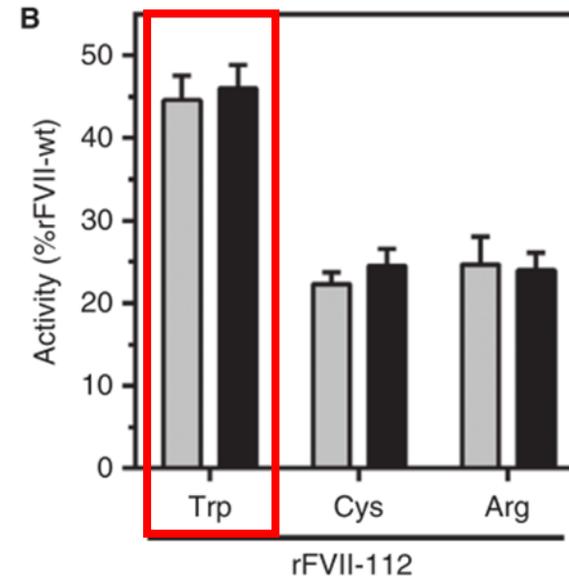
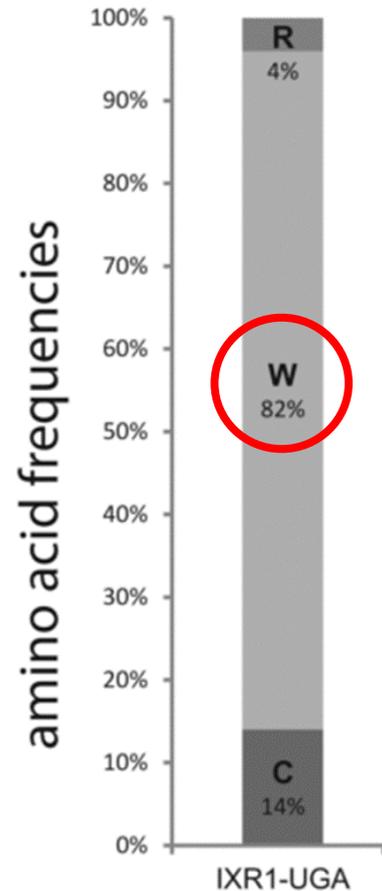
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Life-threatening bleeding phenotype

