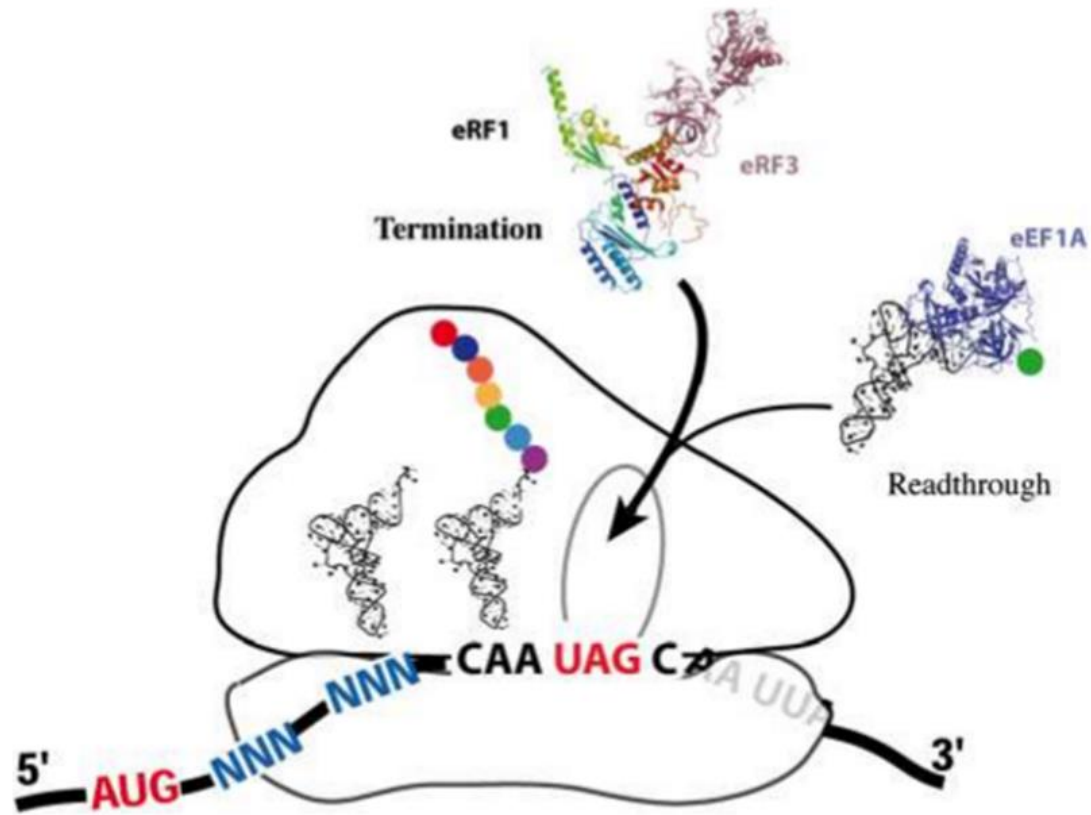


# Ribosome readthrough



# Starting from the base...PROTEIN SYNTHESIS

Eukaryotic translation can be divided into four stages: Initiation, Elongation, Termination and Recycling

During translation, the ribosome catalyzes the sequential addition of amino acids to a growing polypeptide chain, using an mRNA as template and aminoacyl-tRNAs as substrates

Correct base pairing between the three bases of the codon on mRNA and those of the anticodon of the aa-tRNA dictates the sequence of the polypeptide chain

# Translation termination

When the ribosome arrives at a **stop codon**, there is **no corresponding tRNA**. Instead, a **release factor** enters the assembly site and **synthesis is terminated**, releasing the completed polypeptide from the ribosome.

# Translation termination

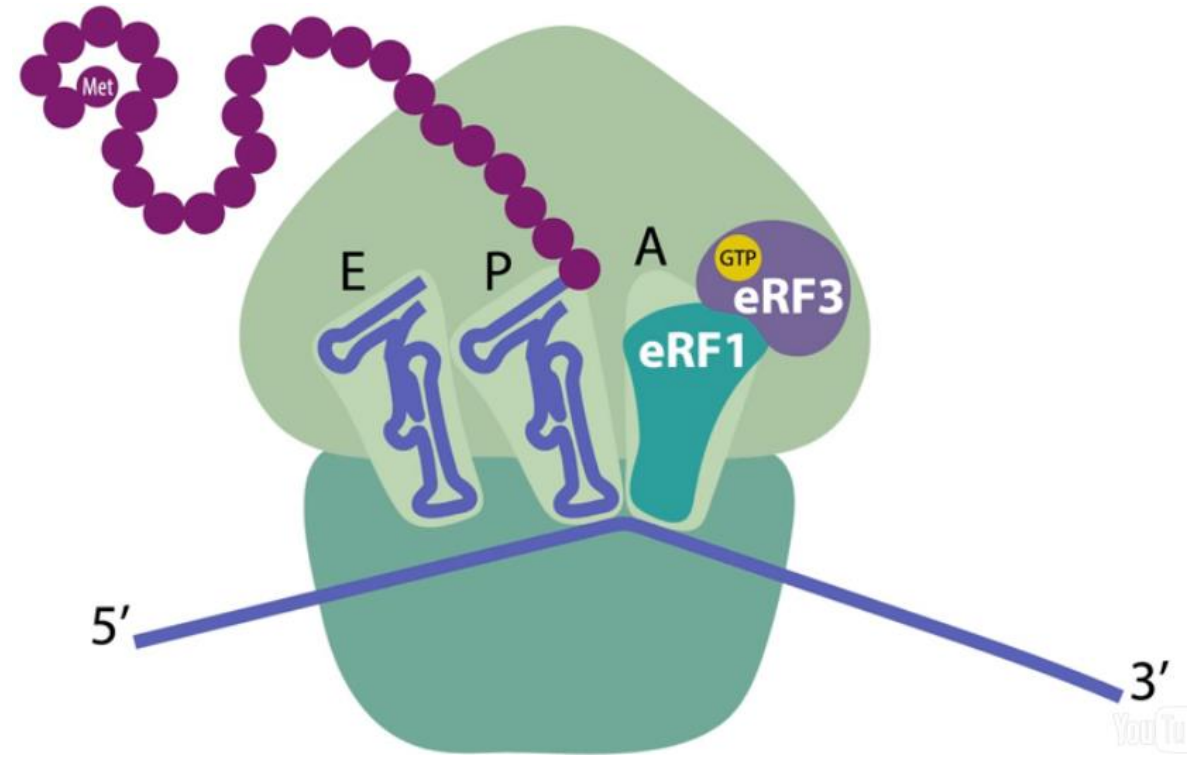
When the ribosome arrives at a **stop codon**, there is **no corresponding tRNA**. Instead, a **release factor** enters the assembly site and **synthesis is terminated**, releasing the completed polypeptide from the ribosome.

Translation termination in eukaryotes is mediated by two release factors:

- **eRF1** recognizes each of the three stop codons (UAG, UAA, and UGA) and facilitates release of the nascent polypeptide chain
- **eRF3** is a GTP binding protein that facilitate the termination process

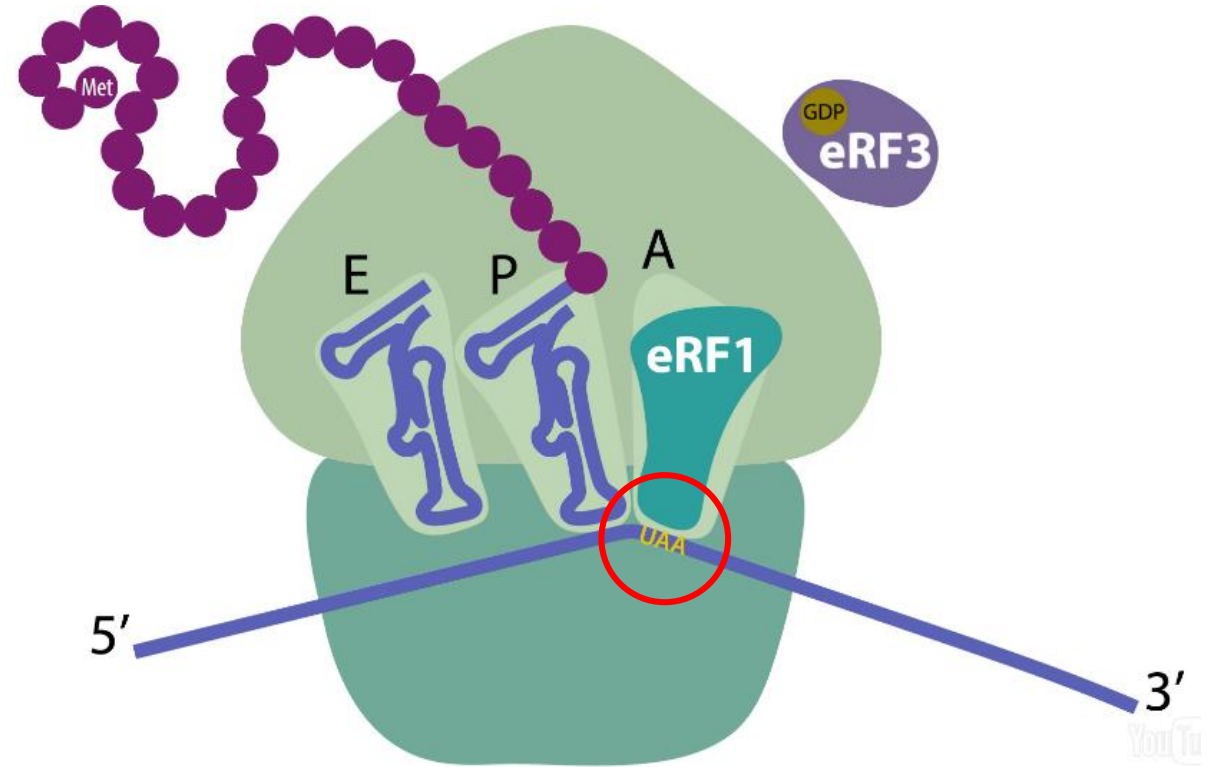
# Translation termination

1. A complex formed by **eRF1** and **eRF3-GTP** enters the ribosome



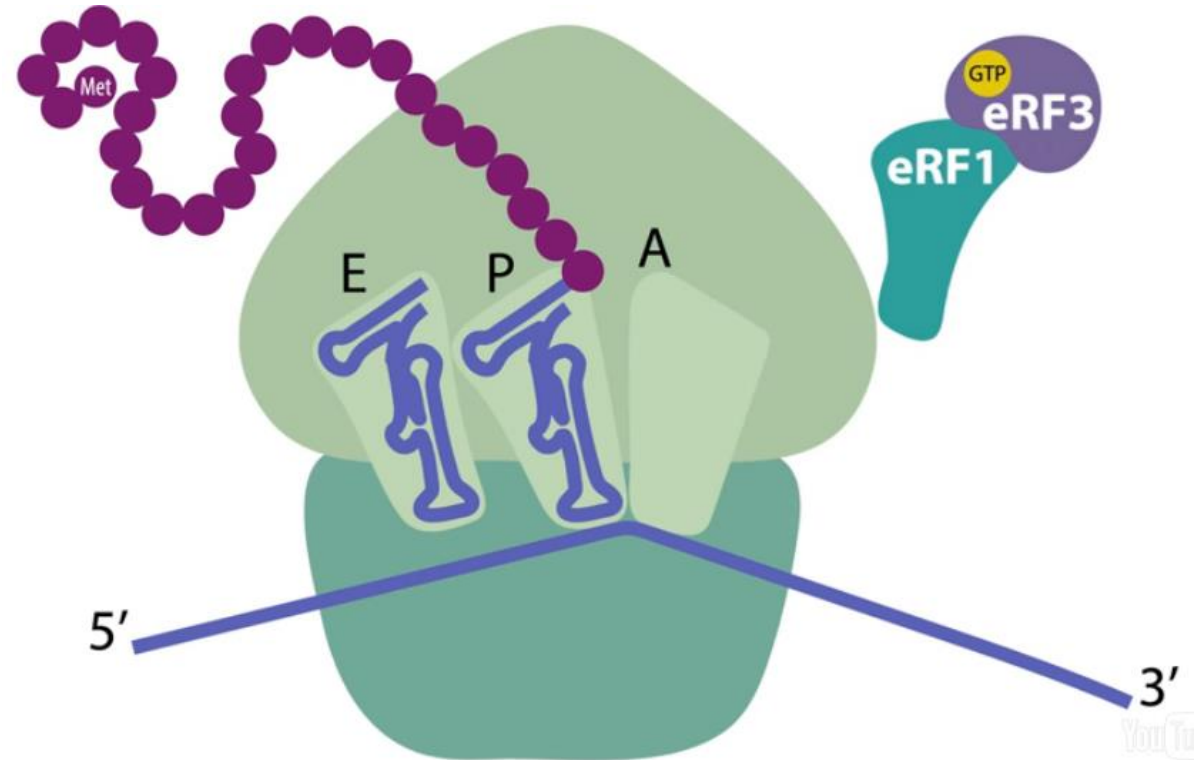
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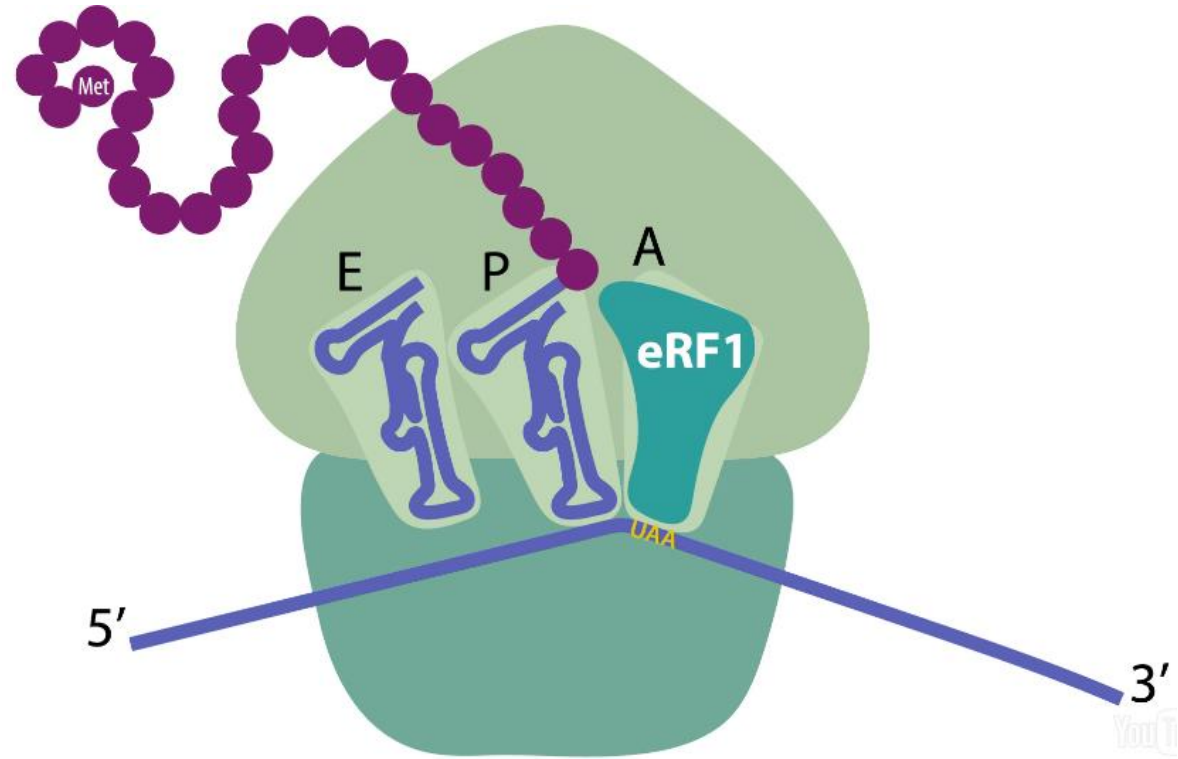
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3. GTP hydrolysis **activates eRF1** so that it can efficiently stimulate nascent chain release



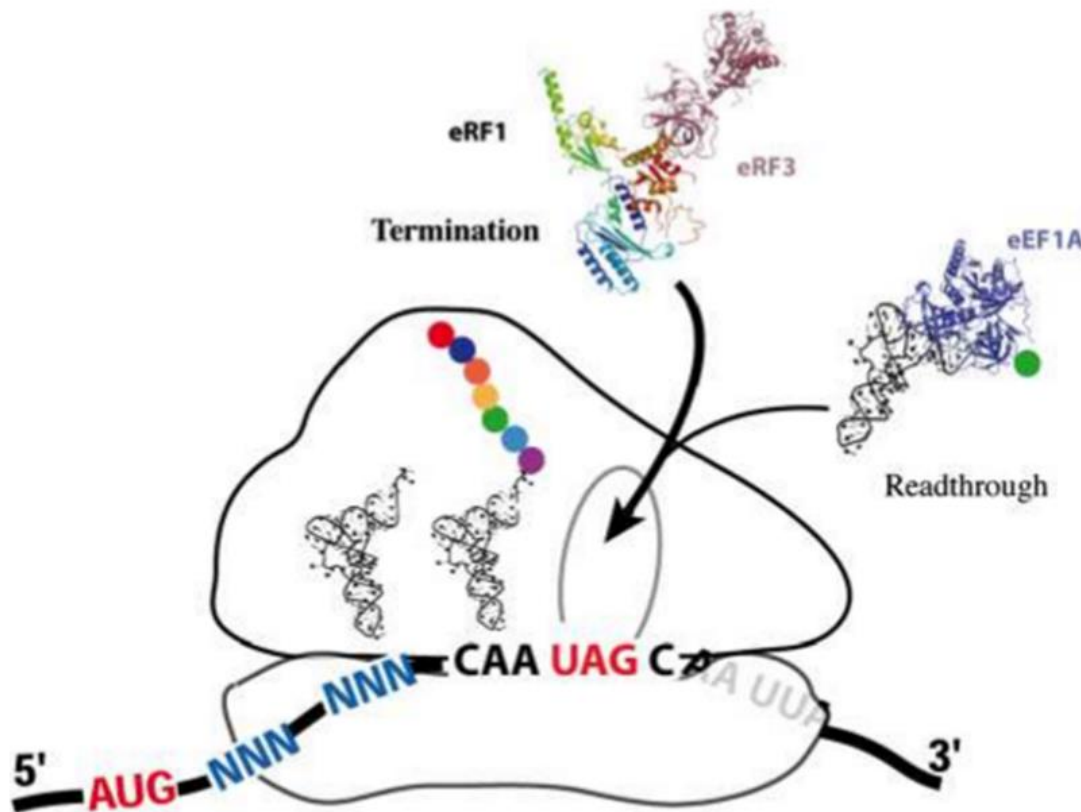


Translation termination is normally a highly efficient process

**BUT** occasionally stop codon recognition by eRF1 can be superseded by selected aminoacyl-tRNAs, resulting in **stop codon suppression**

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This event is called **READTHROUGH** and corresponds to the **incorporation of a tRNA**, or natural suppressor, **at the stop codon**, allowing translation to continue in the same frame until the ribosome reaches the next stop

The more efficient is translation termination, the less frequent is readthrough (and viceversa)



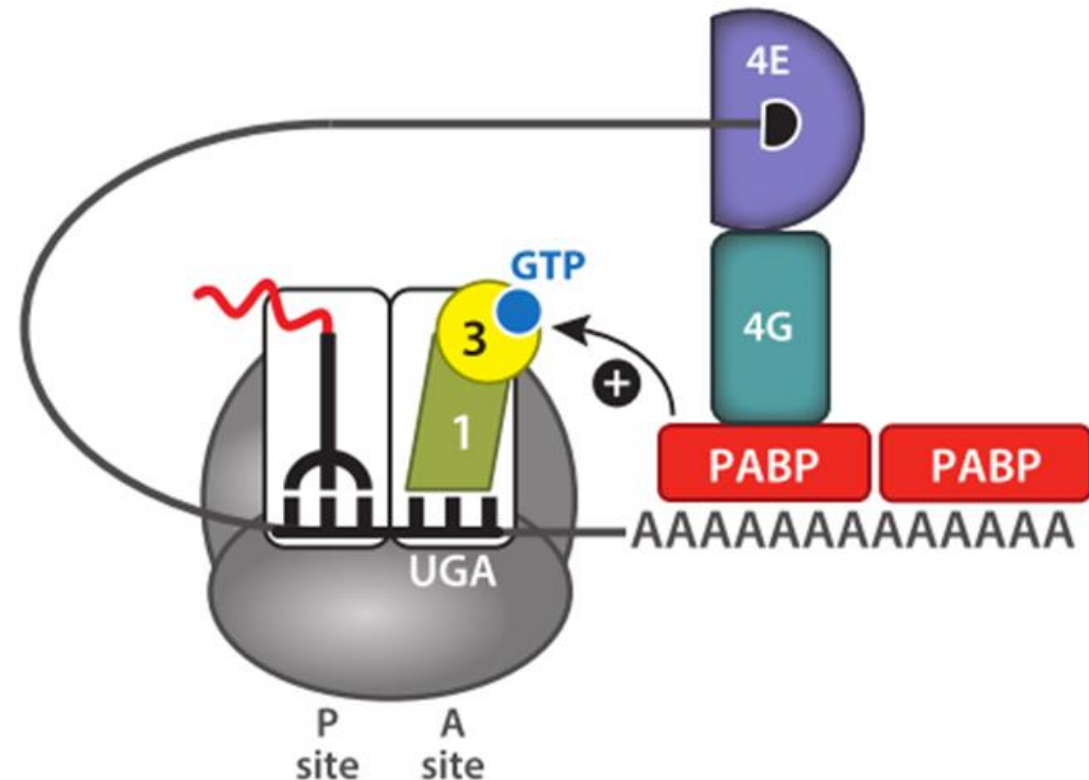
The efficiency of translation termination (and the occurrence of readthrough) can vary depending on many factors

The efficiency of translation termination (and the occurrence of readthrough) can vary depending on:

**1)** The efficiency of termination differs between normal stop codons and premature termination codons (PTC)

Normal STOP codon

PABP interacts with eRF3, promoting translation termination

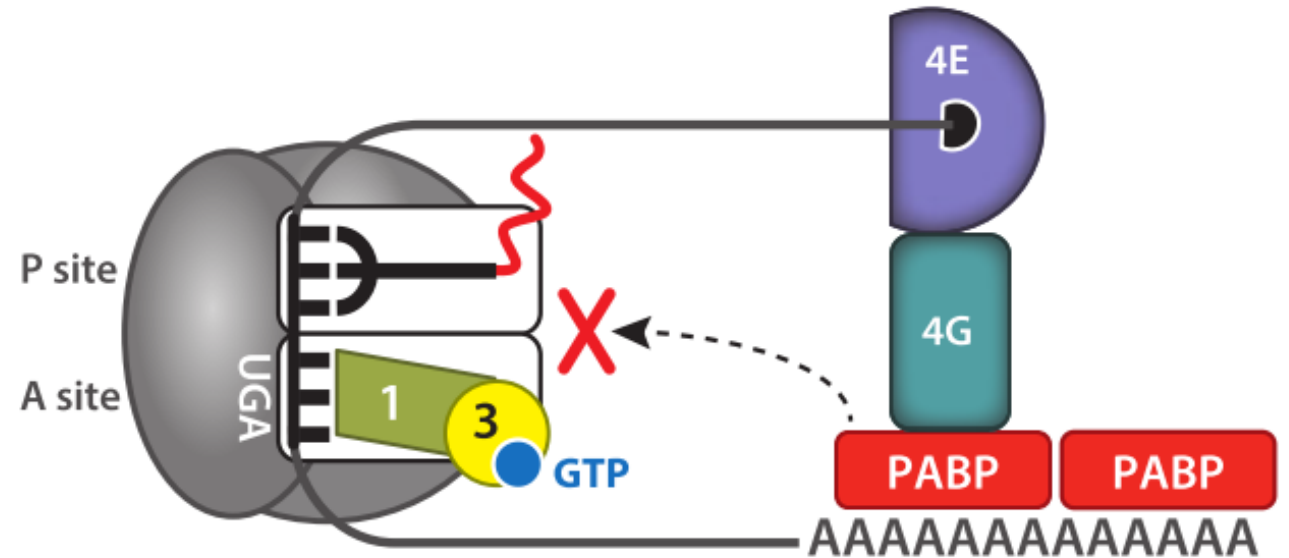


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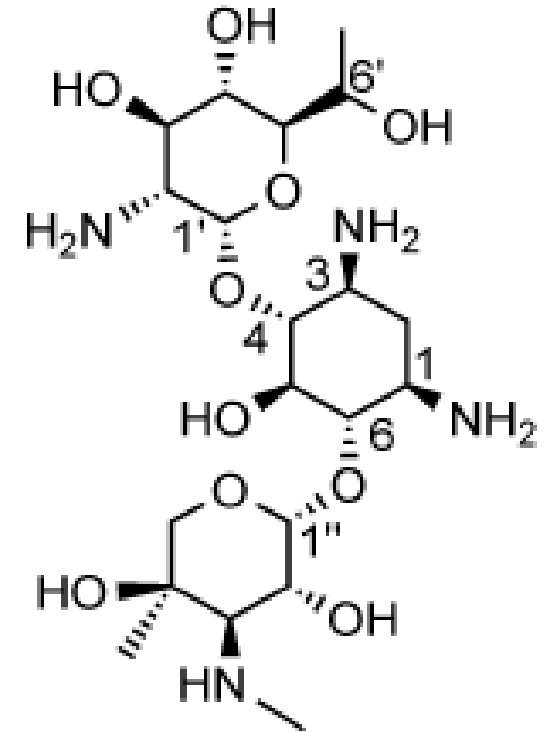
Premature STOP codon

PABP can not interact with eRF3, leading to prolonged ribosomal pausing at PTC and increasing aa-tRNA sampling



2) Aminoglycosides can decrease the fidelity of translation, causing higher frequencies of readthrough

**Aminoglycosides** are a class of antibiotics that interfere with bacterial-protein synthesis. They all have a common 2-deoxystreptamine ring structure, which binds to the ribosome decoding center.



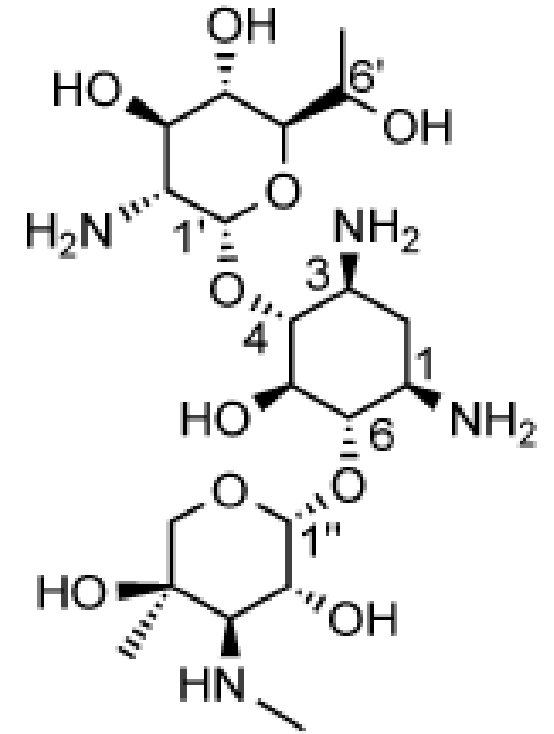
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**Aminoglycosides** are a class of antibiotics that interfere with bacterial-protein synthesis. They all have a common 2-deoxystreptamine ring structure, which binds to the ribosome decoding center.

Ribosome is composed by **two subunits**:

- The large subunit contains the peptidyl transferase center, in which peptide bonds are formed
- The **small subunit** contains the **decoding center**, a region in which the correct codon-anticodon pairing between mRNA and tRNAs is monitored



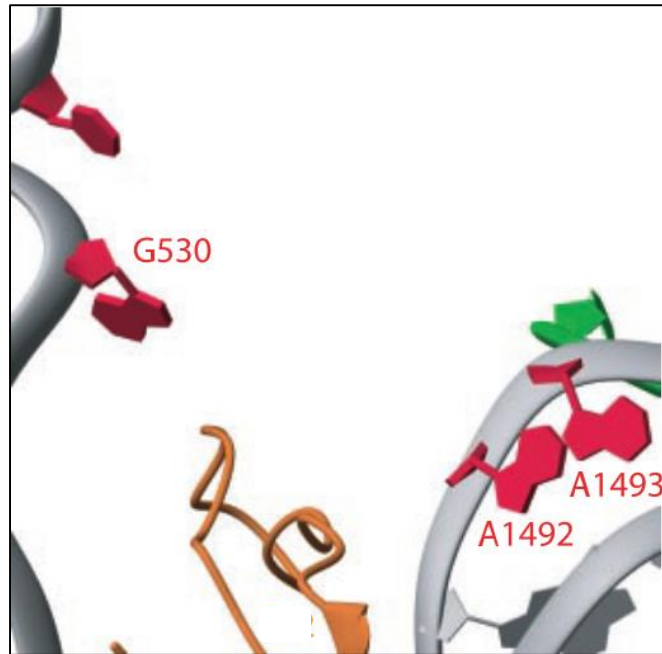
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**Mechanism of action:** aminoglycosides binding to ribosome decoding site induces a conformational change similar to the transition caused by a tRNA binding

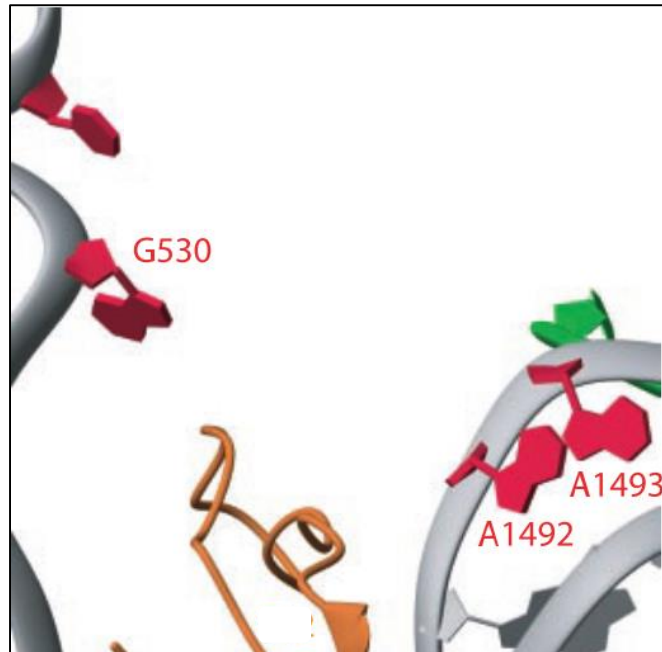
**NATIVE STRUCTURE**



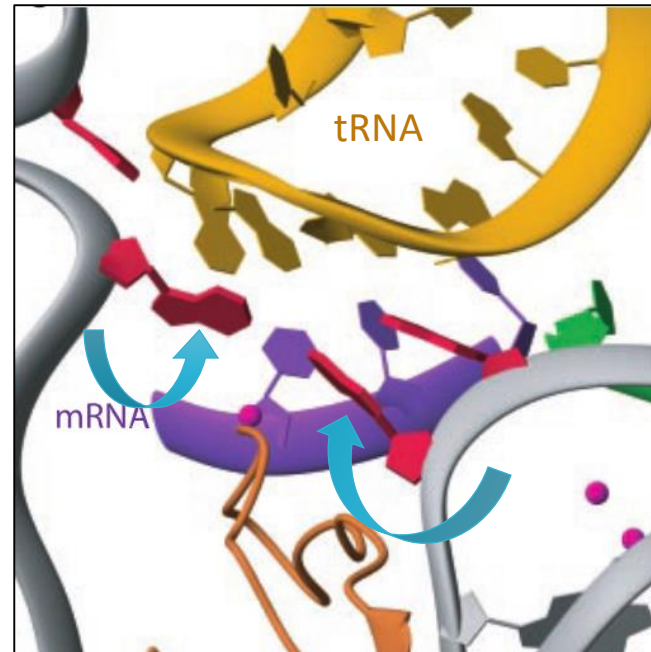
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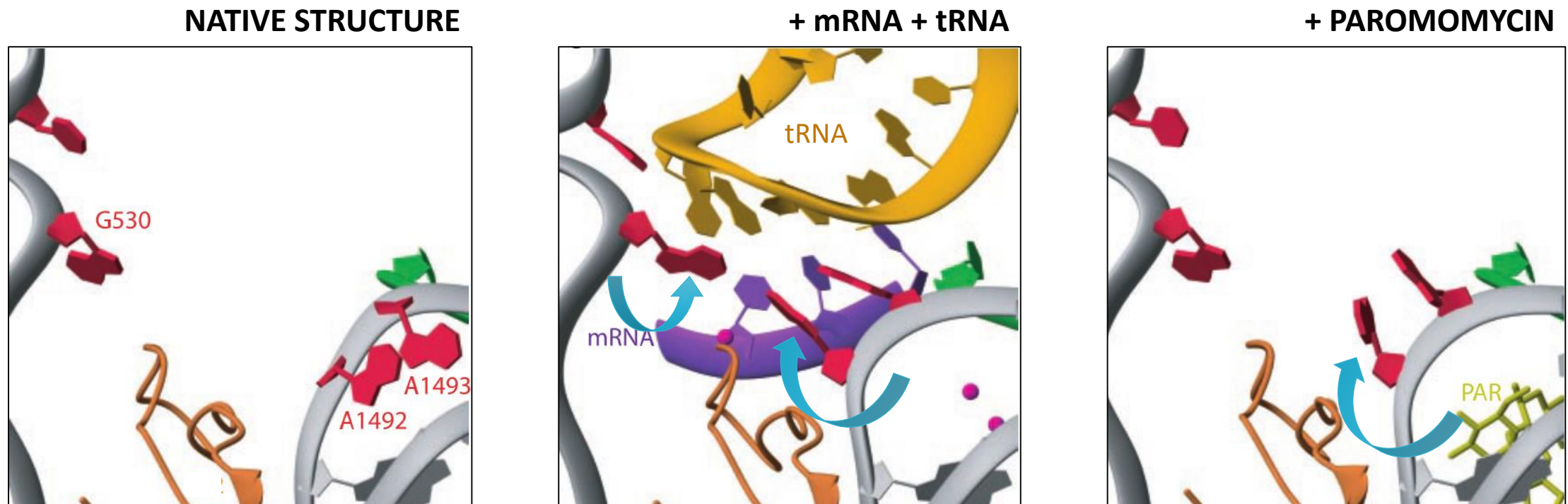


+ mRNA + tRNA



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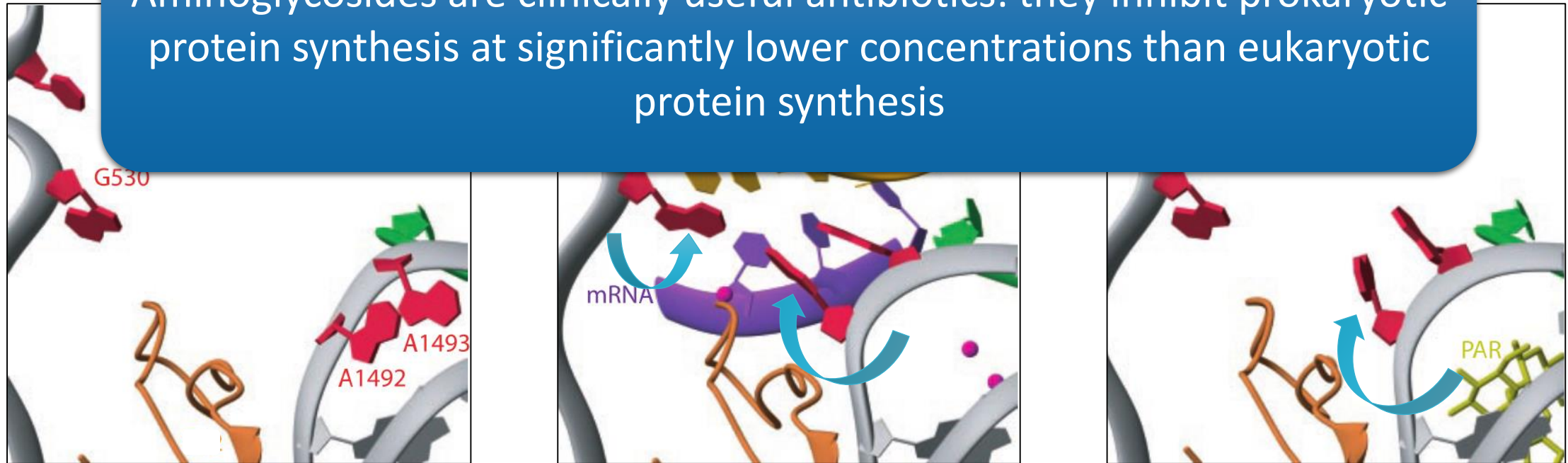
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2) Aminoglycosides can decrease the fidelity of translation, causing higher frequencies of readthrough

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Aminoglycosides are clinically useful antibiotics: they inhibit prokaryotic protein synthesis at significantly lower concentrations than eukaryotic protein synthesis



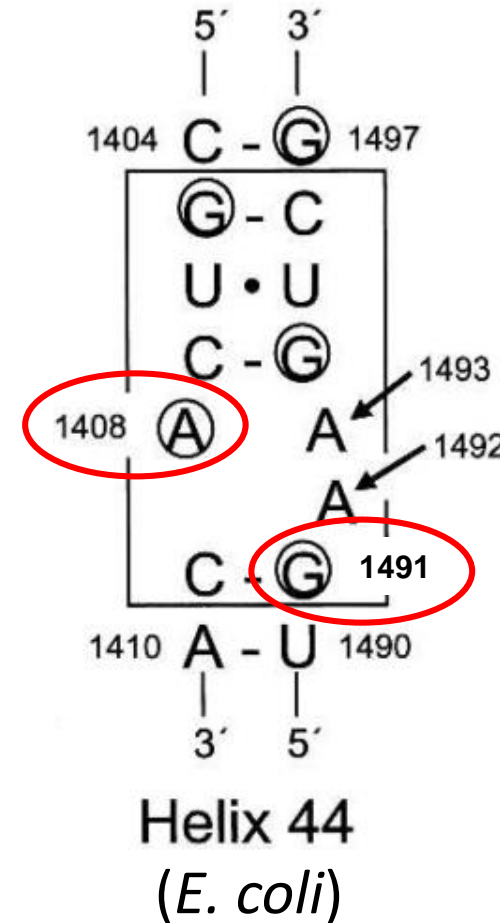
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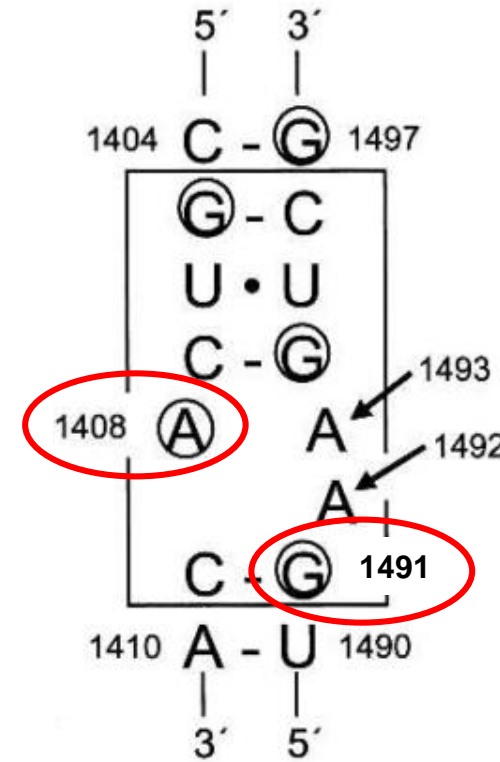
**A1408** and **G1491**