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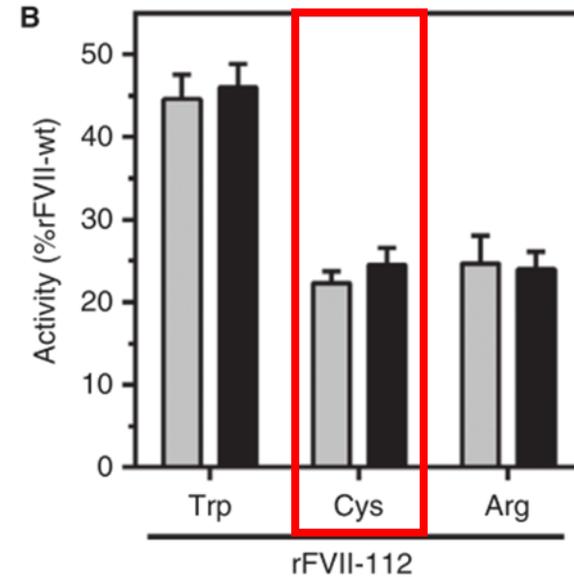
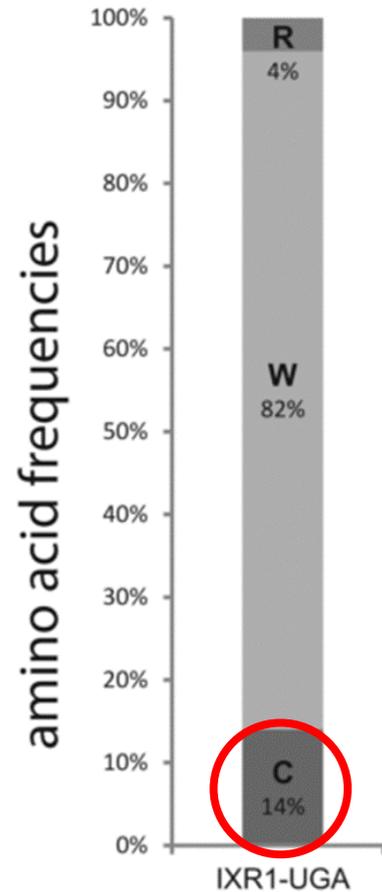
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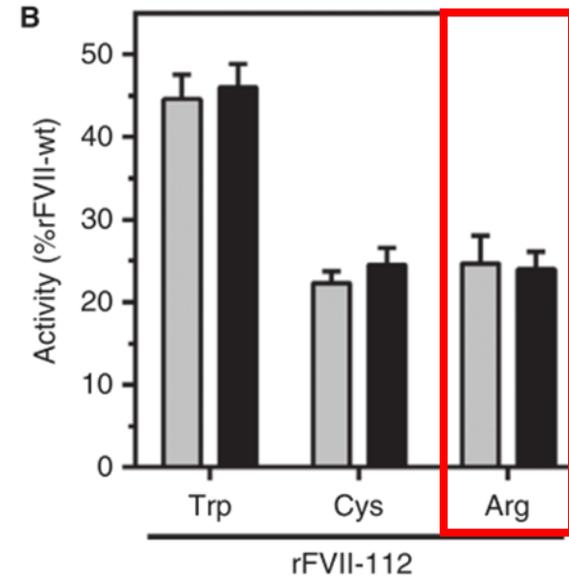
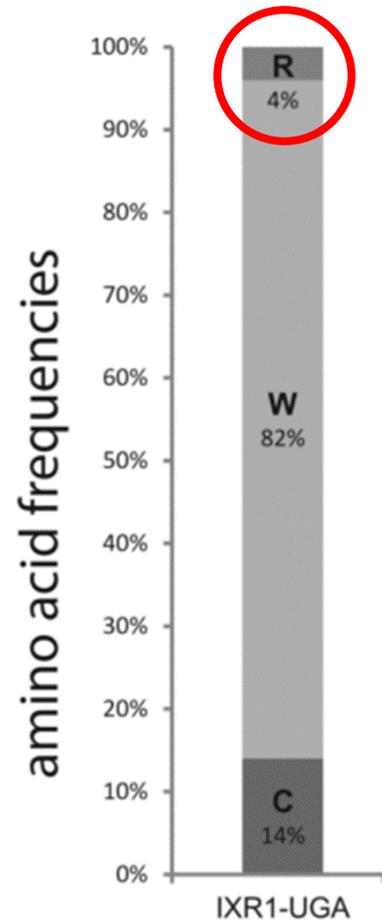
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Life-threatening bleeding phenotype

Experimental findings are consistent with the complex and integrated scenario depicted above and support the notion that many elements account for readthrough efficiency and protein function restoration

Stop codon readthrough is a process that increases the diversity of genomic information.

This mechanism is well documented with respect to viruses, yeast and *Drosophila*, but is believed to occur also in higher animals.

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Received for publication, October 28, 2011, and in revised form, March 22, 2012. Published, JBC Papers in Press, March 28, 2012, DOI 10.1074/jbc.M111.314468

Yoshihide Yamaguchi¹, Akiko Hayashi², Celia W. Campagnoni⁵, Akio Kimura¹, Takashi Inuzuka¹, and Hiroko Baba²