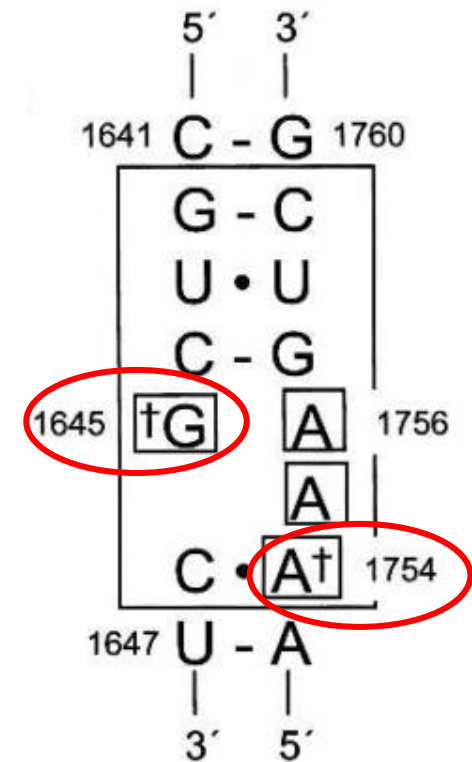
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**A1408** and **G1491** VS **G1645** and **A1754**



Helix 44  
(*E. coli*)



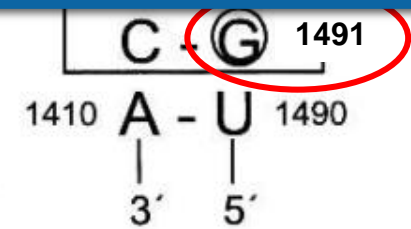
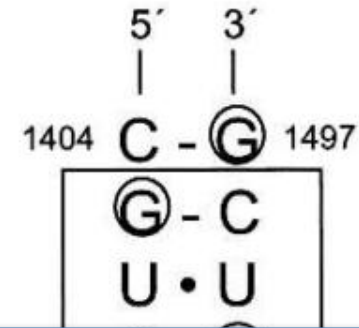
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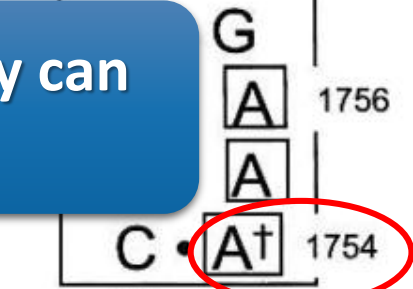
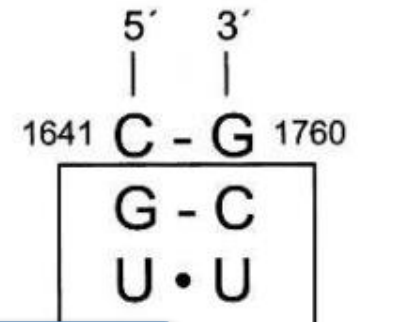
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**In eukaryotes aminoglycosides decrease ribosome fidelity and they can be used to induce readthrough**

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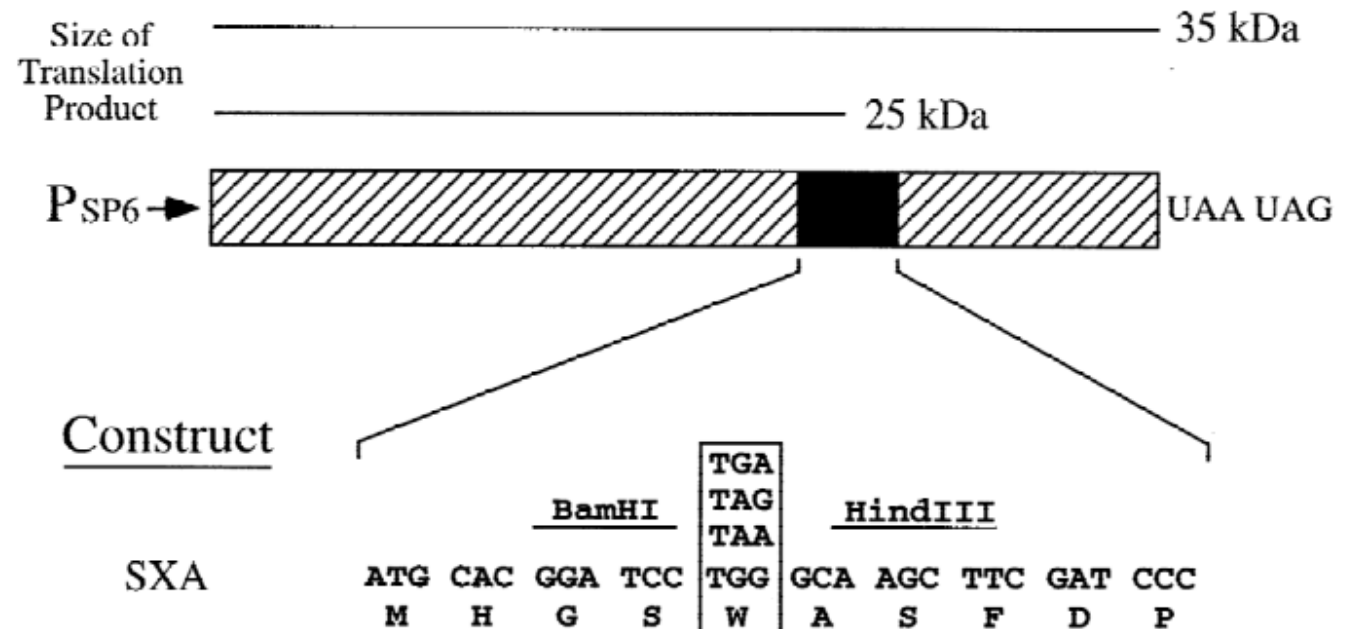


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Different stop codons promote translation termination with different efficiencies

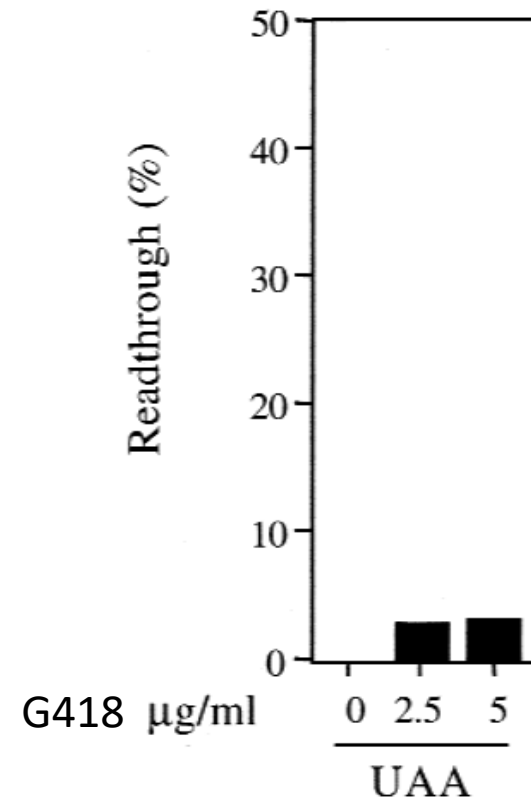
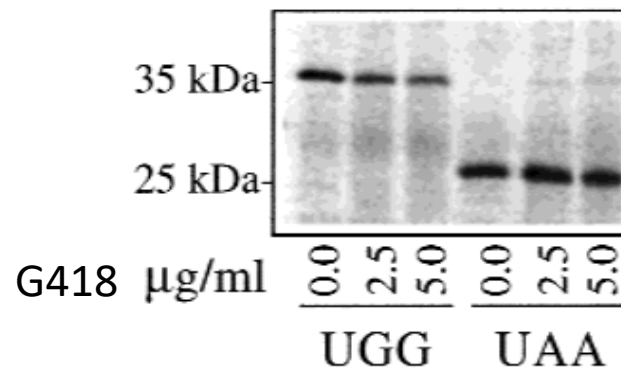
Readthrough reporter system:



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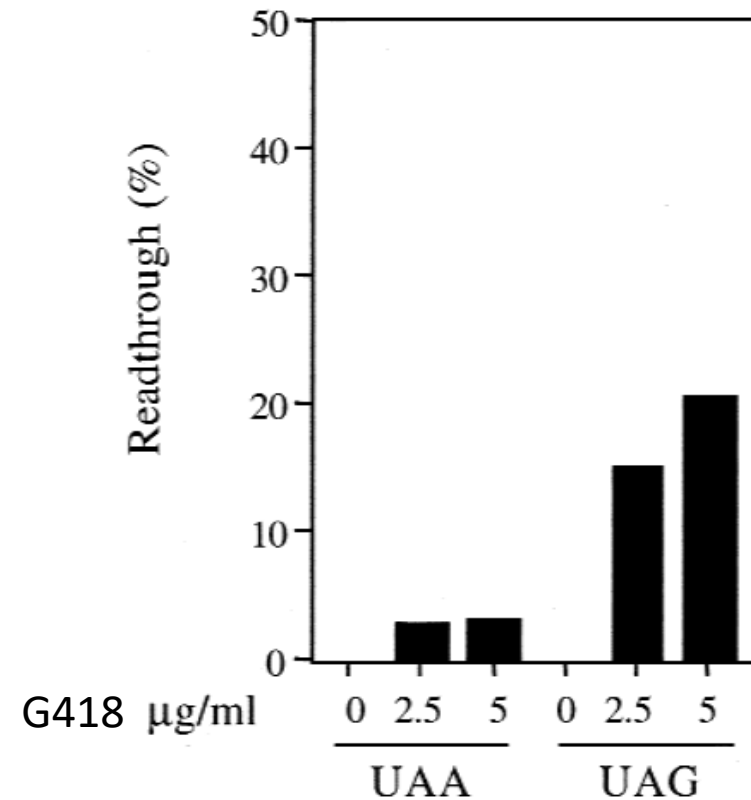
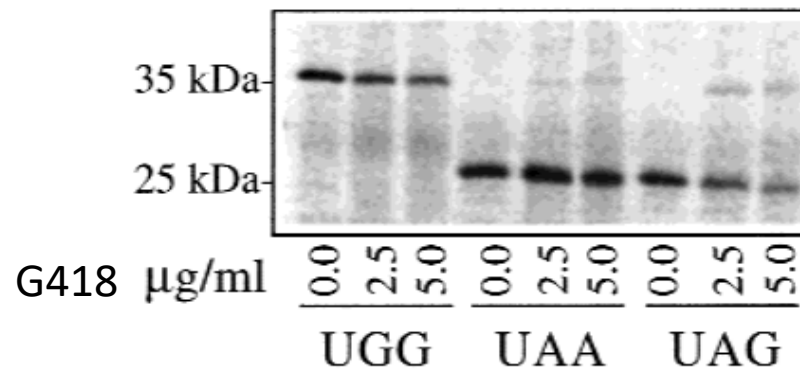
UAA



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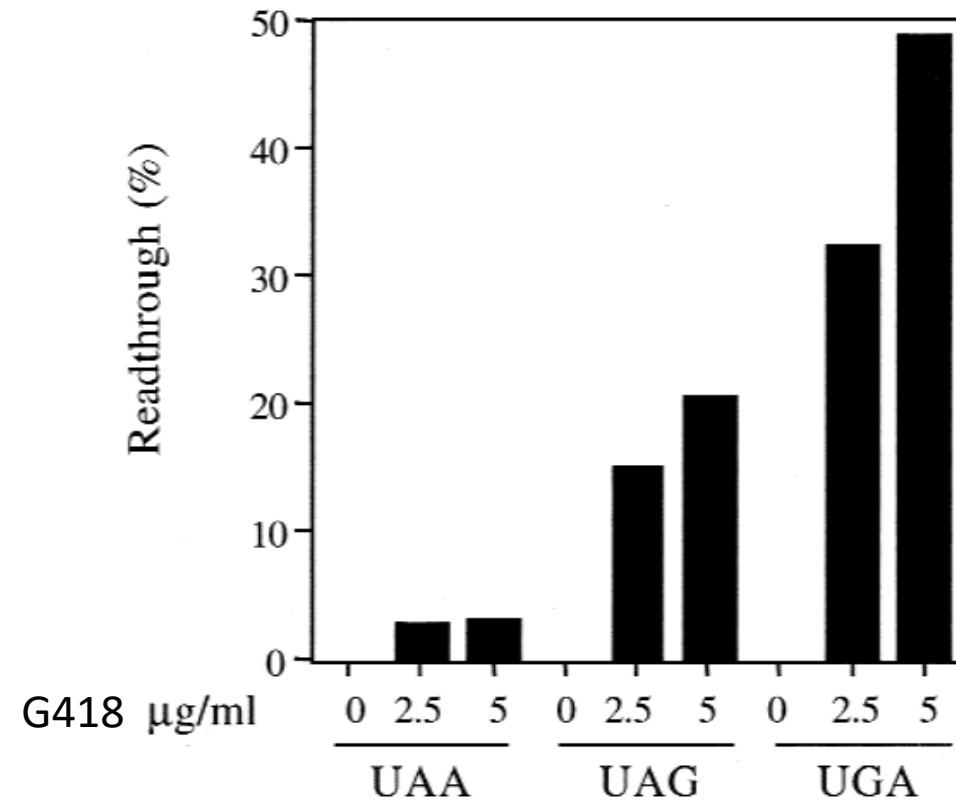
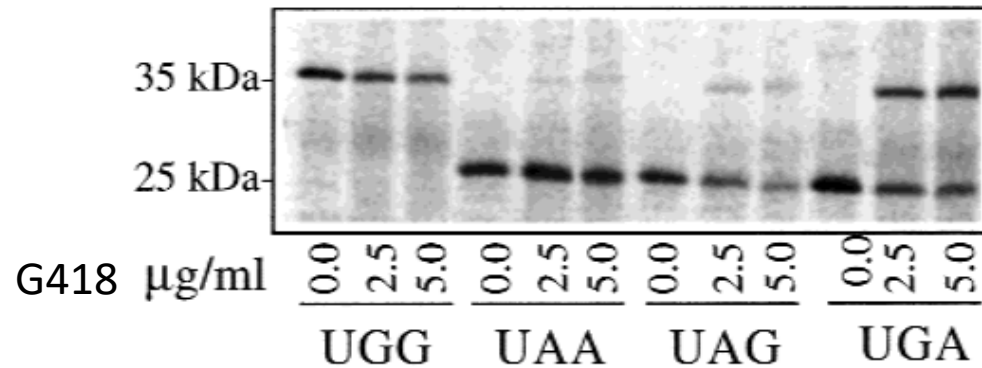
**UAA > UAG**



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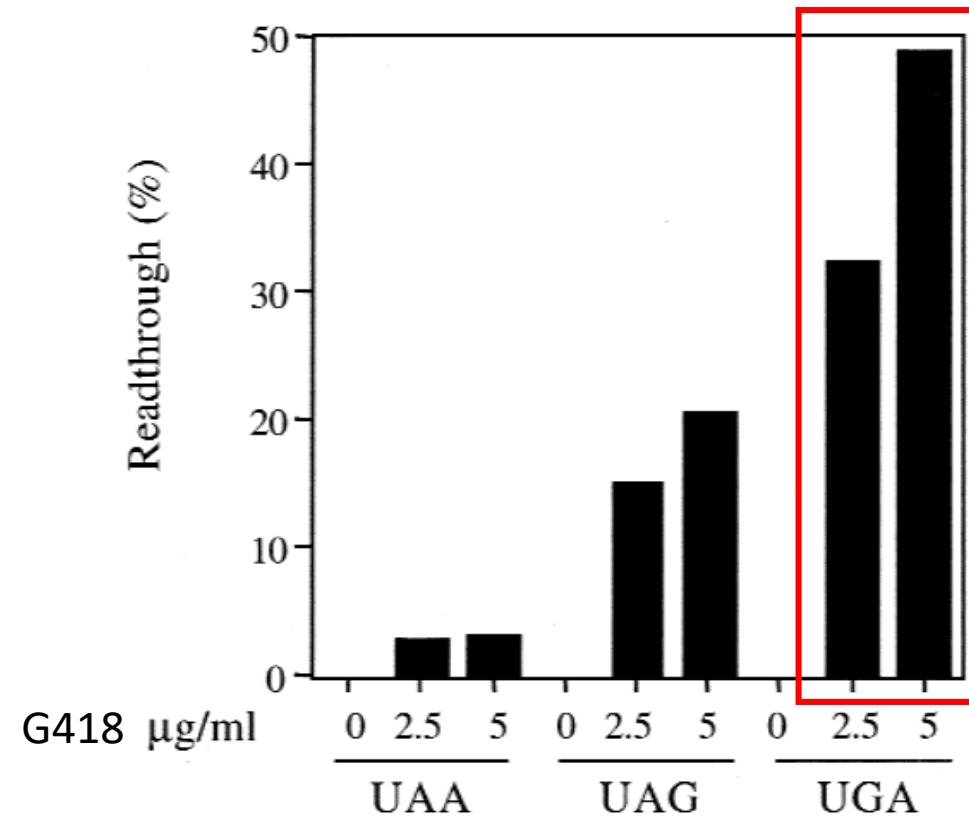
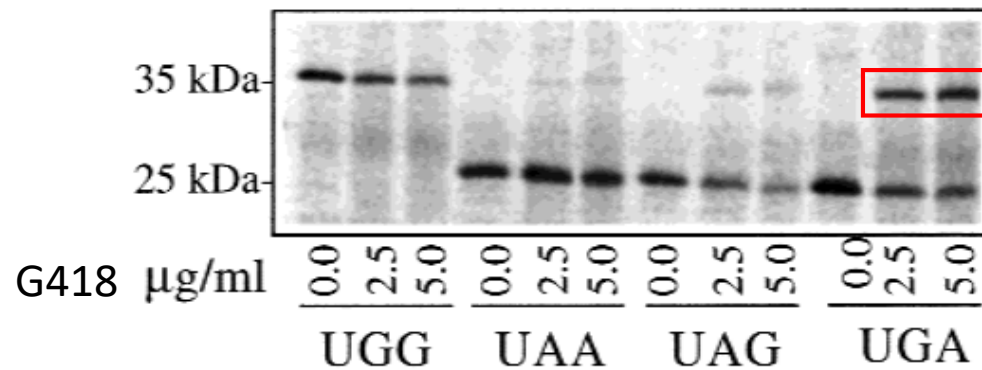
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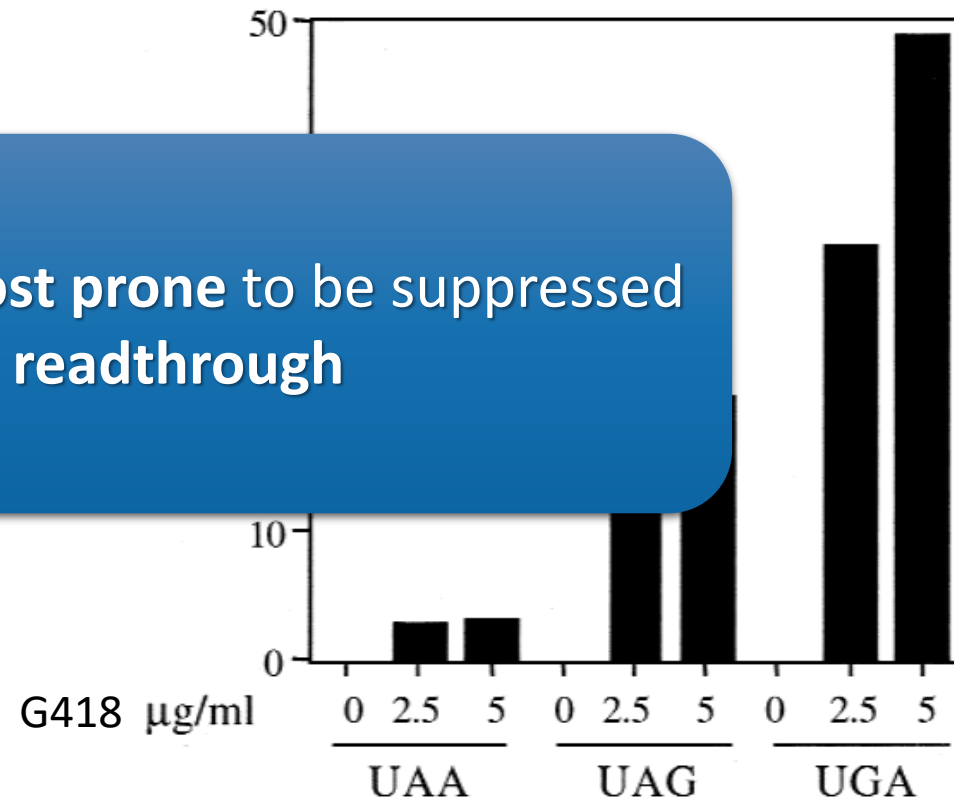
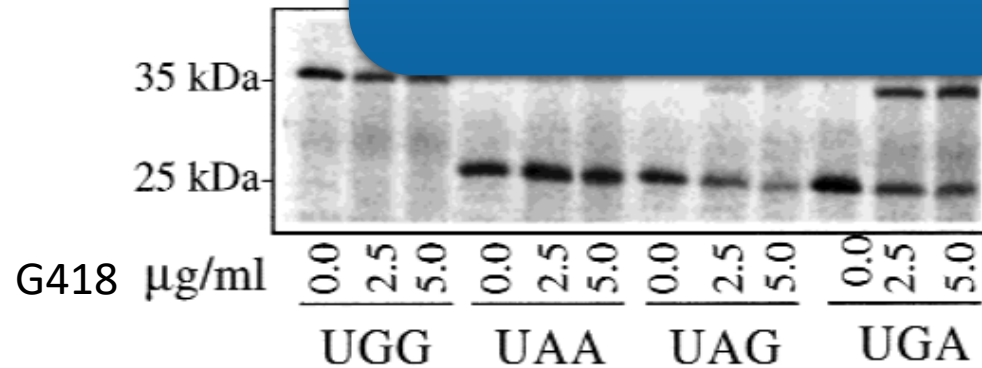


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UAA

**UGA stop codon is the most prone to be suppressed by ribosome readthrough**

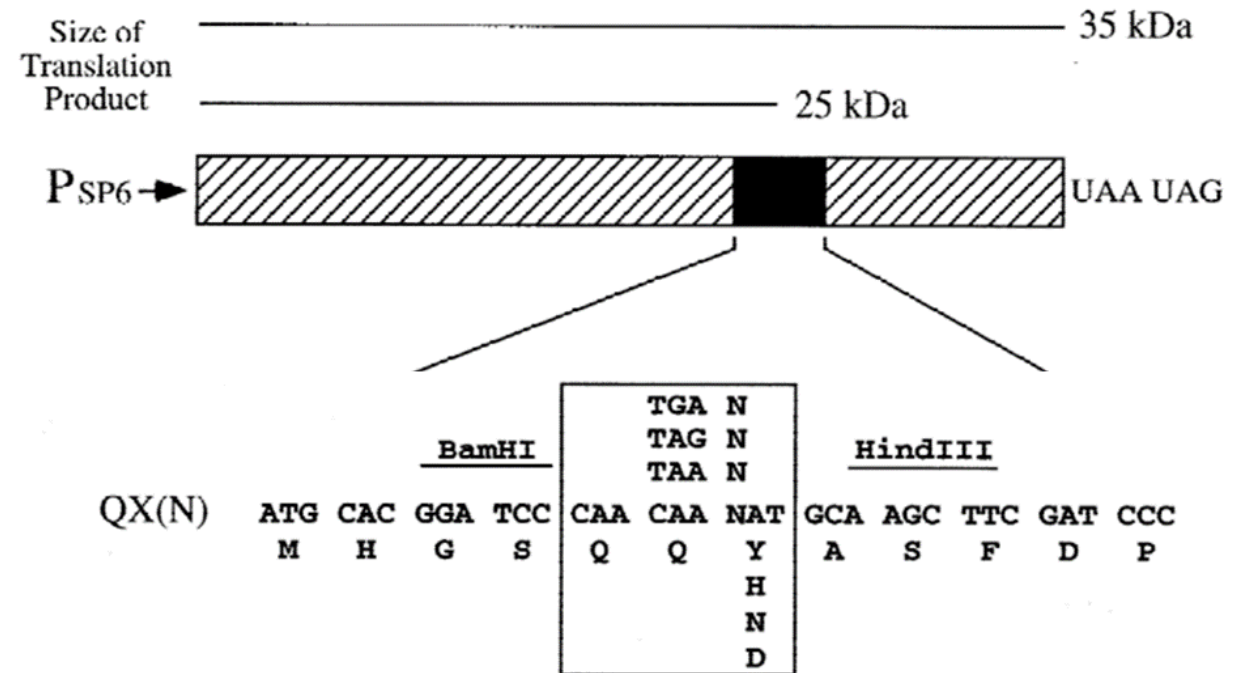


Also the 4th nucleotide strongly influences the occurrence of readthrough



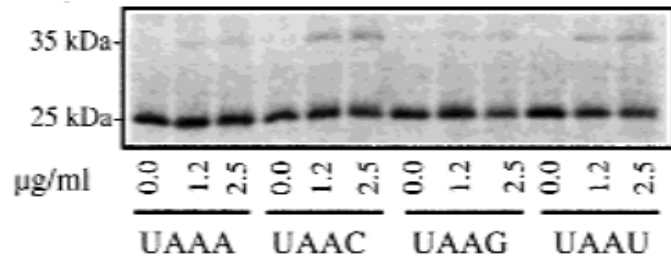
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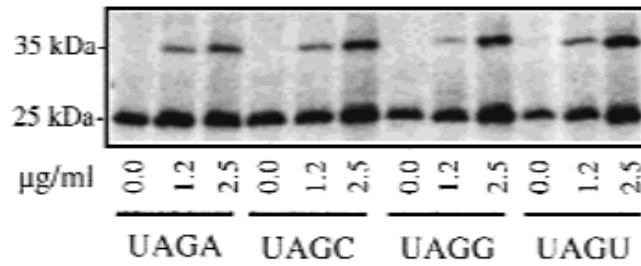


All possible combinations of stop codon and 4<sup>th</sup> nucleotide

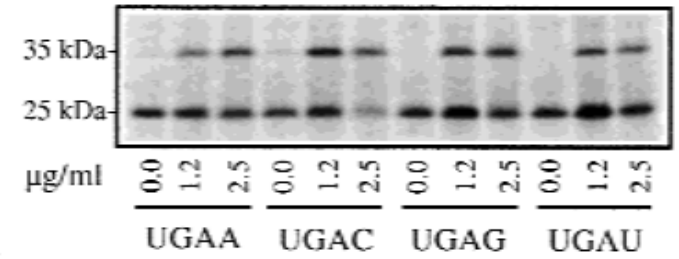
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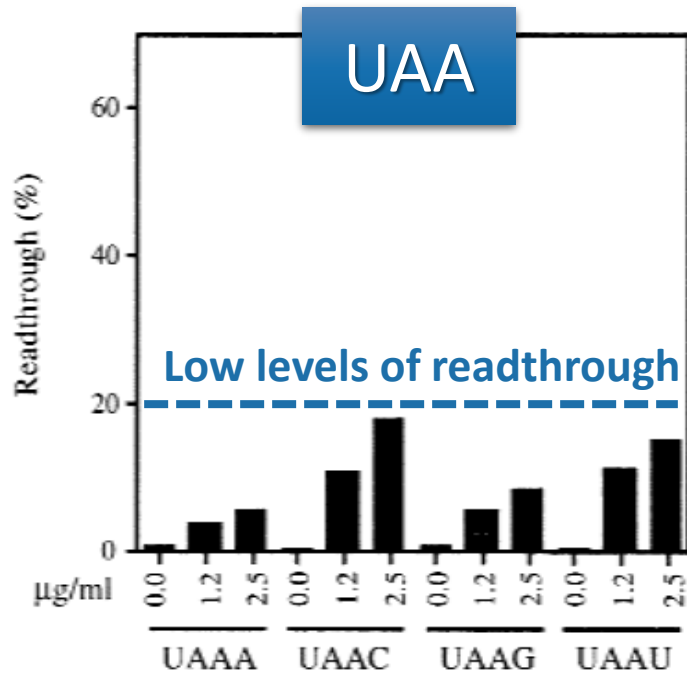
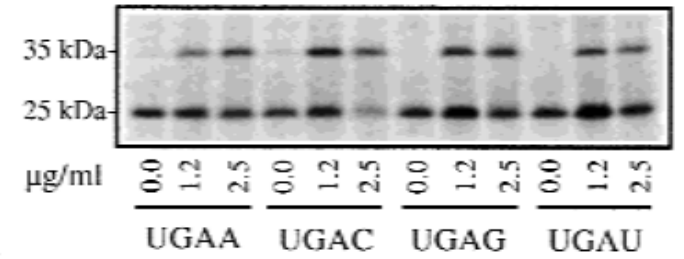
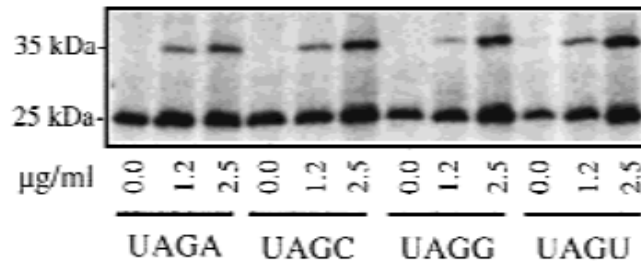
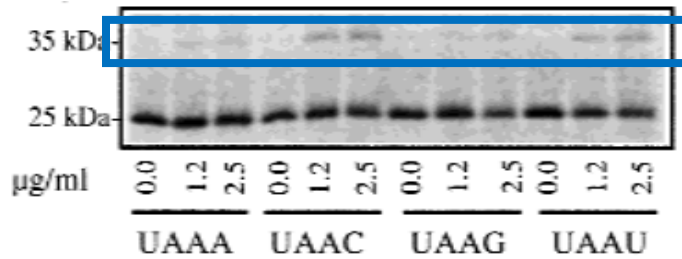
UAG



UGA

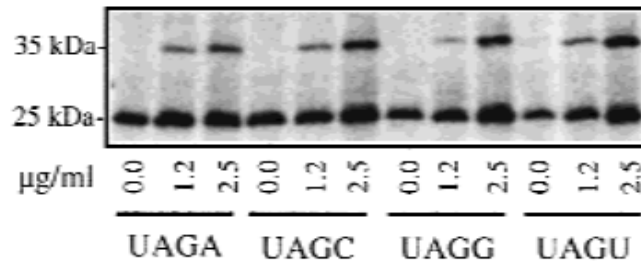
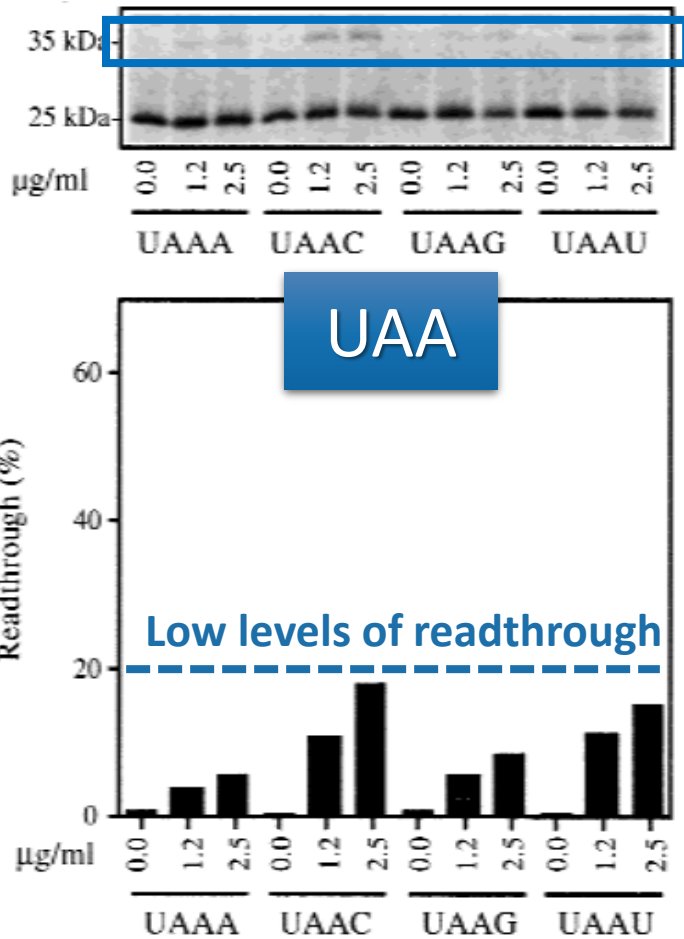
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### Efficient translation termination



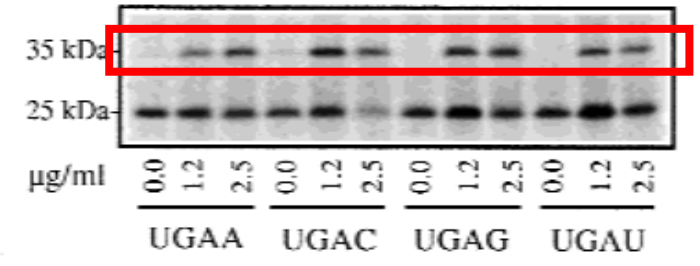
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**Efficient translation termination**

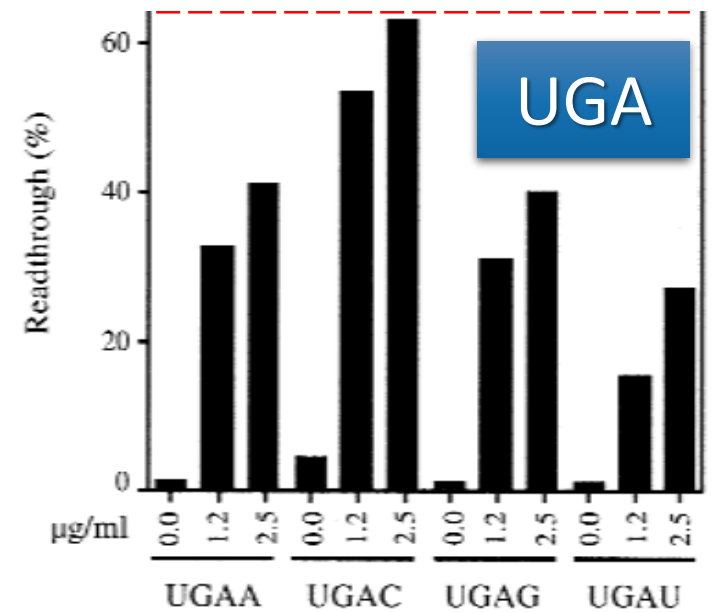


**UAG**

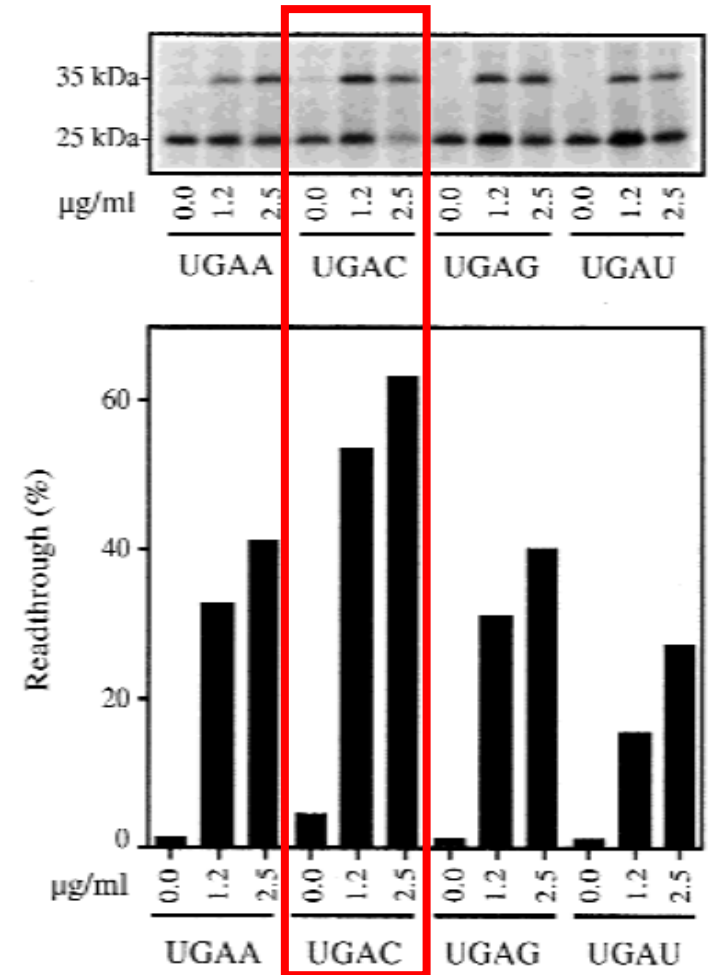
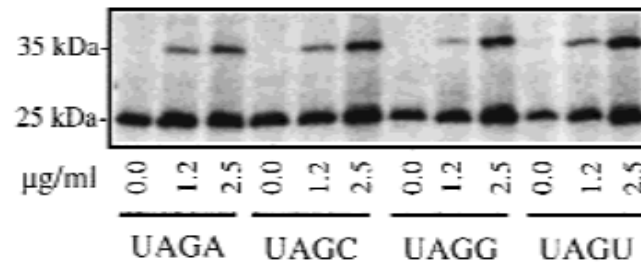
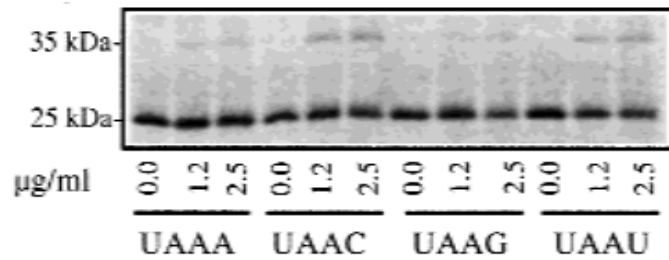
**Less efficient translation termination**



**High levels of readthrough**



Also the 4th nucleotide strongly influences the occurrence of readthrough



The tetranucleotide **UGAC** shows the most frequent stop codon suppression, with readthrough occurring at a frequency of 3–4% (spontaneous) and 63% (G418-induced)

## The efficiency of translation termination (and the occurrence of readthrough) can vary depending on many factors

- 1) The efficiency of termination differs between normal stop codons and premature termination codons
- 2) Aminoglycosides can decrease the fidelity of translation, causing higher frequencies of readthrough
- 3) The stop codon type and the 4<sup>th</sup> nucleotide strongly influence efficiency of translation termination and, as a consequence, occurrence of readthrough



The restoration of protein activity depends strongly on the identity of the amino acid inserted during PTC readthrough