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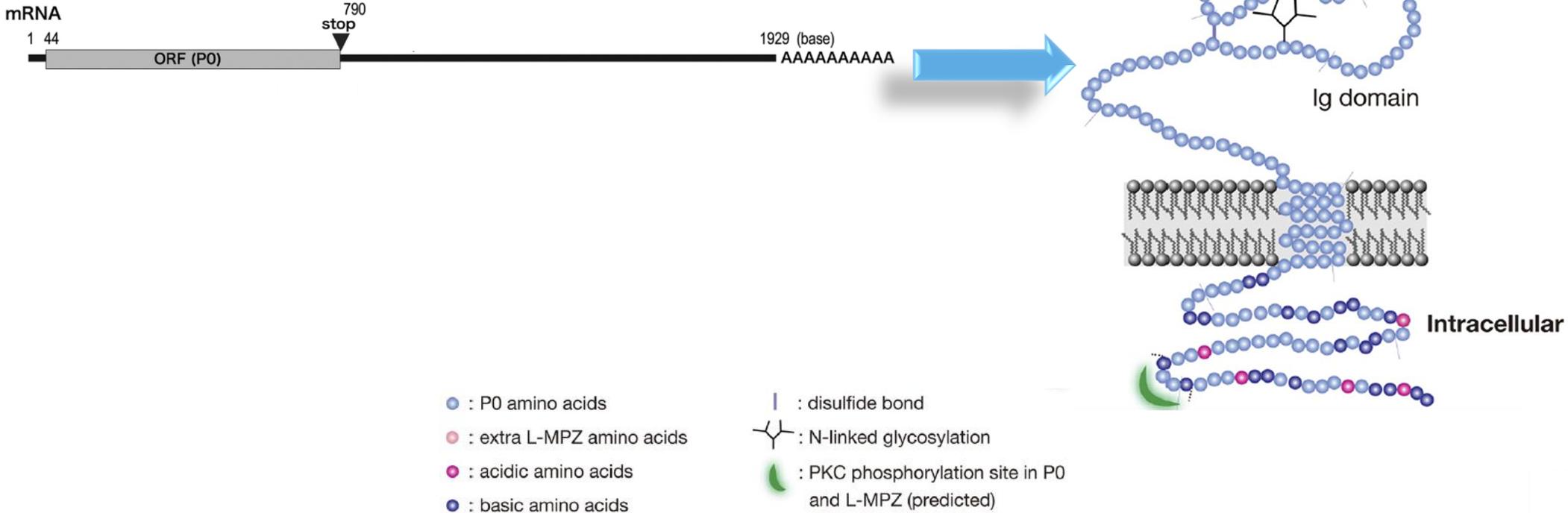
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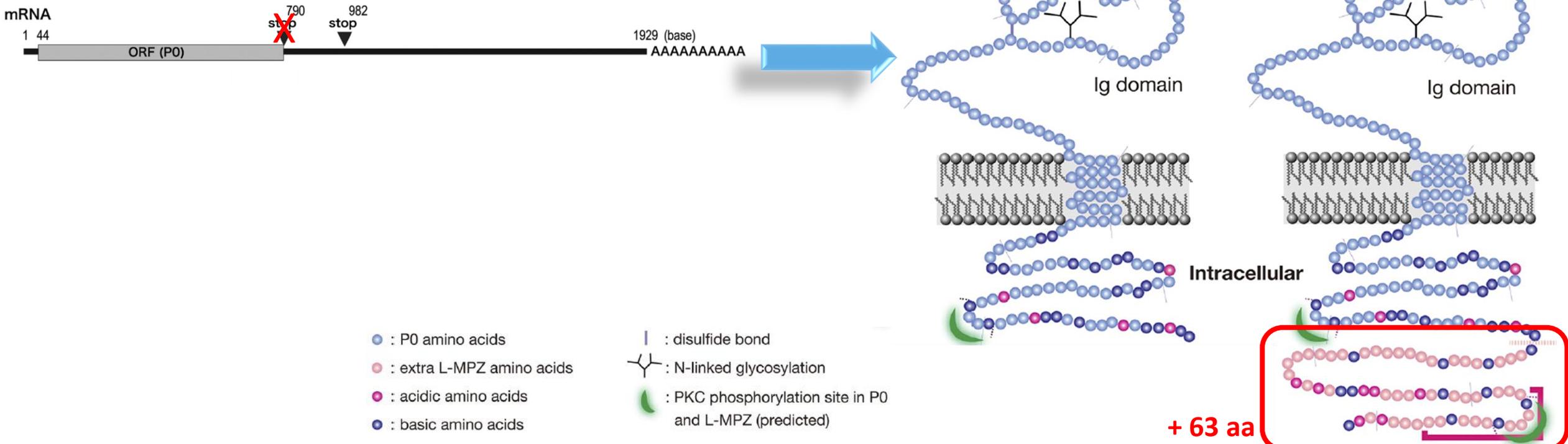