# The restoration of protein activity depends strongly on the identity of the amino acid inserted during PTC readthrough

Published online 23 July 2014

*Nucleic Acids Research*, 2014, Vol. 42, No. 15 10061–10072  
doi: 10.1093/nar/gku663

## **New insights into the incorporation of natural suppressor tRNAs at stop codons in *Saccharomyces cerevisiae***

Sandra Blanchet<sup>1</sup>, David Cornu<sup>2</sup>, Manuela Argentini<sup>2</sup> and Olivier Namy<sup>1,3,\*</sup>

Development of an *in vivo* reporter system to study amino acid insertion at all stop codons

The restoration of protein activity depends strongly on the identity of the amino acid inserted during PTC readthrough

### Results

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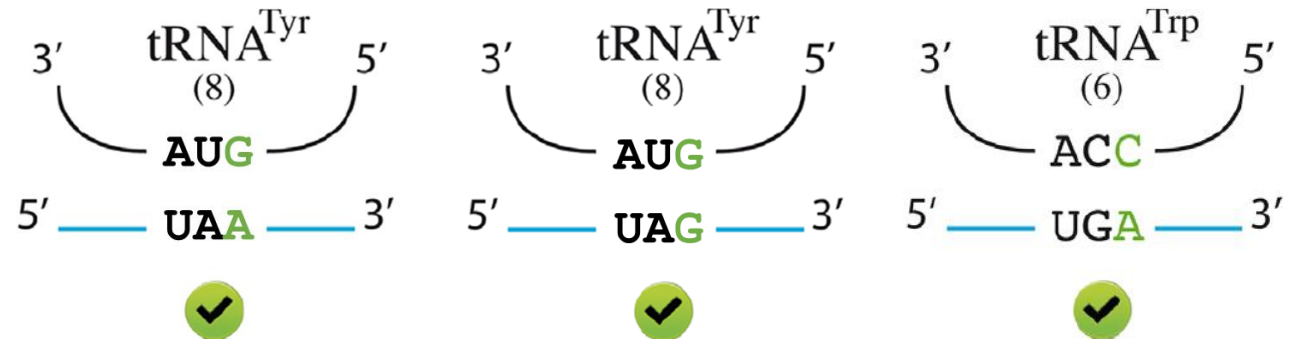
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- Only a subset of predictable suppressor tRNAs are actually incorporated at the various stop codons

**Tyrosine, glutamine** and **lysine** can be inserted at **UAA** and **UAG** codons

**Tryptophan, cysteine** and **arginine** can be inserted at **UGA** codons



**Near-cognate tRNAs:** different in only one position of the codon-anticodon pairing

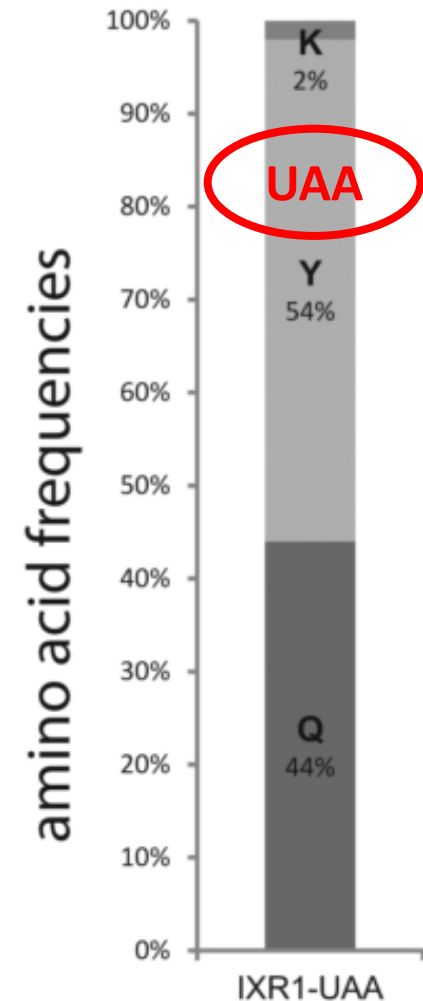
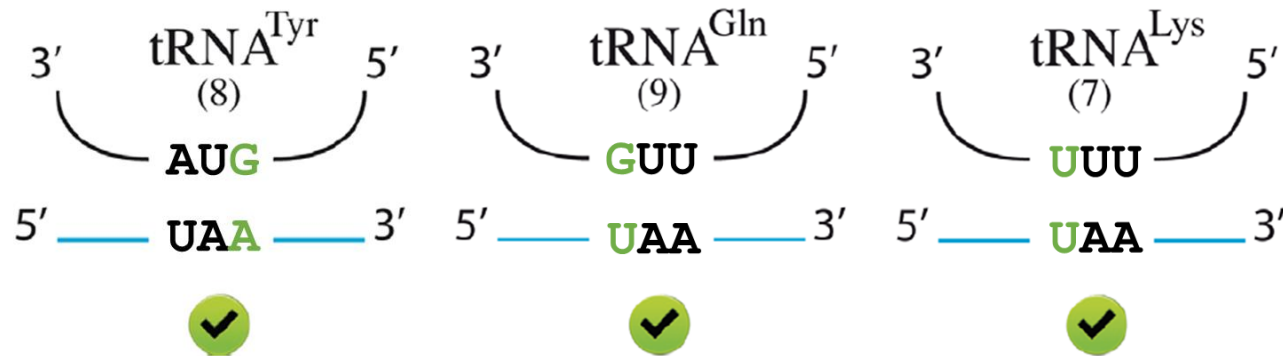
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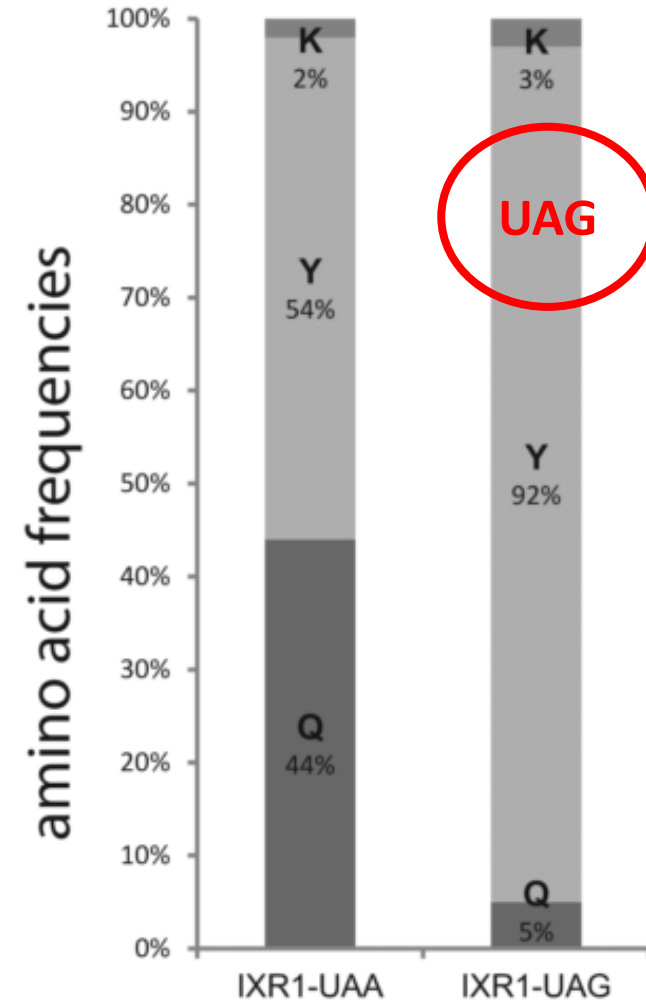
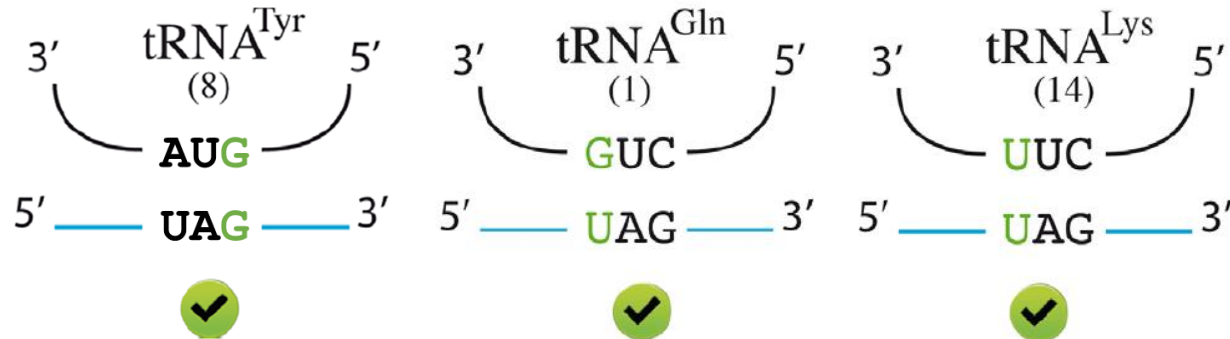
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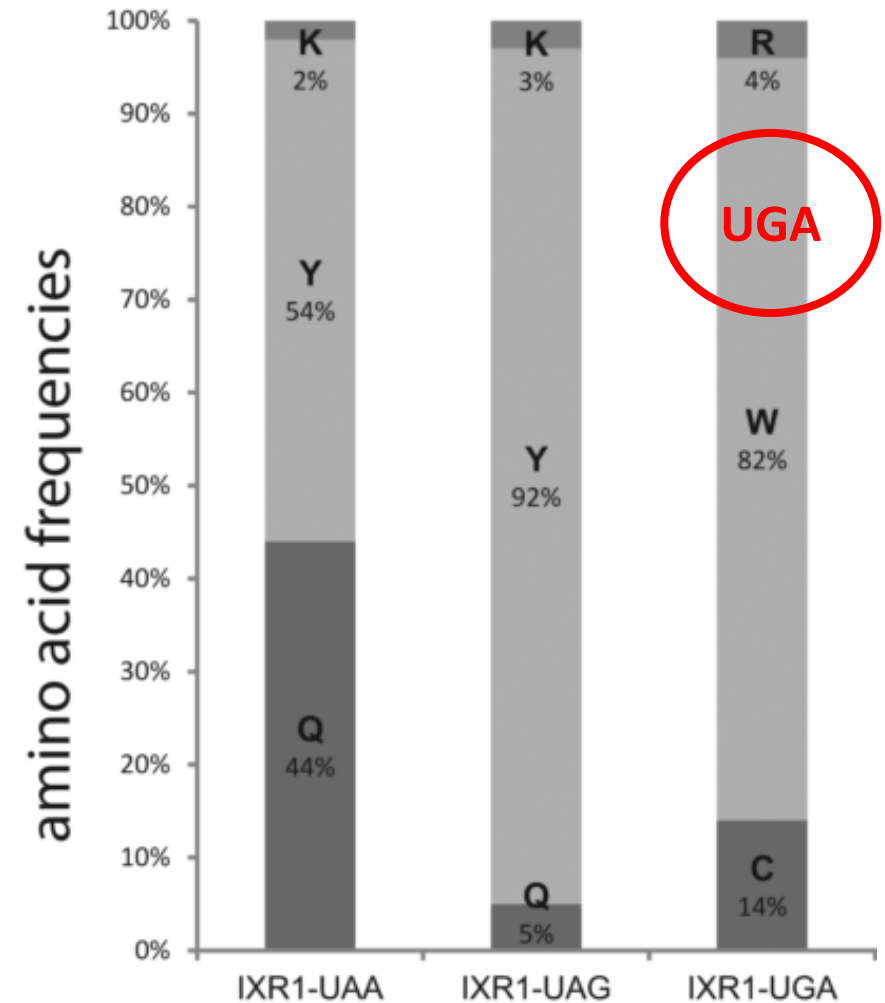
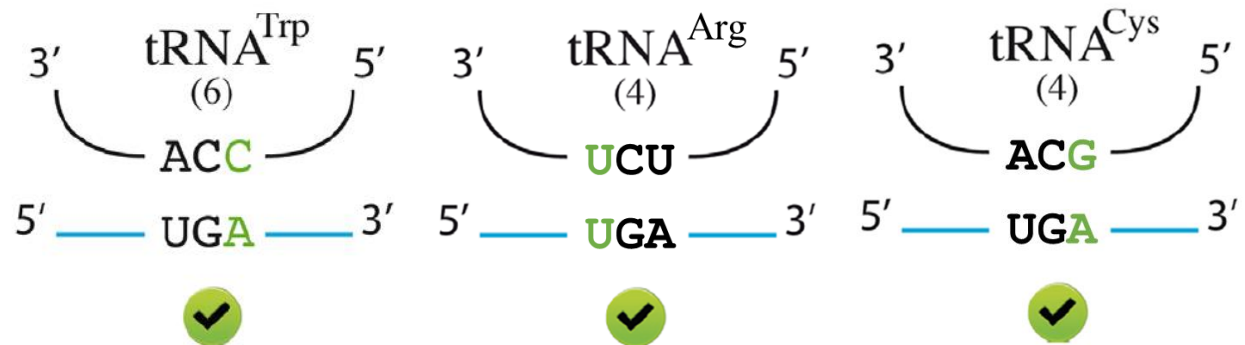
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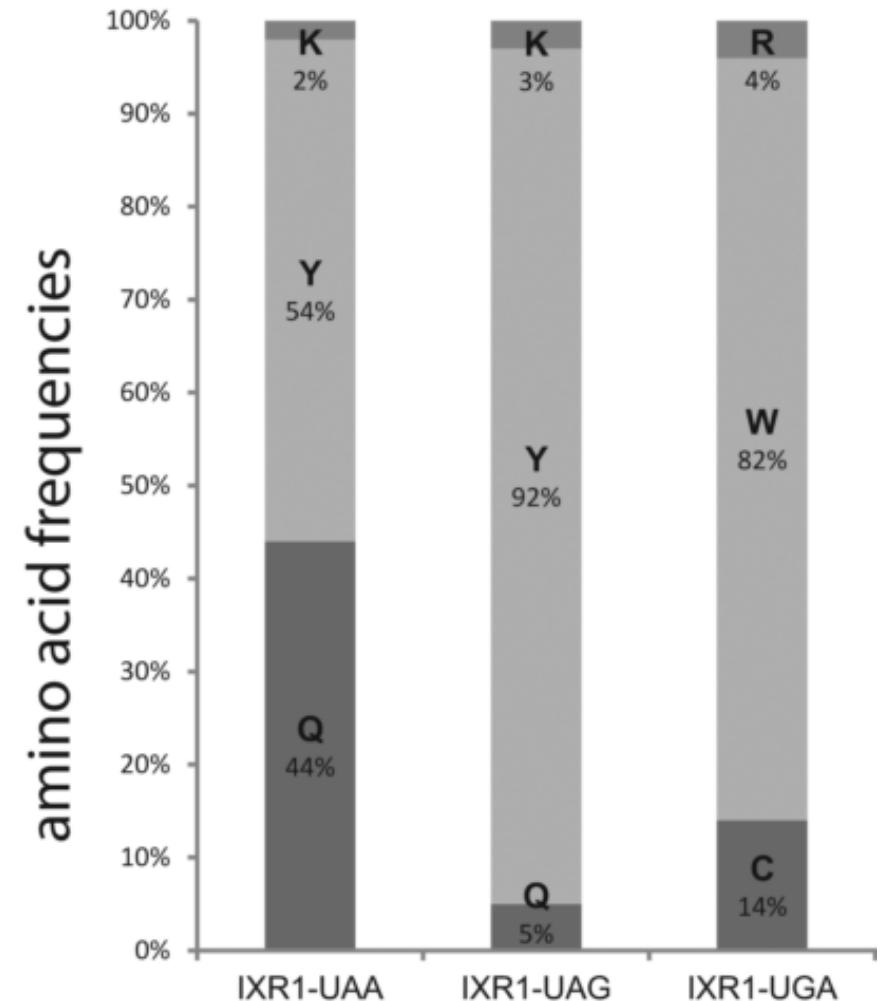




The restoration of protein activity depends strongly on the identity of the amino acid inserted during PTC readthrough

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It is possible to predict the probable protein sequences arising from a readthrough event on the basis of the stop codon present





# Brief overview...

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**BRIEF REPORT**

## Differential functional readthrough over homozygous nonsense mutations contributes to the bleeding phenotype in coagulation factor VII deficiency

A. BRANCHINI,<sup>\*†</sup> M. FERRARESE,<sup>\*</sup> S. LOMBARDI,<sup>\*</sup> R. MARI,<sup>‡</sup> F. BERNARDI<sup>\*†</sup> and M. PINOTTI<sup>\*†</sup>



**BRIEF REPORT**

## Differential functional readthrough over homozygous nonsense mutations contributes to the bleeding phenotype in coagulation factor VII deficiency

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### Objective

Evaluate the spontaneous and drug-induced readthrough levels of two nonsense mutations in coagulation factor VII (FVII) : **p.Ser112X** and **p.Cys132X**

Expected to be lethal

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Life-threatening  
bleeding  
symptoms

Moderate  
bleeding  
symptoms

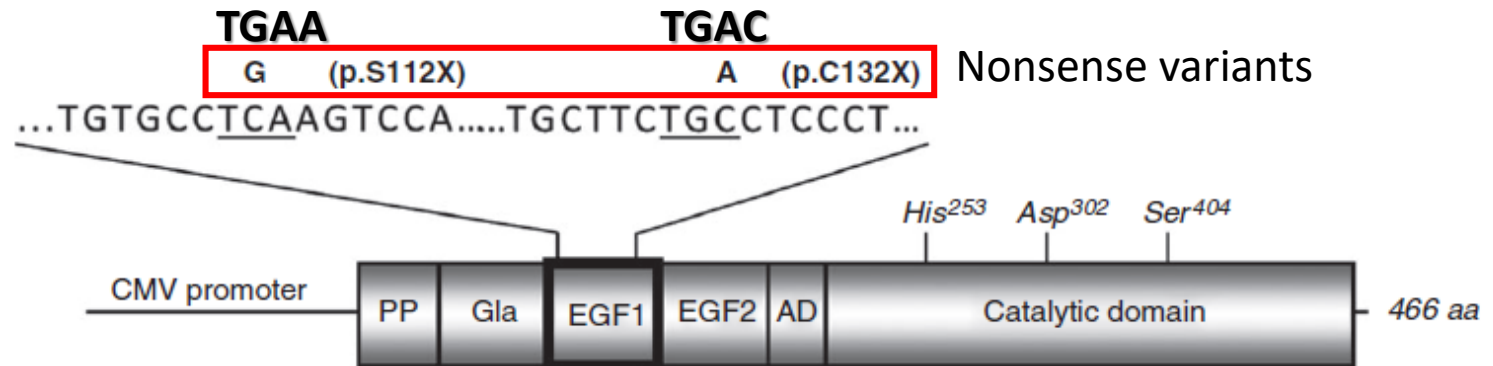
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## Methods

1. Creation of a cellular model through the transient expression of recombinant FVII **nonsense** variants
2. Evaluation of secreted levels of rFVII by ELISA



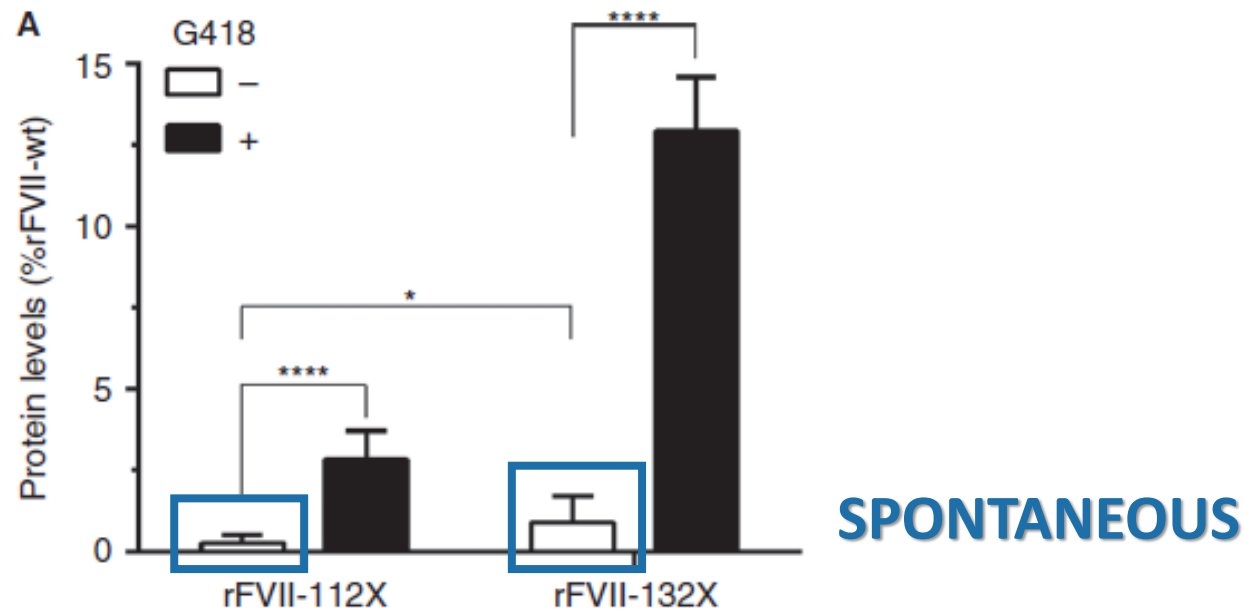
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## Results

### Secretion levels of rFVII nonsense variants



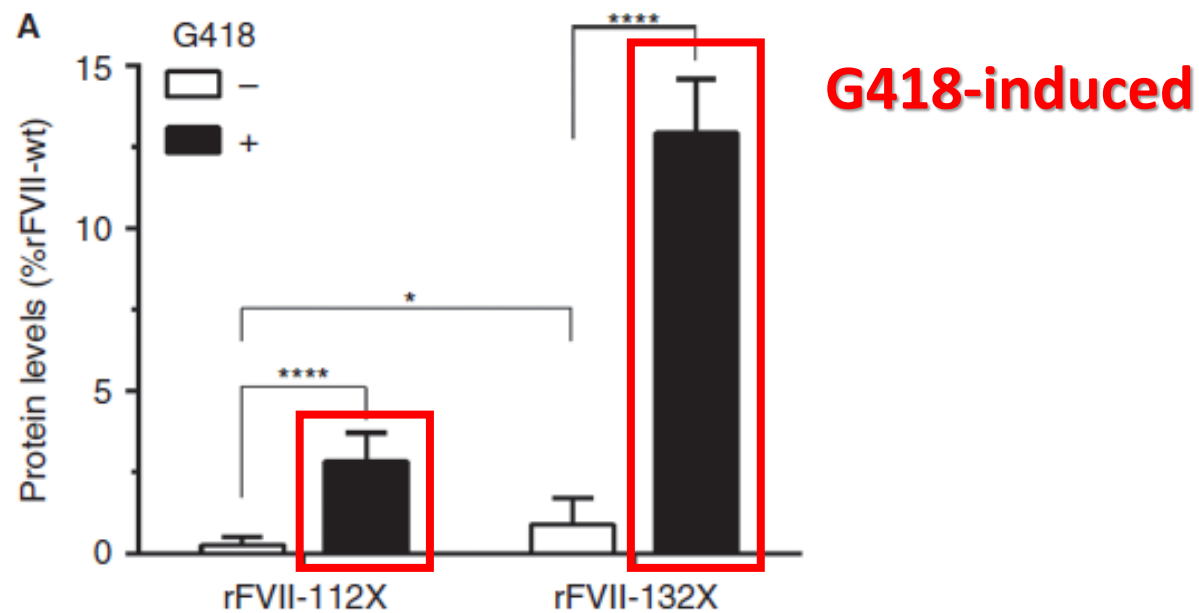
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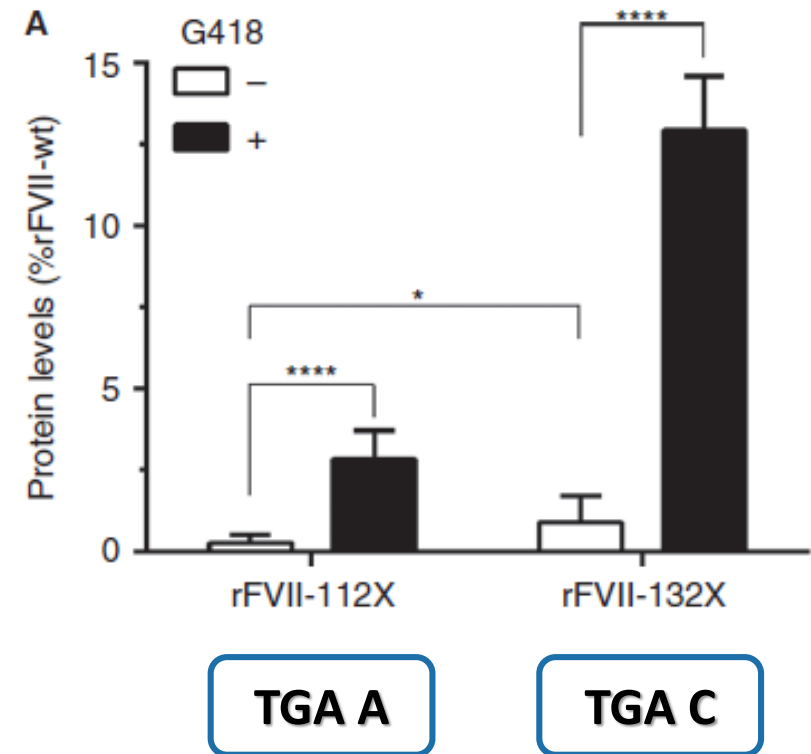
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## Results

Secretion and activity levels of rFVII nonsense variants

rFVII C132X shows the higher degree of suppression, probably due to the more readthrough-favourable sequence context



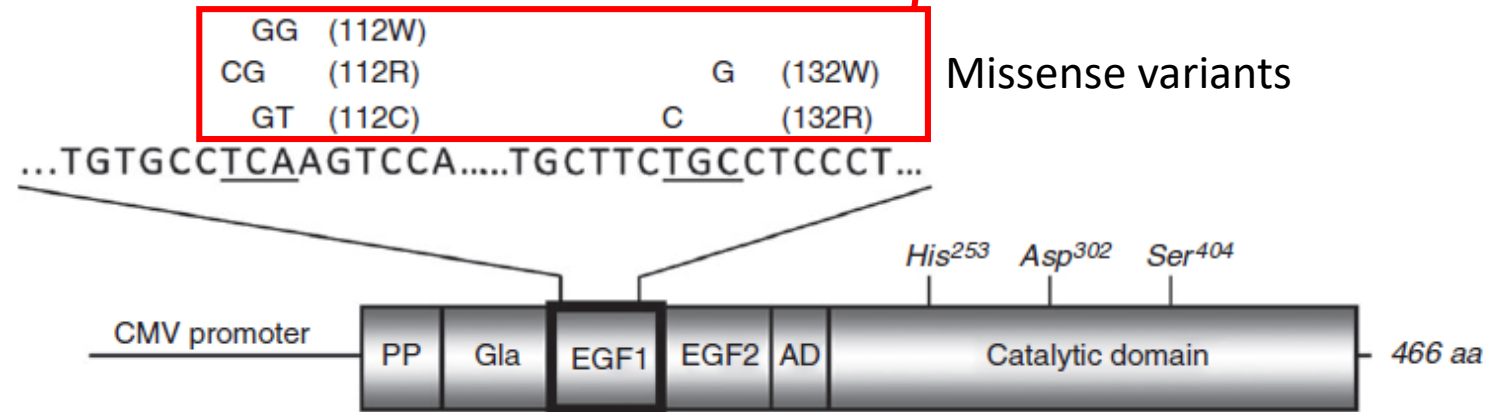
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### Methods

1. Creation of a cellular model through the transient expression of recombinant FVII **missense** variants
2. Evaluation of rFVII protein and **functional** levels by ELISA and activity assays



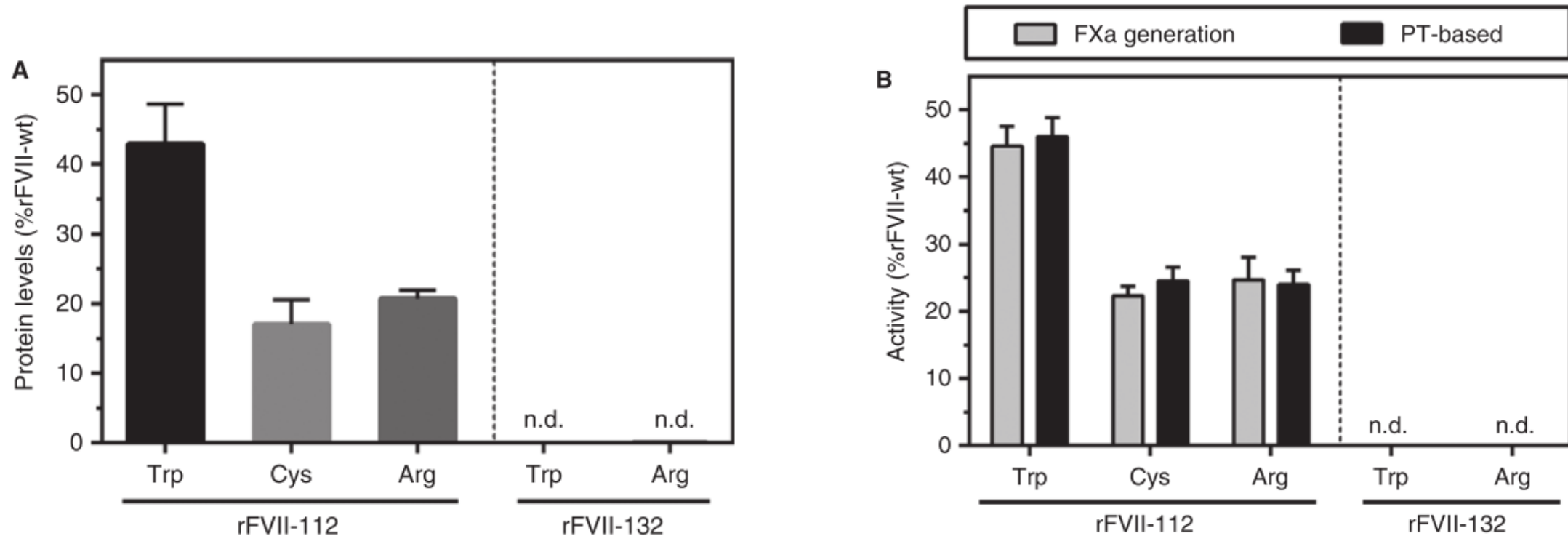
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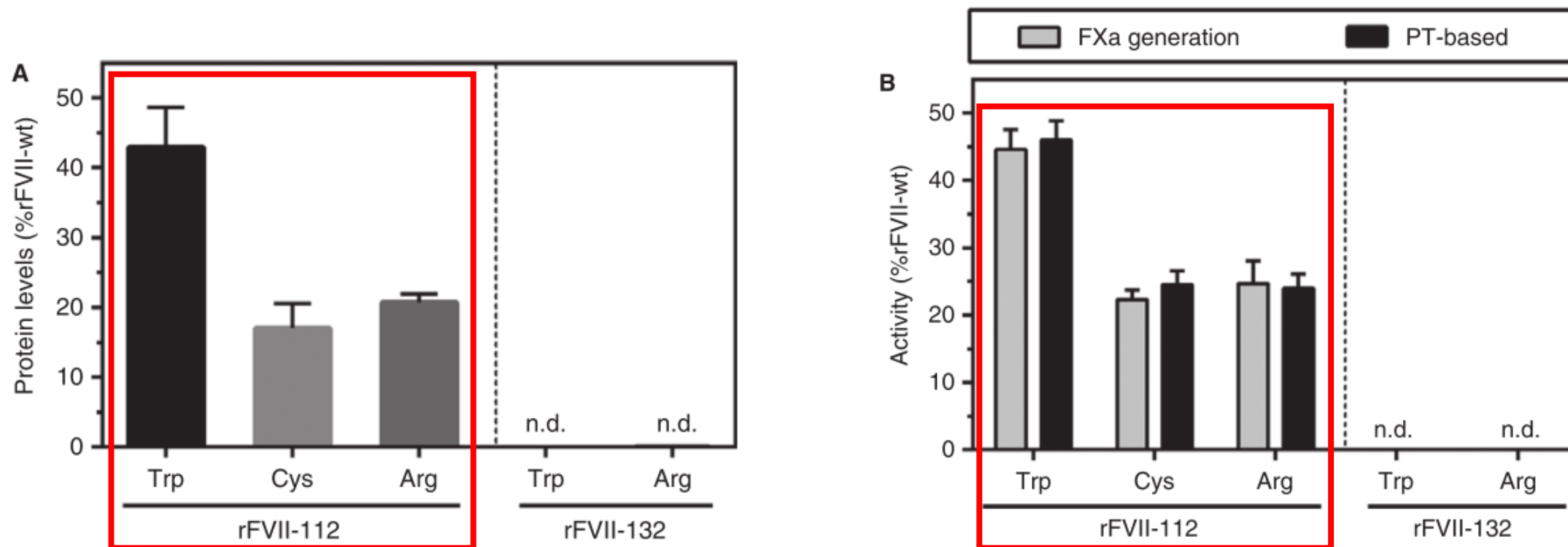
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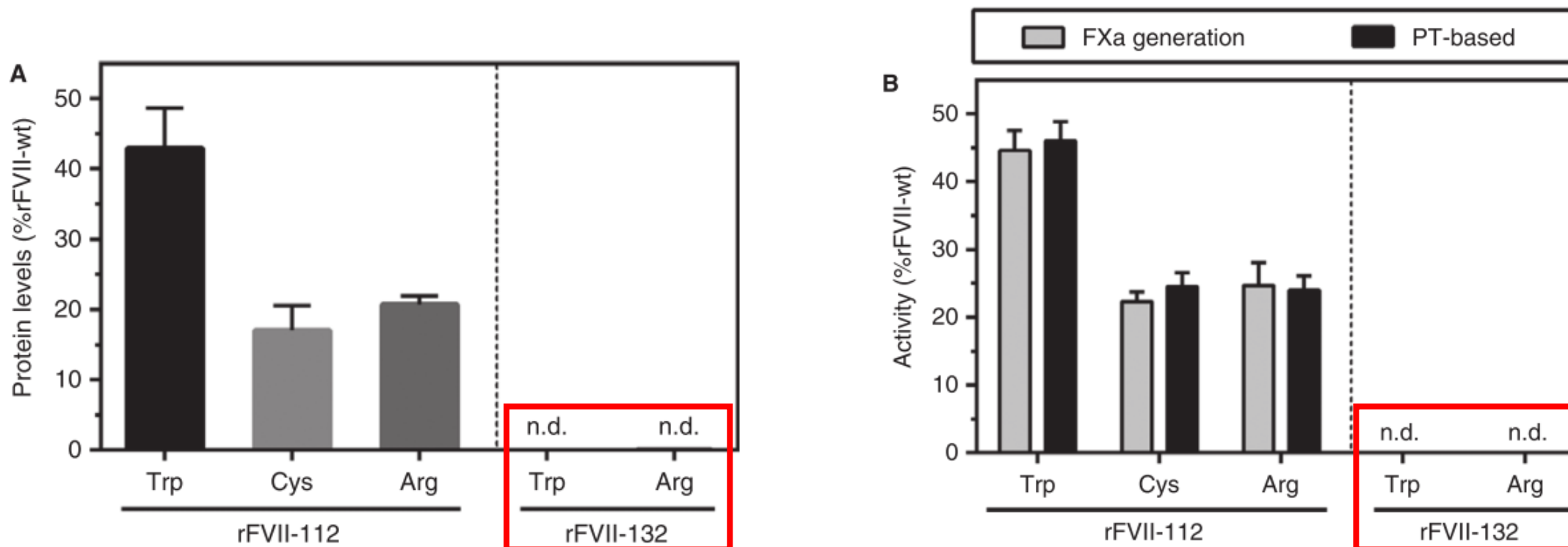
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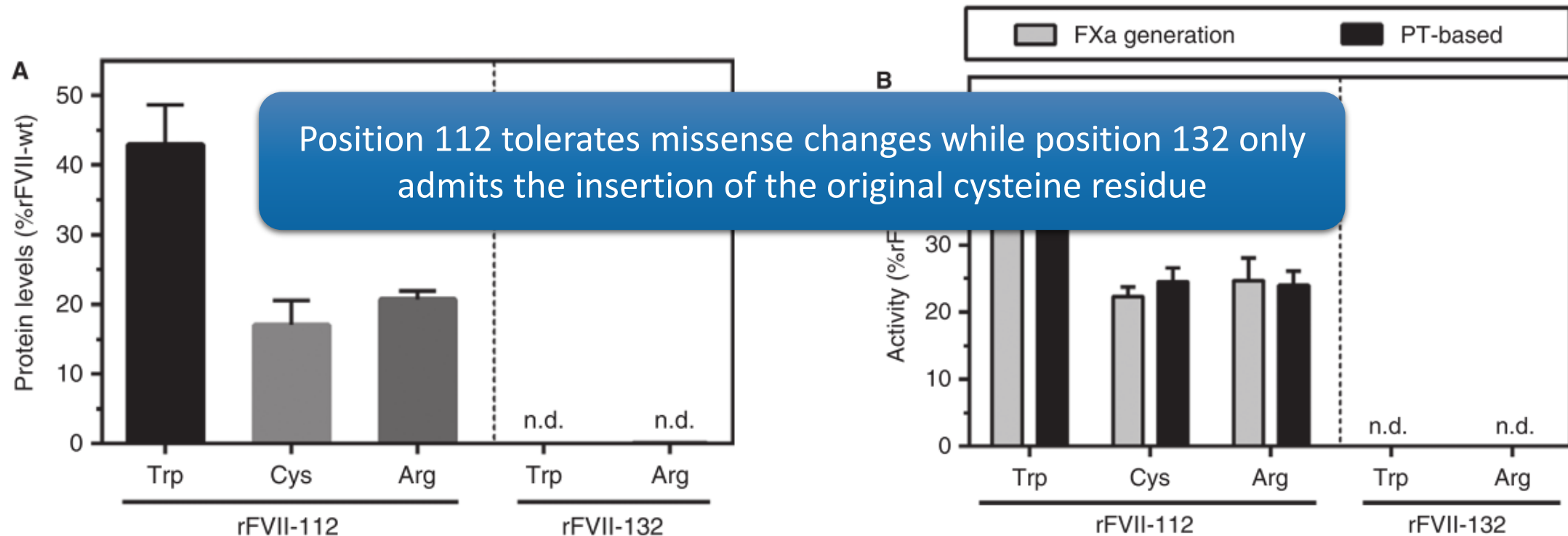
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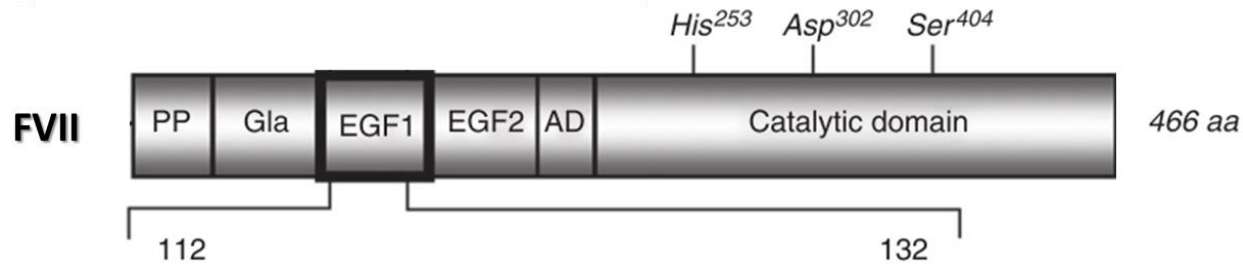
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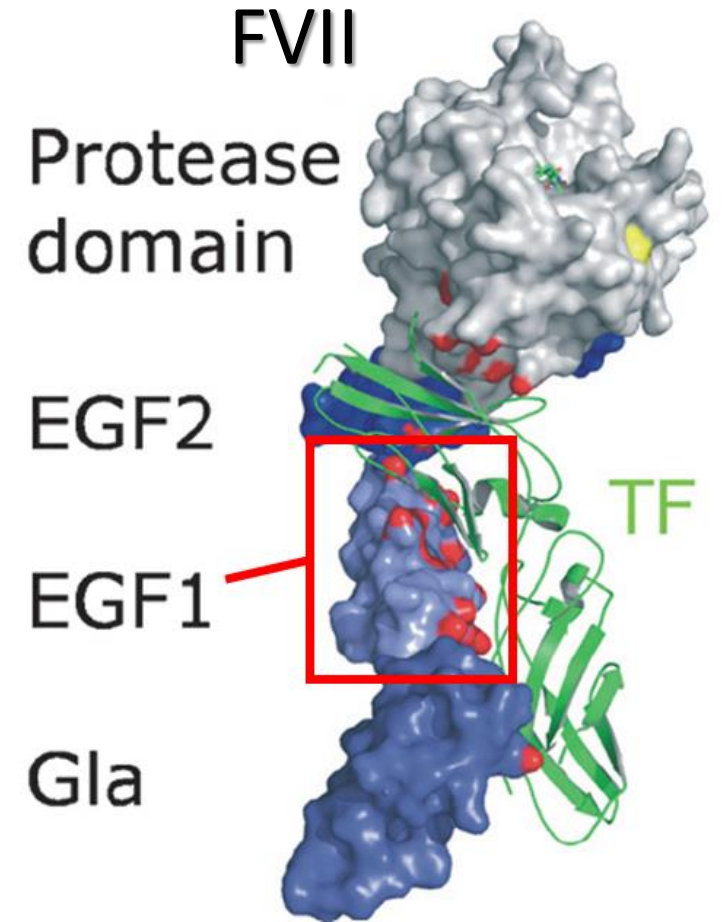
#### Secretion and activity levels of rFVII missense variants