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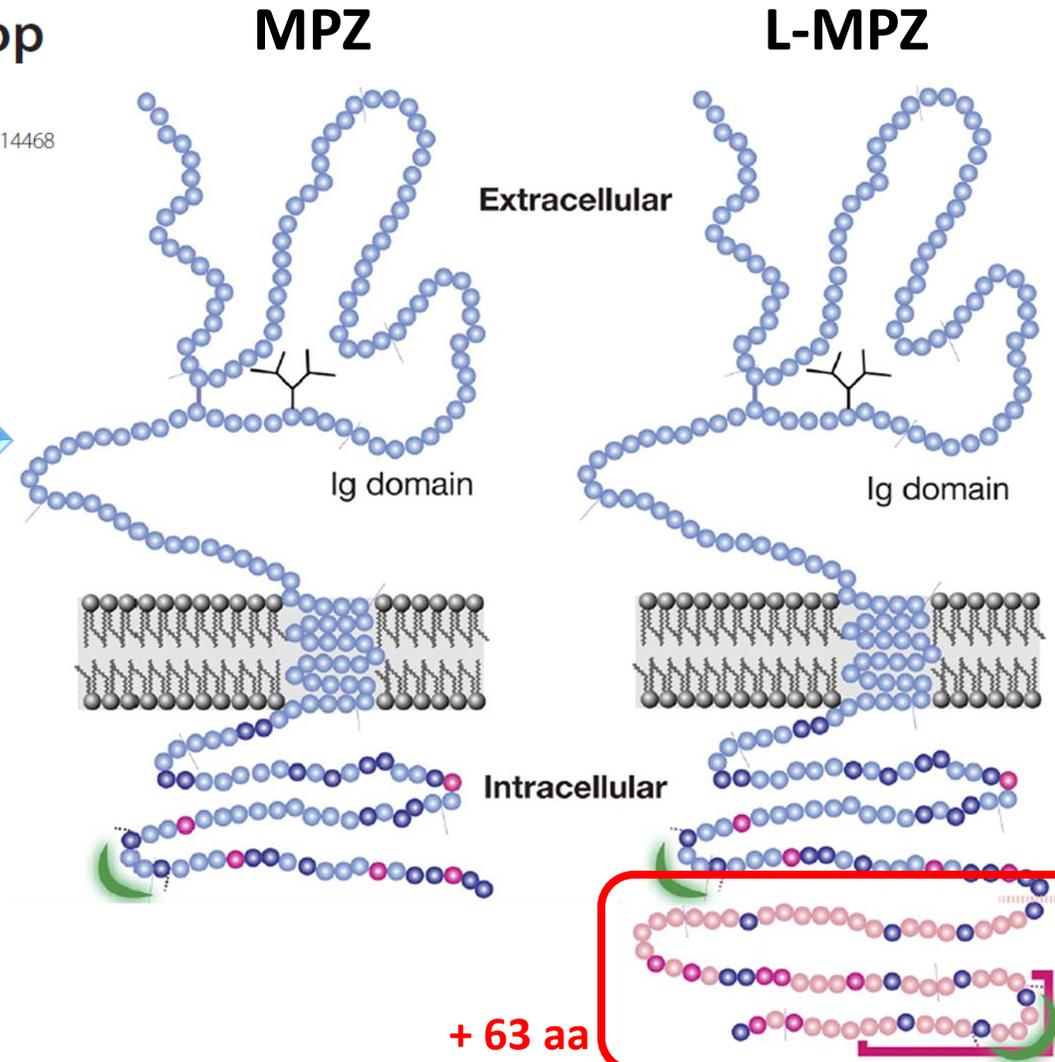
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