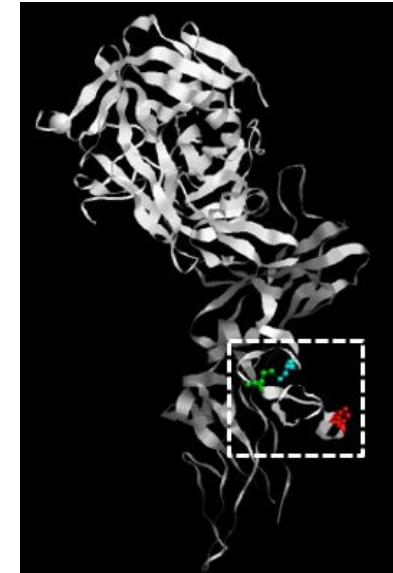
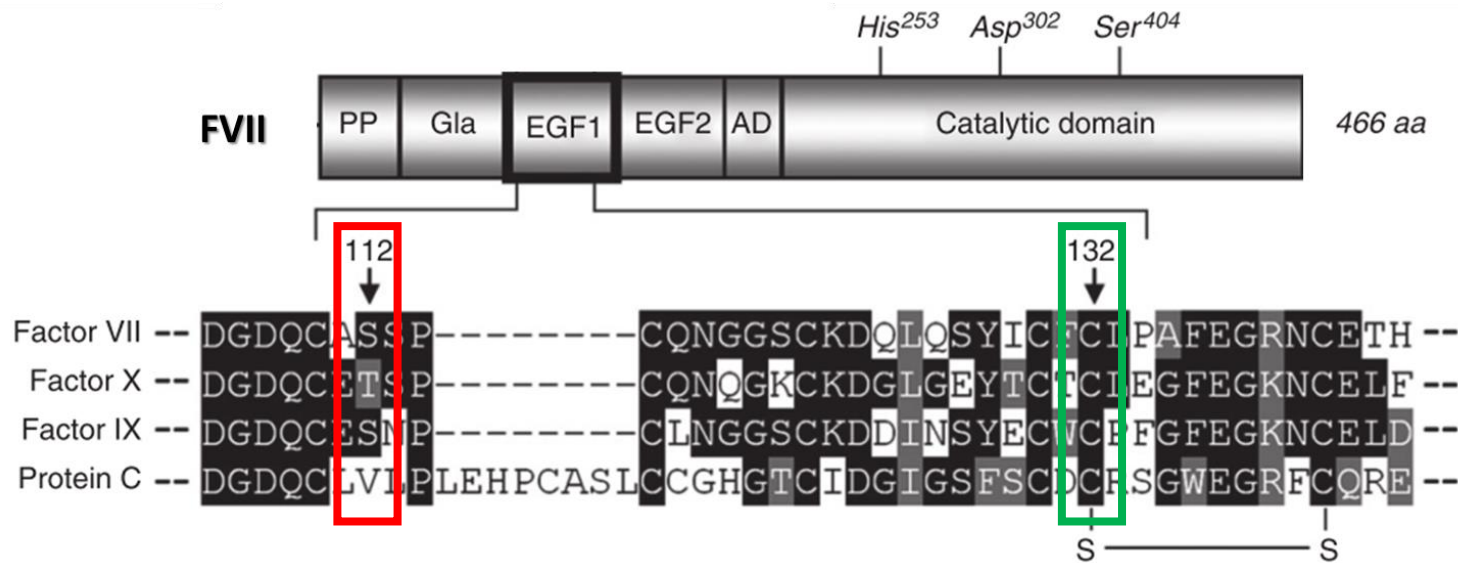
Position 112 tolerates missense changes while position 132 only admits the insertion of the original cysteine residue



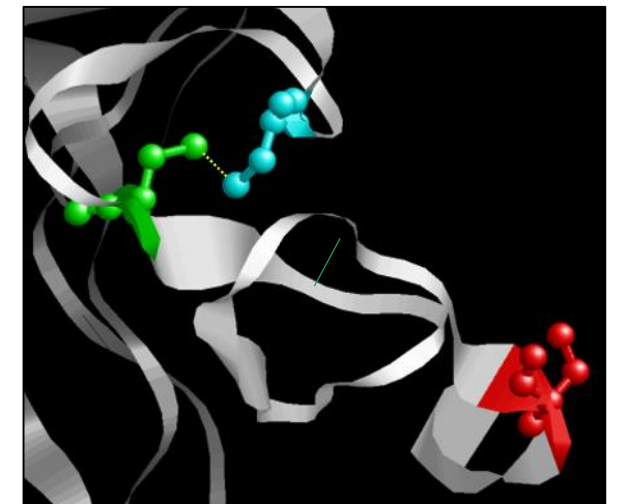
Both residues are part of the first EGF domain of FVII, which is involved in the interaction with tissue factor (TF, the FVII cofactor)



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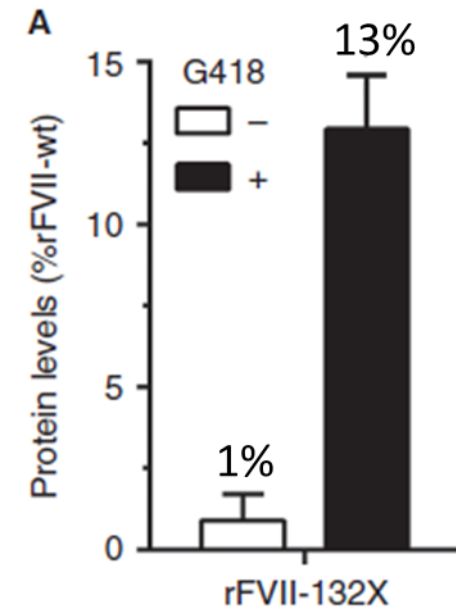
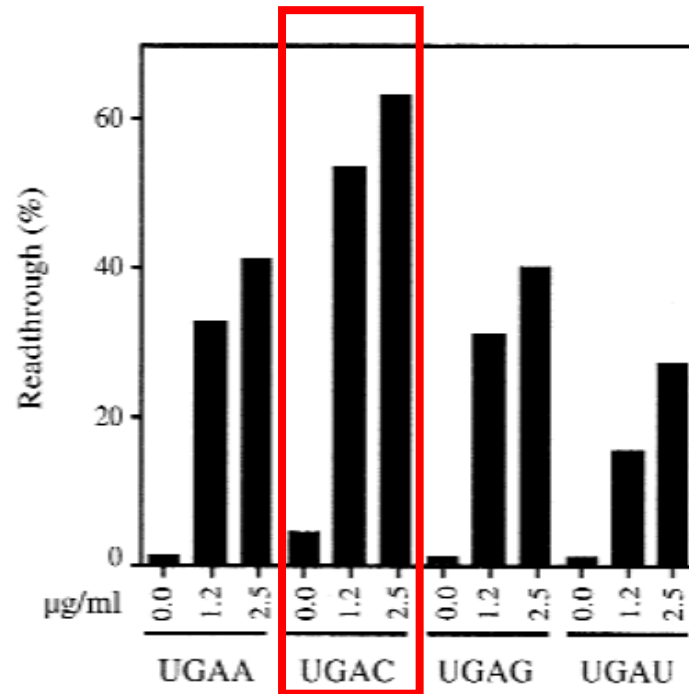


But whereas **Ser112** is surface-exposed and only partially conserved among others coagulation factors, **Cys132** forms a disulfide bridge with **Cys141** within EGF1 and is fully conserved.



## In conclusion:

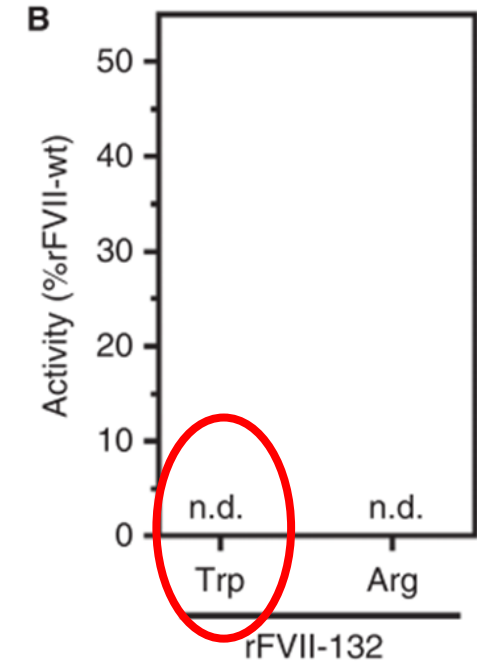
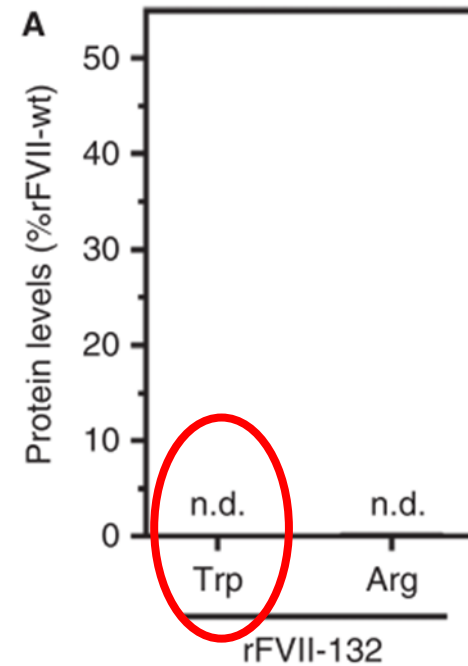
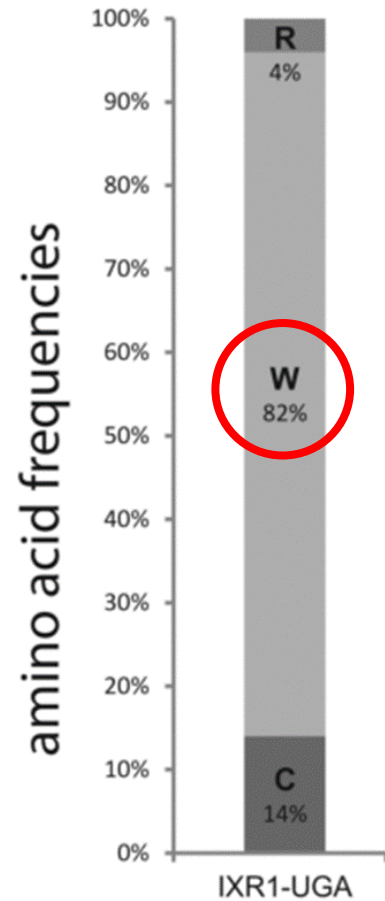
- **p.C132X** variant shows the **most readthrough favourable sequence context**



Moderate bleeding phenotype

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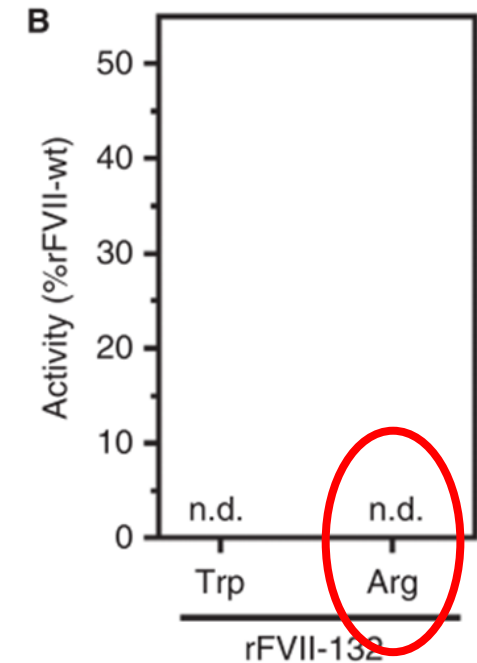
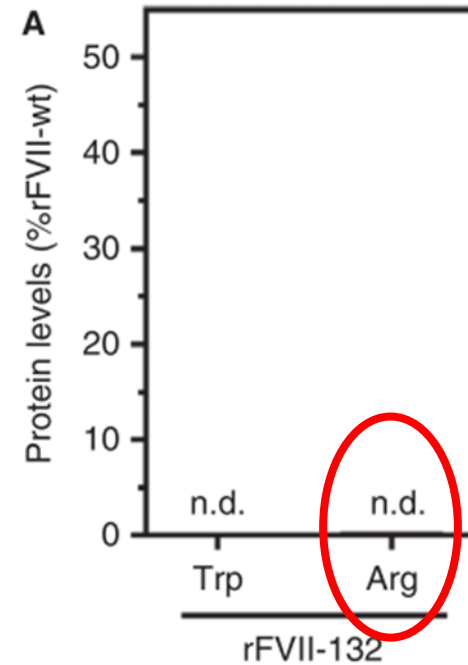
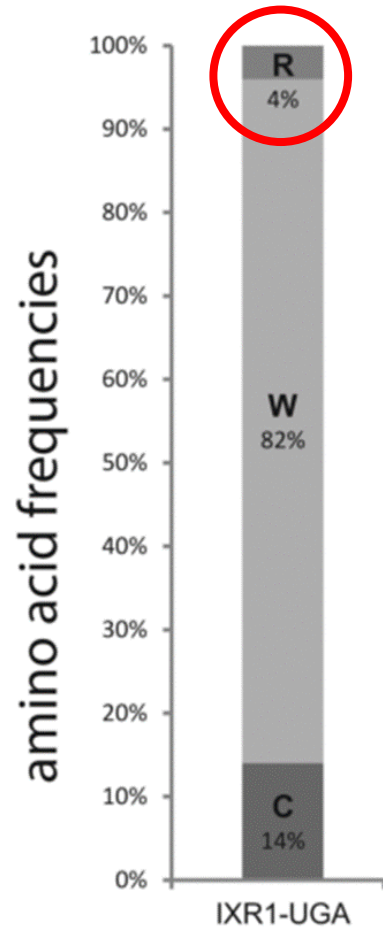
- **p.C132X** variant shows the most readthrough favourable sequence context
- PTC suppression can reinsert the original amino acid, thus leading to the production of wild-type FVII, whereas **other amino acids in this position are not tolerated** for protein secretion and function



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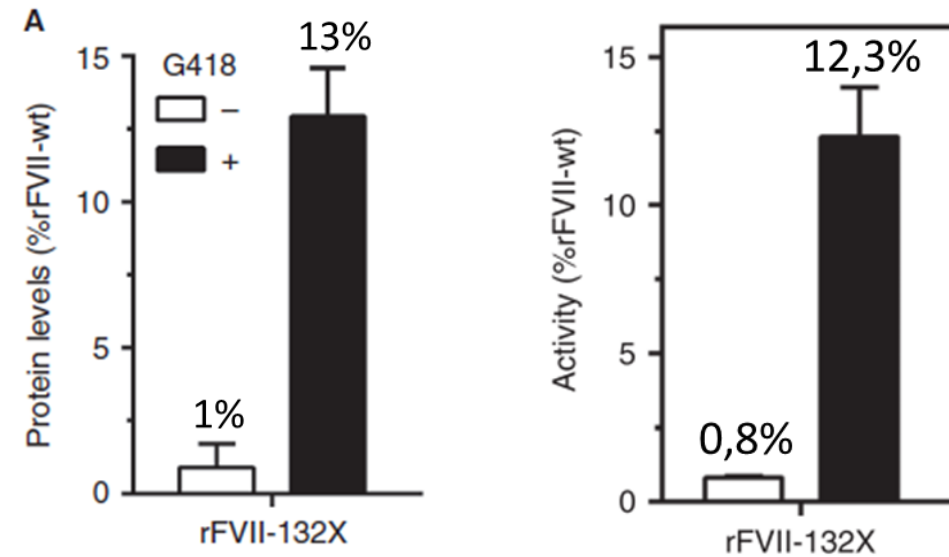
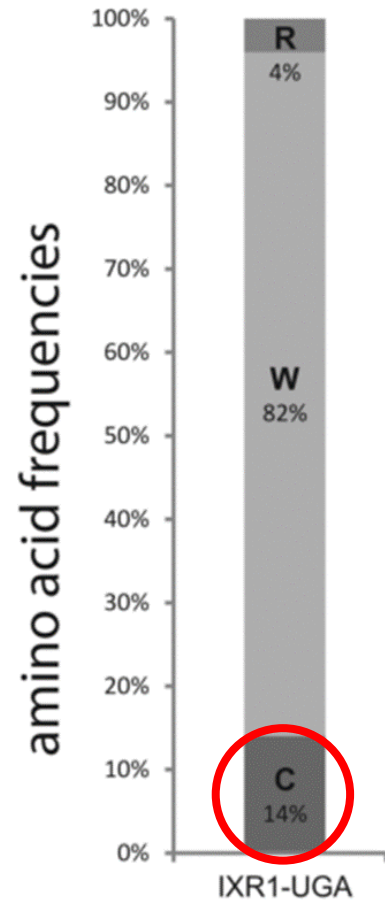


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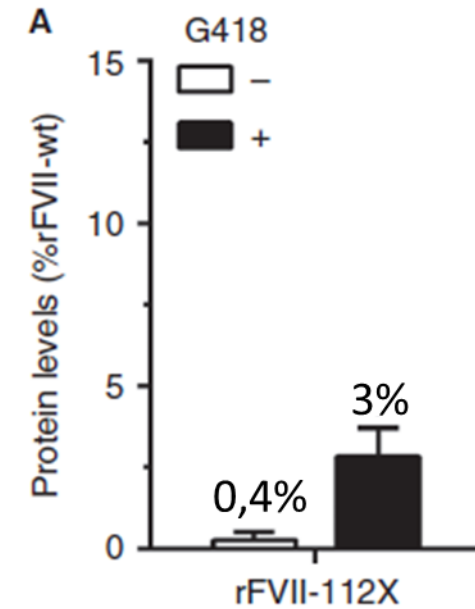
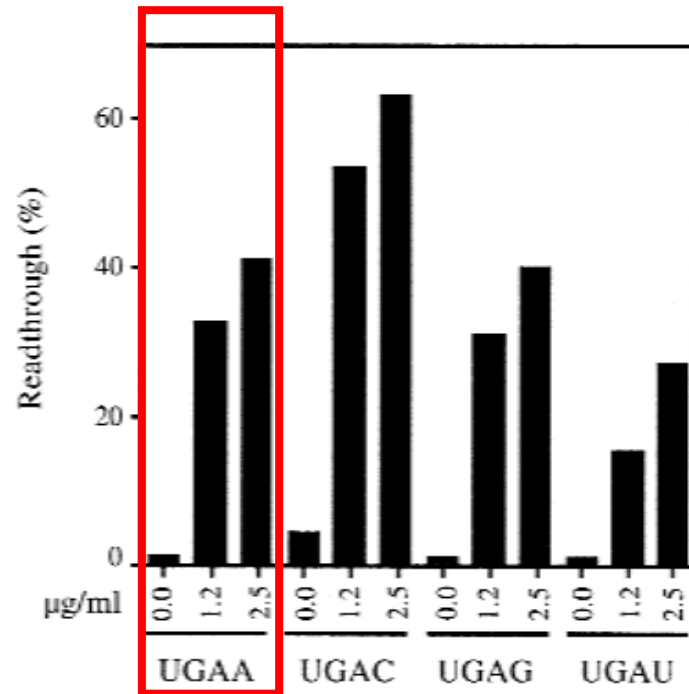


Moderate  
bleeding  
phenotype

**Insertion of original amino acid and  
production of full-length wild-type protein**

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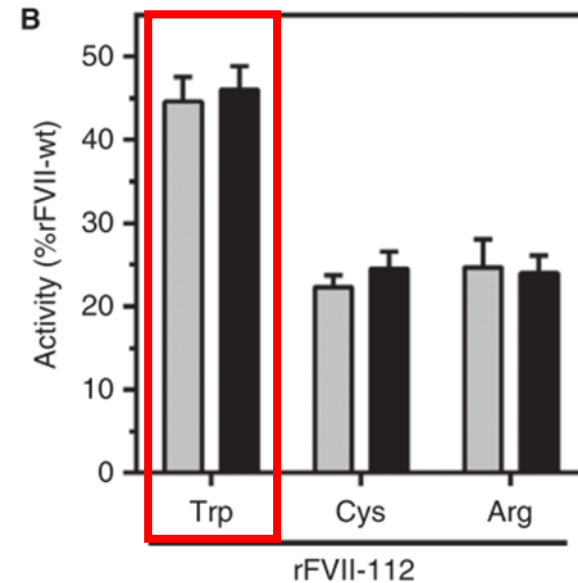
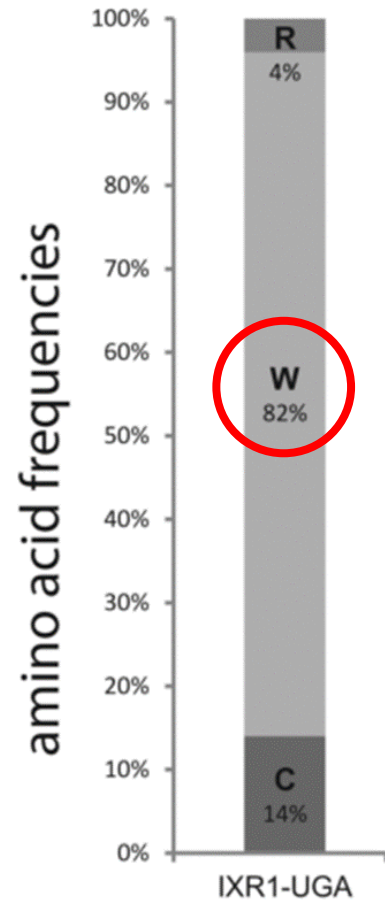
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Life-threatening bleeding phenotype

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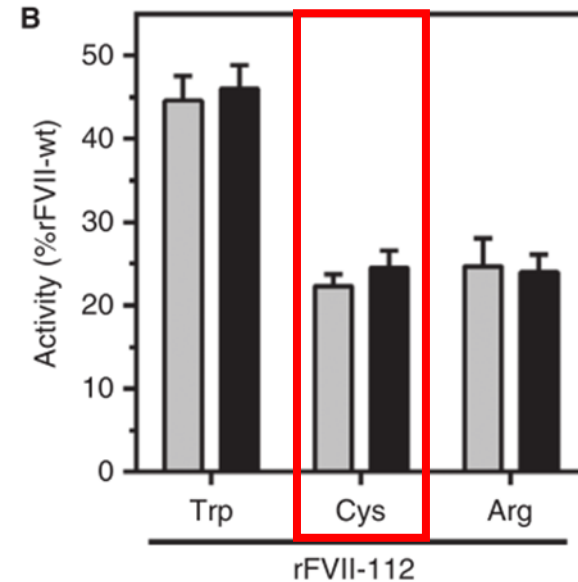
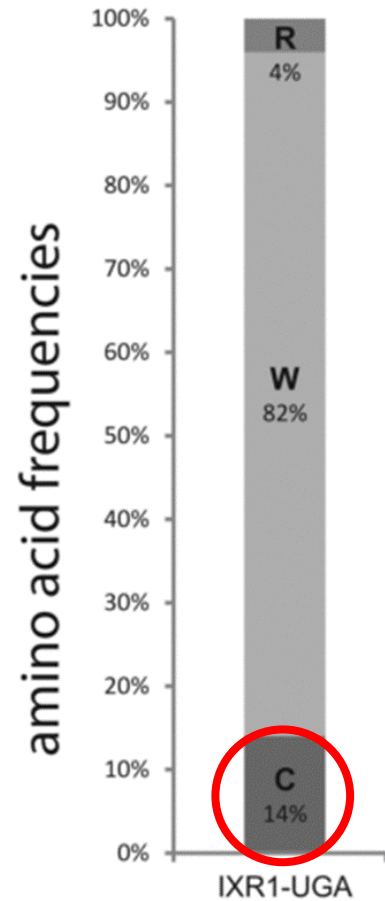
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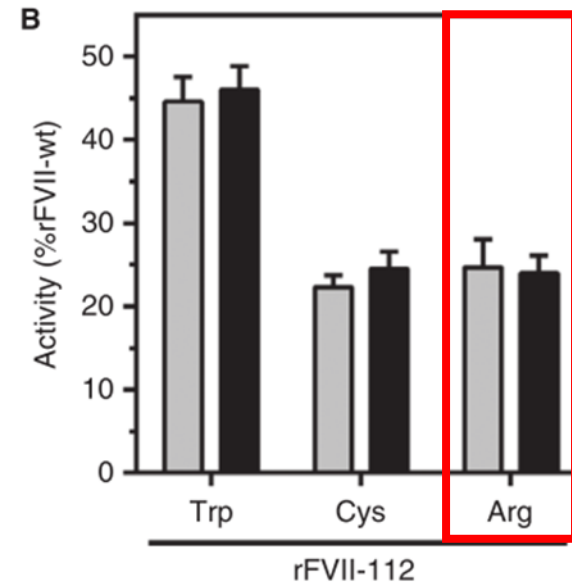
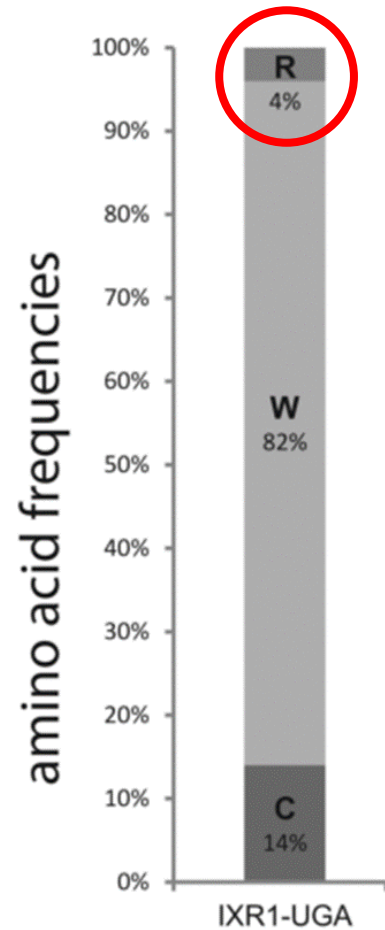
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Life-threatening bleeding phenotype

Experimental findings are consistent with the complex and integrated scenario depicted above and support the notion that many elements account for readthrough efficiency and protein function restoration



Stop codon readthrough is a process that increases the diversity of genomic information.

This mechanism is well documented with respect to viruses, yeast and *Drosophila*, but is believed to occur also in higher animals.



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Received for publication, October 28, 2011, and in revised form, March 22, 2012. Published, JBC Papers in Press, March 28, 2012, DOI 10.1074/jbc.M111.314468

Yoshihide Yamaguchi<sup>1</sup>, Akiko Hayashi<sup>2</sup>, Celia W. Campagnoni<sup>5</sup>, Akio Kimura<sup>1</sup>, Takashi Inuzuka<sup>1</sup>, and Hiroko Baba<sup>2</sup>



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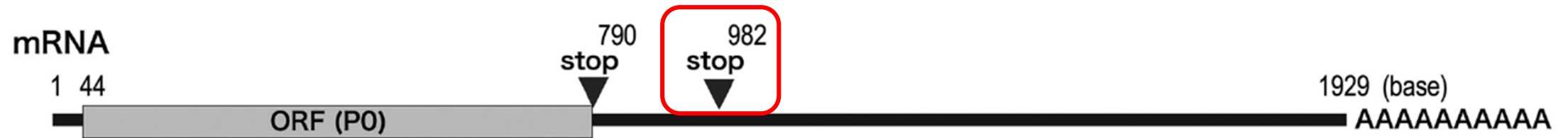
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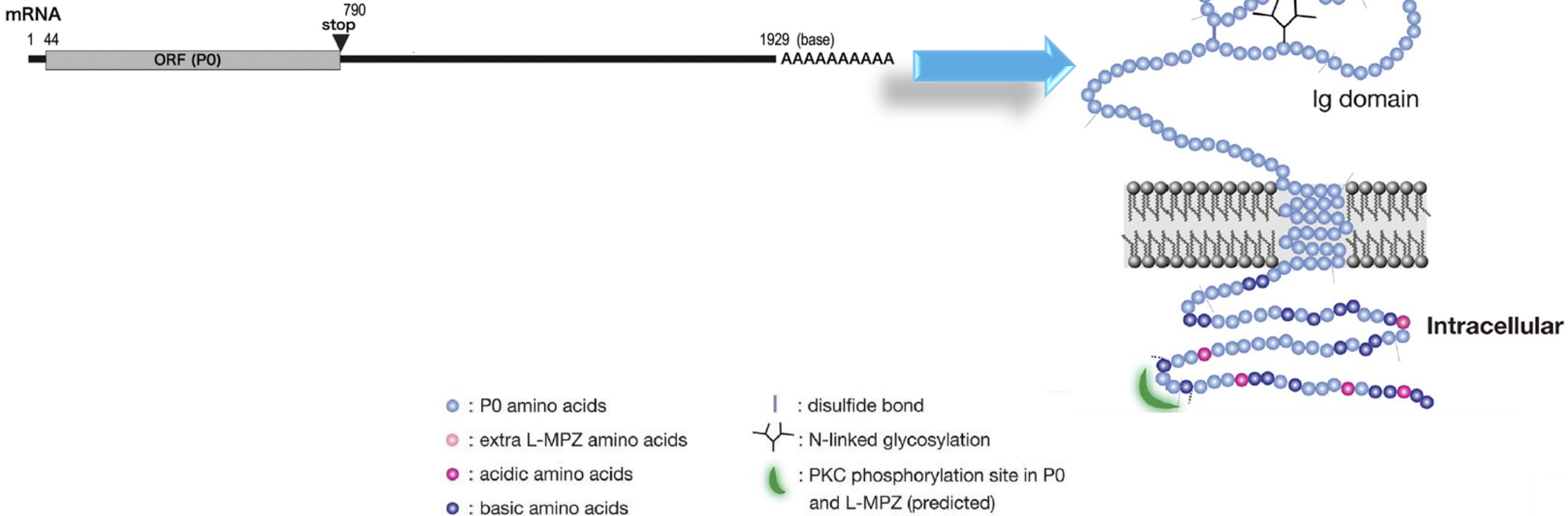
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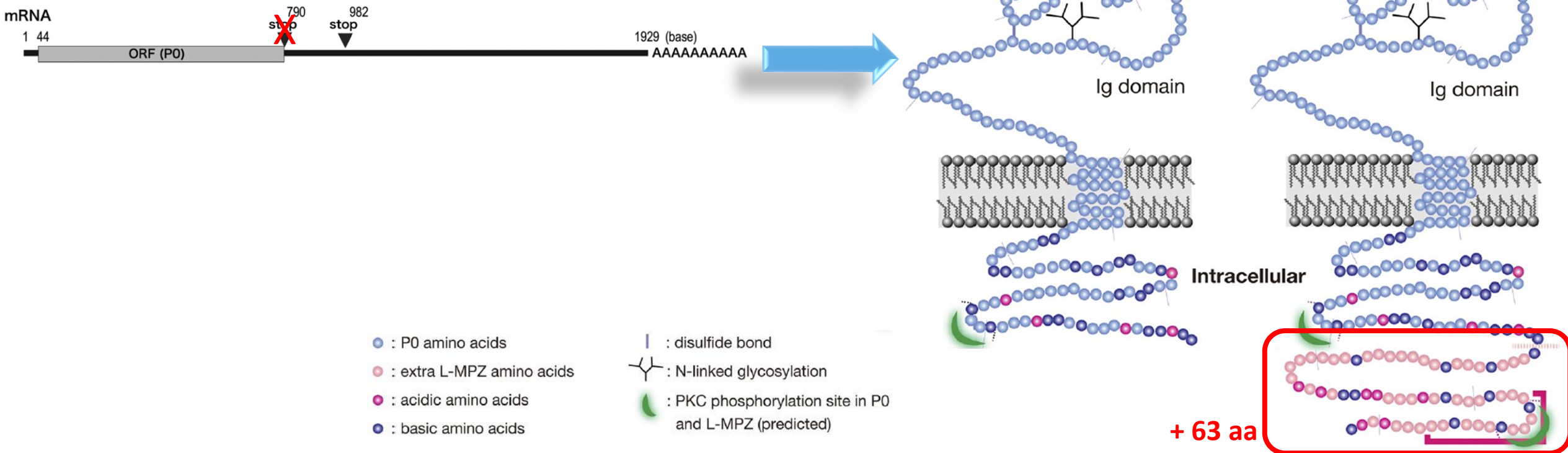
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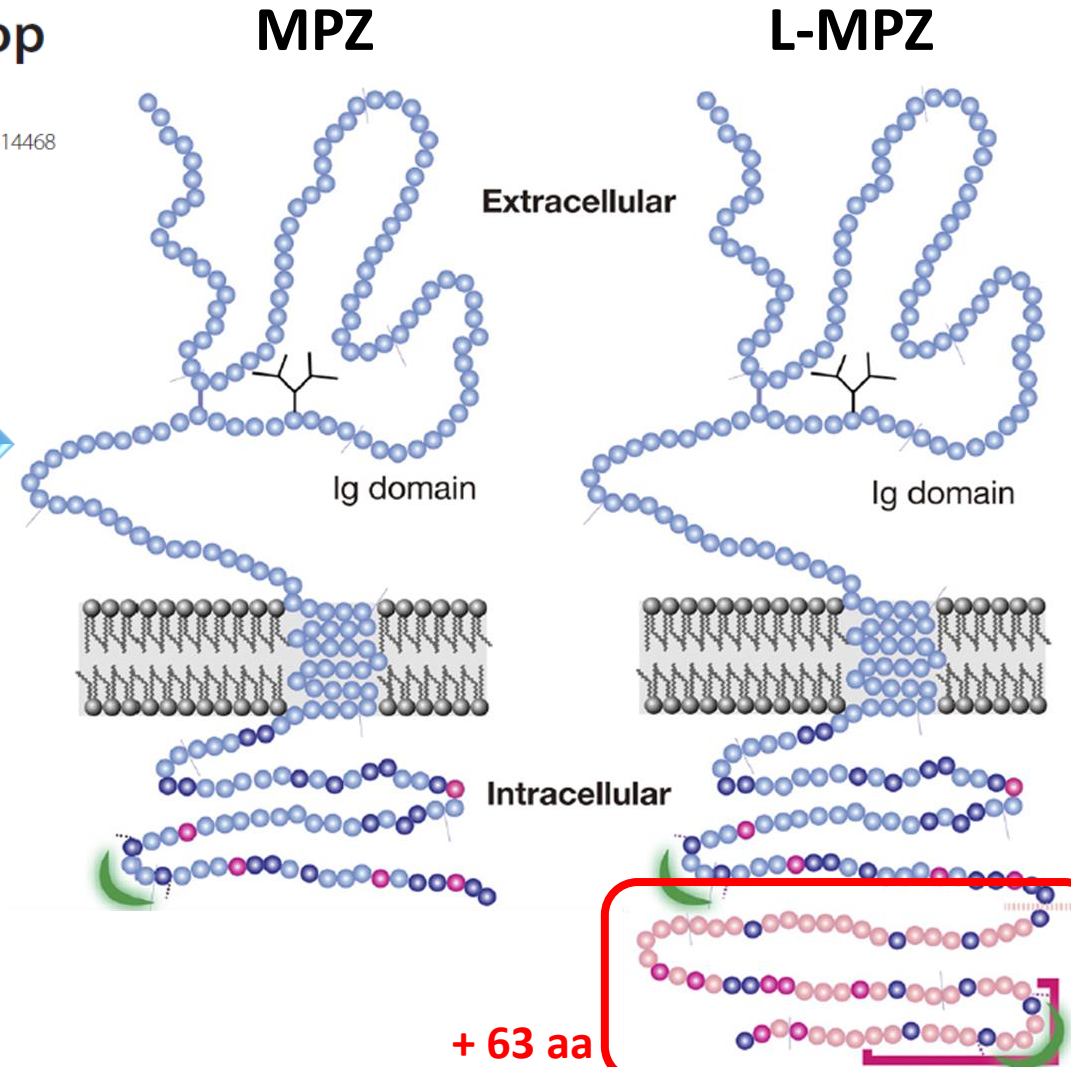
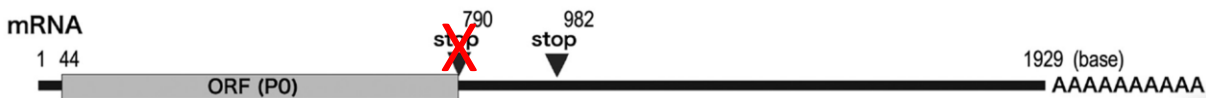
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Because the L-MPZ-specific region contains an additional putative phosphorylation site, it may have some unique roles in normal and pathological myelin conditions.